

The Mediterranean Diet decreases LDL atherogenicity in high cardiovascular risk individuals: a randomized controlled trial

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Abbreviations

ApoB: apolipoprotein B

LDL-C: low-density lipoprotein cholesterol

TMD: Traditional Mediterranean Diet

TMD-VOO: TMD intervention enriched with virgin olive oil

TMD-Nuts: TMD intervention enriched with nuts

Keywords: “LDL cytotoxicity”, “LDL oxidation”, “LDL size”, “low density lipoproteins”, “Mediterranean Diet”

1 **ABSTRACT**

2

3 **Scope.** Traditional Mediterranean Diet (TMD) protects against cardiovascular disease
4 through several mechanisms such as decreasing LDL cholesterol levels. However,
5 evidence regarding TMD effects on LDL atherogenic traits (resistance against
6 oxidation, size, composition, cytotoxicity) is scarce.

7 **Methods and results.** We assessed the effects of a 1-year intervention with a TMD on
8 LDL atherogenic traits in a random sub-sample of individuals from the PREDIMED
9 Study ($N=210$). We compared two TMDs: one enriched with virgin olive oil (TMD-VOO,
10 $N=71$) and another with nuts (TMD-Nuts, $N=68$), versus a low-fat control diet ($N=71$).
11 After the TMD-VOO intervention, LDL resistance against oxidation increased (+6.46%,
12 $P=0.007$), the degree of LDL oxidative modifications decreased (-36.3%, $P<0.05$),
13 estimated LDL particle size augmented (+3.06%, $P=0.021$), and LDL particles became
14 cholesterol-rich (+2.41% $P=0.013$) relative to the low-fat control diet. LDL lipoproteins
15 became less cytotoxic for macrophages only relative to baseline (-13.4%, $P=0.019$). No
16 significant effects of the TMD-Nuts intervention on LDL traits were observed versus the
17 control diet.

18 **Conclusion.** Adherence to a TMD, particularly when enriched with virgin olive oil,
19 decreased LDL atherogenicity in high cardiovascular risk individuals. The development
20 of less atherogenic LDLs could contribute to explaining some of the cardioprotective
21 benefits of this dietary pattern.

22 INTRODUCTION

23

24 Adherence to a Traditional Mediterranean Diet (TMD) protects against the development
25 of cardiovascular diseases as observed in a consistent body of evidence coming from
26 observational and randomized controlled trials [1]. The PREDIMED Study (*Prevención*
27 *con Dieta Mediterránea*), a multi-center, parallel, randomized controlled trial, has
28 demonstrated with a high degree of scientific evidence that a TMD has protective
29 effects on primary cardiovascular disease prevention [2, 3]. Due to its richness in
30 antioxidants and other bioactive molecules (this dietary pattern is based on the
31 consumption of virgin olive oil, nuts, fruit, vegetables, whole grains, legumes, fish,
32 poultry, and moderate quantities of wine at meals) [2], the TMD protects against
33 atherosclerosis by improving blood lipid levels, oxidative/inflammatory status, and gene
34 expression associated with the development of cardiovascular diseases [4–7]. The
35 TMD has also been shown to enhance some characteristics related to low-density
36 lipoproteins (LDLs), such as the levels of total and oxidized LDL particles [8, 9].
37 Besides these properties there are other characteristics that make LDL especially
38 atherogenic including: 1) LDL resistance against oxidative modifications; 2) LDL
39 content of triglycerides, cholesterol, and various proteins; 3) LDL cytotoxic potential on
40 different cell types; and 4) LDL ability to transfer cholesterol to hepatocytes. Our group
41 has previously studied the effects of a typical TMD food, virgin olive oil, on some of
42 these traits [10]. To date, however, the effects of the whole TMD on a complete set of
43 LDL atherogenic properties remain to be fully elucidated.
44 Thus, the aim of the present study was to assess whether a long-term consumption of
45 a TMD, enriched with virgin olive oil or nuts, could decrease the atherogenicity of LDL
46 particles in humans.

47

48

49 MATERIALS AND METHODS

50 **Study design**

51 Our study population was a random subsample of volunteers ($N=210$) from the
52 PREDIMED Study (*Prevención con Dieta Mediterránea*), a randomized, controlled,
53 large-scale, parallel, multicenter trial that assessed the long-term effects of a TMD on
54 the primary prevention of cardiovascular events in a high cardiovascular risk population
55 [2]. Participants were randomly allocated to: 1) a TMD enriched with virgin olive oil
56 (TMD-VOO, $N=71$); 2) a TMD enriched with nuts (TMD-Nuts, $N=68$); and 3) a low-fat
57 control diet following the indications of the American Heart Association ($N=71$).
58 Volunteers allocated to the TMD interventions were instructed to replace cooking fats
59 with virgin olive oil; increase their consumption of fruit, vegetables, nuts, legumes, fish,
60 and poultry; and decrease their intake of red/processed meat and processed foods.
61 Individuals in the low-fat control diet were taught to decrease their consumption of fatty
62 foods (oils, nuts, butter, meat, fish, and processed foods) and to promote their intake of
63 vegetal foods. In addition, TMD-VOO volunteers received 1 L/week of virgin olive oil,
64 and TMD-Nuts individuals were given 210 g/week of mixed nuts (walnuts, hazelnuts,
65 and almonds) to particularly promote the intake of these food items. A more detailed
66 description of the three dietary interventions is available elsewhere [2]. We studied the
67 effects of a TMD on the characteristics related to the atherogenicity of LDL particles
68 before and after one year of intervention. Local institutional ethic committees approved
69 the protocol of the study, and all volunteers provided a signed informed consent before
70 entering the trial. Further details of the study have been previously published [2]. The
71 PREDIMED Study protocol was registered with the International Standard Randomized
72 Controlled Trial Number ISRCTN35739639 (www.controlled-trials.com).

73

74 **Biological samples and clinical information**

75 K3-EDTA plasma samples were obtained from blood collected from the participants
76 before and post-intervention. The samples did not suffer any thaw-freeze cycles before
77 our experiments. We isolated LDL particles from a plasma aliquot by means of a

78 density gradient ultracentrifugation [10]. Samples were stored at -80°C until required.
79 We also gathered the following information before and after the intervention: 1) the
80 general clinical status of the volunteers (sex, age, body mass index, waist
81 circumference, blood pressure); 2) their adherence to the TMD and their usual diet in
82 the previous year (by a food frequency questionnaire); and 3) their levels of physical
83 activity (through a validated Minnesota Leisure-Time Physical Activity questionnaire)
84 [2].

85

86 **Biochemical profile**

87 We performed all systemic determinations in plasma samples in an ABX Pentra-400
88 autoanalyzer (Horiba-ABX, Montpellier, France). We determined the levels of fasting
89 glucose, triglycerides, and total cholesterol by enzymatic methods (ABX Pentra
90 Glucose HK CP, ABX Pentra Triglycerides CP, and ABX Pentra Cholesterol CP, all
91 from Horiba-ABX), the levels of HDL cholesterol by the Accelerator Selective Detergent
92 method (ABX Pentra HDL Direct CP, Horiba-ABX), and the levels of apolipoproteins B
93 (ApoB) (ABX Pentra ApoB, Horiba-ABX) and A-I (ApoA-I) (ABX Pentra ApoA1, Horiba-
94 ABX) by immunoturbidimetry. The inter-assay coefficients of variation (CVs) of these
95 determinations were: 1.91% for fasting glucose, 4.07% for triglycerides, 1.24% for total
96 cholesterol, 1.79% for HDL cholesterol, 1.59% for ApoB, and 1.68% for ApoA-I. We
97 also calculated LDL cholesterol (LDL-C) levels (according to the Friedewald formula
98 whenever triglycerides were <300 mg/dL) and the ApoB/ApoA-I ratio.

99

100 **LDL resistance against oxidation**

101 We incubated isolated LDL particles with an oxidizing agent (CuSO₄) to assess their
102 resistance to accumulate Cu²⁺-induced conjugated dienes. We dialyzed LDL
103 lipoproteins against PBS and incubated them (final concentration: 10 mg
104 cholesterol/dL) with CuSO₄ (final concentration: 5 μM) in 96-well transparent plates at
105 37°C in an Infinite M200 reader (Tecan Ltd, Männedorf, Switzerland). We measured

106 absorbance at 234 nm every 3 minutes for 4 hours to obtain the curves of LDL
107 oxidation. From the curves, we calculated the lag time (the time when maximal
108 oxidation started, in minutes). High LDL lag time values are associated with a greater
109 resistance of LDL particles against oxidation [10]. The inter-assay CV was 12.4%.

110

111 **Degree of LDL oxidative modifications**

112 We measured the quantity of oxidative modifications in LDL particles (malondialdehyde
113 equivalents) by the thiobarbituric reactive acid species technique in isolated LDL
114 samples [11]. We then divided the malondialdehyde equivalents by the cholesterol
115 content in each LDL sample (see “LDL composition”). The inter-assay CV was 9.21%.

116

117 **Estimated LDL particle size**

118 From the data of the volunteers’ plasma lipid profile we calculated a surrogate marker
119 for LDL size, the LDL-C/ApoB ratio (unitless). Low ratio values are associated with LDL
120 particles of smaller size [12].

121

122 **LDL composition**

123 We analyzed the composition of isolated LDL lipoproteins in an ABX Pentra-400
124 autoanalyzer (Horiba-ABX). We measured the levels of triglycerides (ABX Pentra
125 Triglycerides CP, Horiba-ABX) and cholesterol (Cholesterol-LQ, Spinreact) by
126 enzymatic methods, total protein (ABX Pentra Total Protein CP, Horiba-ABX) by the
127 Biuret reaction, and ApoB (ABX Pentra ApoB, Horiba-ABX) by immunoturbidimetry.
128 The inter-assay CVs of these measurements were: 4.62% for triglycerides, 3.86% for
129 cholesterol, 2.47% for total protein, and 1.59% for ApoB.

130 From these values, we determined the content of cholesterol and triglycerides in
131 isolated LDL particles (adjusted for the ApoB quantity of the lipoproteins), the
132 triglyceride/cholesterol ratio, and the percentage of LDL proteins other than ApoB, as

133 follows: (total protein in LDL – ApoB in LDL)/total protein in LDL x100.

134

135 **LDL cytotoxicity in macrophages**

136 We grew human THP-1 in RPMI-1640 medium (complemented with 10% fetal bovine
137 serum, 1% penicillin-streptomycin, 1% L-glutamine, and 1% sodium pyruvate),

138 refreshed them every 72h, and differentiated them into macrophages (by incubating
139 them with 200 nM phorbol-myristate-acetate –Sigma, Barcelona, Spain–, for 96h).

140 Next, we washed the macrophages and incubated them with isolated LDL particles

141 (concentration: 10 mg/dL cholesterol in LDL particles [10, 13]) or without LDL (as

142 negative control), for 16h. After incubation, we washed the cells and incubated them

143 with 0.5 mg/mL soluble MTT bromide (Thiazolyl Blue Tetrazolium bromide, Sigma),

144 during 4h. Then, we removed the supernatant, washed the cells again, and dissolved

145 the cell content (and the MTT-formazan crystals inside the cells) with DMSO (Sigma),

146 for 1h under stirring. Finally, we measured absorbance at 570 nm in an Infinite M200

147 reader (Tecan Ltd). If the viability of the cells was high, they would transform the

148 soluble MTT pigment more rapidly into insoluble MTT-formazan crystals, and the

149 absorbance of the DMSO-dissolved cell content would be greater. Therefore, high LDL

150 cytotoxicity would be related to low MTT-absorbance values [13].

151 To calculate the index of LDL cytotoxicity in macrophages, we subtracted the blank

152 (absorbance of the cells non-treated with MTT) from all absorbance values, and

153 calculated the difference in the MTT-absorbance in the LDL-treated cells versus the

154 untreated cells (the negative control): (MTT-absorbance in LDL-treated cells – MTT-

155 absorbance in untreated cells)/MTT-absorbance in untreated cells*100. The inter-assay

156 CV of the experiment ($N=7$) was 35.5%.

157

158 **Data quality control**

159 LDL oxidation and cytotoxicity experiments followed a predefined process to control

160 inter-assay variability. In all these experiments, we analyzed the samples from the

161 same volunteers in the same analytical run, in duplicate, and we did not allow intra-
162 repetition CVs over 15%. We also included an LDL pool (isolated from a pool of plasma
163 from 20 healthy volunteers) in each experiment. We finally divided the values obtained
164 in the samples by the value of the pool for each parameter, to obtain normalized ratios
165 without units.

166

167 **Sample size calculation**

168 A sample size of 68 participants per group allowed $\geq 80\%$ power to detect a significant
169 difference of 0.05 points in LDL lag time values (expressed as normalized units)
170 between pre- and post-intervention values, and of 0.07 points among the three
171 interventions, considering a 2-sided type I error of 0.05, a loss rate of 1%, and the
172 standard deviation of the differences in normalized LDL lag time values (SD=0.144)
173 after an analogous dietary intervention [10].

174

175 **Statistical analyses**

176 We examined the distribution of continuous variables in normality plots and the
177 Shapiro-Wilk test, and log-transformed the non-normally distributed variables. To find
178 possible differences in the baseline characteristics of our subsample and the whole
179 PREDIMED population, we performed a T-test. To investigate possible differences in
180 baseline values among the three intervention groups, we carried out a chi-square test
181 for categorical variables and a one-way ANOVA for continuous variables.

182 We studied the differences between pre- and post-intervention values after the three
183 interventions in a paired T-test. We also analyzed the effects of the TMD interventions
184 (relative to the low-fat diet) on the changes in the variables of interest in a multivariate
185 regression analysis (using two dummy variables, one for each intervention group)
186 adjusted for: sex; age; center of origin of the volunteer ($k-1$ dummy variables); baseline
187 value of the variable; and changes in the presence of dyslipidemia, diabetes,
188 hypertension, and tobacco habit ($k-1$ dummy variables) throughout the study.

189 To detect potential relationships among LDL atherogenic traits, we carried out
190 Spearman's correlation analyses among the baseline values of these determinations.
191 In addition, to assess the relationships among the changes in LDL atherogenic
192 characteristics after the TMD-VOO intervention (the one after which most of the
193 differences occurred), we carried out Spearman's correlation analyses and a principal
194 component analysis among these variables.
195 We accepted any two-sided P -value <0.05 as significant. We performed all the
196 previous analyses in R Software, version 3.0.2 (*R: A language and environment for*
197 *statistical computing. R Foundation for Statistical Computing, Vienna, Austria*).

198

199

200 **RESULTS**

201

202 **Participants and dietary adherence**

203 Study design is available in **Supplemental Figure 1**. No differences in the baseline
204 characteristics were found among the three groups in our subsample (**Table 1**). With
205 respect to the whole PREDIMED Study population, our volunteers were on average 1.6
206 years younger, with 9.2% more males, and 6.9% more dyslipidemic individuals at
207 baseline ($P<0.05$) (**Supplemental Table 1**). We found no differences in energy
208 expenditure in leisure-time physical activity among interventions.

209 Subjects appeared to be relatively compliant to the diets. The augmented TMD
210 adherence after the TMD-VOO intervention was observed as: 1) increments in the
211 consumption of virgin olive oil, legumes, and fish; and 2) decreases in the intake of red
212 and processed meat, refined olive oil, and precooked meals ($P<0.05$). The augmented
213 TMD adherence after the TMD-Nuts intervention was due to: 1) increases in the intake
214 of nuts, virgin olive oil (less than in the TMD-VOO intervention), and canned and oily
215 fish; and 2) decrements in the consumption of meat, refined olive oil, precooked meals,
216 and industrial confectionery ($P<0.05$). Finally, adherence to the low-fat diet was

217 reflected as a reduction in the intake of saturated fats, due to decreases in the
218 consumption of whole-fat dairy products, meat (particularly red and processed),
219 processed meals, and industrial confectionery ($P<0.05$). Total, monounsaturated, and
220 polyunsaturated fat consumption was significantly reduced in the low-fat diet relative to
221 both TMD interventions (**Supplemental Tables 2-3**).

222

223 **Biochemical profile**

224 We observed a 10.9 mg/dL decrease in the levels of total cholesterol after the low-fat
225 diet ($P=0.023$ and $P=0.007$, relative to baseline values and the TMD-VOO
226 intervention). The reduction took place essentially through a 10.5 mg/dL decline in
227 LDL-C levels ($P=0.007$ and $P=0.003$, when compared to baseline and the TMD-VOO
228 intervention, respectively) (**Figure 1A-1B**). Despite the changes in LDL-C levels, ApoB
229 concentrations (**Figure 1C-1D**) and the ratio between ApoB and apolipoprotein A-I
230 levels remained unchanged after the three interventions. Finally, there was a significant
231 decrease in remnant cholesterol (-15.1%, **Figure 1E-1F**) and triglyceride
232 concentrations (-2.98%) after the TMD-Nuts intervention when compared to the low-fat
233 diet ($P=0.020$ and $P=0.021$, respectively) (**Supplemental Tables 4-5**).

234

235 **Estimated LDL particle size**

236 The LDL-C/ApoB ratio in plasma diminished after the low-fat diet relative to baseline (-
237 4.47%, $P<0.001$). In concordance, we observed a significant increase (+3.06%) in
238 estimated LDL particle size after the TMD-VOO intervention relative to the low-fat diet
239 ($P=0.021$) (**Figure 1G-1H**).

240

241 **LDL oxidation-related parameters**

242 LDL resistance against oxidation (LDL lag time) increased after both TMD
243 interventions. After the TMD-VOO intervention, LDL lag time increased relative to
244 baseline (+6.77%) and the low-fat diet (+6.46%) ($P<0.001$ and $P=0.007$, respectively).

245 After the TMD-Nuts intervention, it increased significantly only relative to baseline
246 (+6.45%) ($P=0.002$) (**Figure 2A-2B**).

247 Degree of LDL oxidative modifications (malondialdehyde equivalents in LDL, adjusted
248 for the content of cholesterol in LDL particles) decreased significantly after the TMD-
249 VOO intervention when compared with the low-fat diet (-36.3%) ($P<0.05$) (**Figure 2C-**
250 **2D**).

251

252 **LDL composition**

253 Cholesterol content in LDL particles increased after the TMD-VOO intervention relative
254 to the low-fat diet (+2.41%) ($P=0.013$) (**Figure 3A-3B**).

255 Triglyceride content in LDL lipoproteins and the ratio between triglycerides and
256 cholesterol in isolated LDL particles (data not shown) did not vary significantly after any
257 intervention.

258 Finally, the content of LDL proteins other than ApoB decreased relative to baseline
259 after the three dietary interventions (-5.06% $-P=0.001-$, -4.99% $-P=0.006-$, and -
260 3.99% $-P=0.020-$ for the TMD-VOO, the TMD-Nuts, and the low-fat diet, respectively)
261 (**Figure 3C-3D**). We found no statistically significant differences among the three
262 interventions.

263

264 **LDL cytotoxicity**

265 After the TMD-VOO intervention, the cytotoxicity of LDL particles in human
266 macrophages lessened relative to baseline (-13.4%, $P=0.019$) (**Figure 4A-4B**). We
267 found no effects after the TMD-Nuts intervention.

268

269 **Relationships among LDL atherogenic traits**

270 LDL atherogenic characteristics that reflect limited atherogenic properties (high lag time
271 values, low levels of oxidative modifications, high average estimated LDL particle size,
272 low triglyceride load, and high cholesterol content) were all inter-correlated ($P<0.05$ in

273 all cases except the relationship between LDL lag time and the ratio between
274 triglycerides and cholesterol in LDL). Low LDL cytotoxicity in macrophages was also
275 associated with a low degree of LDL oxidative modifications, and triglyceride-poor,
276 protein-poor, cholesterol-rich LDL particles (all $P < 0.001$), and with increases in
277 estimated LDL particle size ($P = 0.056$) (**Supplemental Table 6**).

278 Changes in LDL atherogenic traits after the TMD-VOO intervention also correlated
279 amongst each other (**Supplemental Table 7**). First, decreases in LDL oxidation after
280 this intervention were associated with increases in triglyceride-poor, protein-poor,
281 cholesterol-rich LDL particles, and low LDL cytotoxicity in macrophages (all $P < 0.001$).

282 Second, increases in cholesterol content and decreases in the relative levels of
283 triglycerides in LDL particles were linked to decreases in LDL cytotoxicity ($P = 0.009$ and
284 $P = 0.090$, respectively). Finally, as observed in the principal component analysis
285 (**Supplemental Figure 2**): 1) changes in LDL lag time and estimated LDL particle size
286 were inter-related; 2) changes in the degree of LDL oxidative modifications, the
287 triglyceride/cholesterol ratio in LDL particles, and the percentage of LDL proteins other
288 than ApoB were associated, and probably inter-related to changes in LDL cytotoxicity;
289 and 3) all these effects were independent from the changes in LDL-C and ApoB levels.

290

291 Values of the comparisons between post- and pre-intervention values, and between the
292 changes in the TMD interventions relative to the low-fat diet for LDL atherogenic traits,
293 are available in **Supplemental Tables 4** and **5**, respectively.

294

295

296 **DISCUSSION**

297 Our results indicate that a 1-year intervention with a TMD improves several LDL traits
298 related to its atherogenicity (resistance against oxidation, size, composition, and
299 cytotoxicity) in high cardiovascular risk individuals, particularly when the TMD is
300 enriched with virgin olive oil. To the best of our knowledge, this is the first time that the

301 effect of a healthy dietary pattern on a complete set of LDL atherogenic properties has
302 been studied in humans.

303 LDL oxidation is one of the most relevant biochemical events that leads to the
304 formation of an atherosclerotic plaque [14]. Oxidized LDL particles are avidly
305 phagocytized by macrophages which results in their transformation to foam cells [15],
306 and they also induce cytotoxic responses in endothelial cells [16]. Although the causal
307 relationship between LDL oxidation and atherosclerosis is still a controversial topic [17],
308 increased oxidized LDL levels and high susceptibility of LDL lipoproteins to oxidation
309 have been associated with greater cardiovascular risk in some clinical trials [18, 19],
310 but not in an independent manner in others [20]. In our trial, the TMD (especially when
311 enriched with virgin olive oil) augmented LDL resistance against oxidation and
312 decreased the quantity of LDL oxidative modifications. Some of these effects have
313 been previously observed after similar dietary interventions [9, 10]. As a possible
314 explanation, TMD dietary antioxidants may bind to LDL or preserve other dietary
315 antioxidants in the lipoprotein (e.g., vitamin E) in a non-oxidized state, increasing the
316 resistance of the lipoprotein against oxidative attacks [21].

317 Small LDL particles are also more atherogenic [22]: they remain longer in circulation
318 (they interact more poorly with LDL receptors), are more easily oxidized, and tend to
319 traverse the endothelial barrier more than large ones [23]. Therefore, high
320 concentrations of small LDL lipoproteins have been associated with a greater incidence
321 of coronary heart disease [24]. In our trial, the TMD-VOO intervention increased
322 estimated LDL particle size (measured as the LDL-C/ApoB ratio [12]), in agreement
323 with the effects induced by other similar interventions such as the consumption of virgin
324 olive oil [10] or adherence to a TMD enriched with nuts [8]. The improvement in the
325 general oxidative status after the intervention could contribute to explaining this benefit,
326 since pro-oxidative states are linked to an increased number of small LDL particles in
327 circulation [25].

328 LDL composition affects the atherogenicity of the particle. On the one hand,
329 cholesterol-poor, triglyceride-rich LDL particles are present in high cardiovascular risk
330 states [26] and have been related to changes in ApoB conformation that hinder its
331 binding to LDL receptors [27]. On the other hand, although 95% of LDL protein is
332 ApoB, LDLs are known to be able to bind some proteins that may be detrimental
333 (apolipoprotein C-III, pro-inflammatory proteins such as serum amyloid A4 and
334 elements of the complement system, and pro-thrombotic proteins such as the
335 fibrinogen α chain). Therefore, an increase in LDL protein content different from ApoB
336 may be considered an indirect sign of a dysfunctional, pro-inflammatory, pro-thrombotic
337 particle [28]. Moreover, the most atherogenic LDL (small, dense, electronegative) is
338 also protein-rich [28]. According to our data, adherence to the TMD-VOO intervention
339 made LDL particles cholesterol-rich (they carried more cholesterol per each ApoB
340 molecule). In addition, the levels of proteins other than ApoB in LDL lipoproteins
341 decreased after both TMD interventions and the low-fat diet. These two changes could
342 have contributed to diminishing LDL atherogenicity.

343 Atherogenic LDL particles are toxic for some cell types: when macrophages phagocyte
344 modified LDL lipoproteins, the cells begin to release pro-inflammatory signals and
345 finally become foam cells [15]. In the present trial, the TMD-VOO intervention
346 decreased LDL cytotoxicity in human macrophages. In this regard, an *in vitro* treatment
347 with a flavonoid-rich extract has been previously reported to decrease the cytotoxic
348 response induced by oxidized LDL on macrophages [13]. However, this is the first time
349 that an intervention in humans has been able to decrease the *ex vivo* cytotoxicity of
350 LDL particles. The improved oxidative status, estimated size, and composition of LDL
351 lipoproteins after the intervention could help to explain this enhancement [14].

352 Nevertheless, the relevance of LDL *ex vivo* cytotoxicity in the development of
353 cardiovascular outcomes remains to be elucidated in future trials.

354 According to our data, all the benefits of the TMD-VOO intervention on LDL
355 atherogenic characteristics seemed inter-related (anti-atherogenic LDL traits were

356 associated among each other at baseline, as well as most changes after the TMD-VOO
357 intervention) and independent from LDL-C or ApoB quantity. This evidence supports
358 the hypothesis that adherence to a TMD (particularly when enriched with virgin olive
359 oil) may lead to the development of a less atherogenic LDL phenotype [29]. Although
360 not directly examined in this study, this phenotype could be partially responsible for
361 some of the cardioprotective benefits of the Mediterranean Diet.

362 Another general comment in this work could be the potentially deleterious effect of the
363 low-fat diet on characteristics beyond the lipid profile. Although this diet was able to
364 decrease the quantity of LDL-C in plasma, it reduced the estimated values of LDL
365 particle size (LDL-C levels decreased whilst ApoB levels did not, possibly leading to an
366 increase in the pro-atherogenic, cholesterol-poor, small LDLs) and also increased
367 remnant cholesterol levels (another lipid parameter associated with greater
368 cardiovascular risk [30]). These detrimental traits may contribute to explaining why
369 TMD is more cardioprotective than a low-fat diet, and could also highlight that,
370 regarding the lipid profile, quality may be more relevant than quantity.

371 Our study has various strengths. First, it presents a randomized design and involves an
372 active comparator (the low fat control intervention). Second, it comprises a large
373 sample size ($N=210$) and a long intervention duration (one year). Finally, it studies
374 comprehensively several LDL atherogenic traits and their interrelationships.

375 Nevertheless, the study also has limitations. The volunteers were elderly people with
376 high cardiovascular risk values; hence the extrapolation of our results to the general
377 population is complex. The results obtained were modest because: 1) the dietary
378 intervention in our trial is based on discreet lifestyle changes; and 2) the low-fat control
379 intervention is a well-known healthy diet. Finally, although the sample selection was
380 random, and the baseline characteristics of the three groups were comparable, they
381 varied modestly from the baseline characteristics of the whole PREDIMED Study
382 population. Differences among the changes observed in our results and other
383 PREDIMED Study sub-samples, particularly relative to lipid profile, could be due to the

384 longer duration of the intervention in our sub-group, and the varying proportion of
385 patients with dyslipidemia. Nevertheless, to take into consideration all the possible
386 confounders, we included age, sex, center, and changes in classical cardiovascular
387 risk factors as co-variables in our multivariate linear regression analyses.

388 In conclusion, adherence to a TMD, particularly when enriched with virgin olive oil,
389 decreased LDL atherogenicity (ameliorating LDL characteristics related to oxidation,
390 estimated size, and composition) and LDL *ex vivo* cytotoxicity. These data reinforce the
391 previous evidence regarding the healthy effects of the Mediterranean Diet, since the
392 development of a less atherogenic LDL phenotype could be a possible explanation for
393 some of the cardioprotective benefits of this dietary pattern.

394 **AUTHOR CONTRIBUTIONS**

395 A.H. and M.Fitó designed the experiments. A.H. performed the experimental work,
396 interpreted the data, and drafted the manuscript. R.T. and M.C.L-S. contributed to the
397 experimental development. A.H., O.C., A.G., and M.Fitó contributed in the search of
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401

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413 **CONFLICT OF INTEREST**

414 Authors have no conflict of interest to declare for this research.

415 **REFERENCES**

416

417 [1] Martínez-Gonzalez, M.A., Bes-Rastrollo, M., Dietary patterns, Mediterranean
418 diet, and cardiovascular disease. *Curr. Opin. Lipidol.* 2014, 25, 20–6.

419 [2] Estruch, R., Ros, E., Salas-Salvadó, J., Covas, M.-I., et al., Primary prevention
420 of cardiovascular disease with a Mediterranean diet. *N. Engl. J. Med.* 2013, 368,
421 1279–90.

422 [3] Ros, E., Martínez-González, M.A., Estruch, R., Salas-Salvadó, J., et al.,
423 Mediterranean diet and cardiovascular health: Teachings of the PREDIMED
424 study. *Adv. Nutr.* 2014, 5, 330S–6S.

425 [4] Estruch, R., Martínez-González, M.A., Corella, D., Salas-Salvadó, J., et al.,
426 Effects of a Mediterranean-style diet on cardiovascular risk factors: a
427 randomized trial. *Ann. Intern. Med.* 2006, 145, 1–11.

428 [5] Fitó, M., Estruch, R., Salas-Salvadó, J., Martínez-Gonzalez, M.A., et al., Effect of
429 the Mediterranean diet on heart failure biomarkers: a randomized sample from
430 the PREDIMED trial. *Eur. J. Heart Fail.* 2014, 16, 543–50.

431 [6] Mitjavila, M.T., Fandos, M., Salas-Salvadó, J., Covas, M.-I., et al., The
432 Mediterranean diet improves the systemic lipid and DNA oxidative damage in
433 metabolic syndrome individuals. A randomized, controlled, trial. *Clin. Nutr.* 2013,
434 32, 172–8.

435 [7] Castañer, O., Corella, D., Covas, M.-I., Sorlí, J. V, et al., In vivo transcriptomic
436 profile after a Mediterranean diet in high-cardiovascular risk patients: a
437 randomized controlled trial. *Am. J. Clin. Nutr.* 2013, 98, 845–53.

438 [8] Damasceno, N.R.T., Sala-Vila, A., Cofán, M., Pérez-Heras, A.M., et al.,
439 Mediterranean diet supplemented with nuts reduces waist circumference and
440 shifts lipoprotein subfractions to a less atherogenic pattern in subjects at high
441 cardiovascular risk. *Atherosclerosis* 2013, 230, 347–53.

442 [9] Fitó, M., Guxens, M., Corella, D., Sáez, G., et al., Effect of a traditional

- 443 Mediterranean diet on lipoprotein oxidation: a randomized controlled trial. *Arch.*
444 *Intern. Med.* 2007, 167, 1195–203.
- 445 [10] Hernáez, Á., Remaley, A.T., Farràs, M., Fernández-Castillejo, S., et al., Olive Oil
446 Polyphenols Decrease LDL Concentrations and LDL Atherogenicity in Men in a
447 Randomized Controlled Trial. *J. Nutr.* 2015, 145, 1692–7.
- 448 [11] Hernáez, Á., Castañer, O., Elosua, R., Pintó, X., et al., Mediterranean Diet
449 Improves High-Density Lipoprotein Function in High-Cardiovascular-Risk
450 IndividualsClinical Perspective. *Circulation* 2017, 135, 633–643.
- 451 [12] Wägner, A.M., Jorba, O., Rigla, M., Alonso, E., et al., LDL-
452 cholesterol/apolipoprotein B ratio is a good predictor of LDL phenotype B in type
453 2 diabetes. *Acta Diabetol.* 2002, 39, 215–20.
- 454 [13] Gajaria, T.K., Patel, D.K., Devkar, R. V, Ramachandran, A. V, Flavonoid rich
455 extract of *Murraya Koenigii* alleviates in-vitro LDL oxidation and oxidized LDL
456 induced apoptosis in raw 264.7 Murine macrophage cells. *J. Food Sci. Technol.*
457 2015, 52, 3367–75.
- 458 [14] Steinberg, D., The LDL modification hypothesis of atherogenesis: an update. *J.*
459 *Lipid Res.* 2009, 50 Suppl, S376-81.
- 460 [15] Ross, R., Atherosclerosis--an inflammatory disease. *N. Engl. J. Med.* 1999, 340,
461 115–26.
- 462 [16] Badimon, L., Martínez-González, J., Llorente-Cortés, V., Rodríguez, C., et al.,
463 Cell biology and lipoproteins in atherosclerosis. *Curr. Mol. Med.* 2006, 6, 439–
464 56.
- 465 [17] Ho, E., Karimi Galougahi, K., Liu, C.-C., Bhindi, R., et al., Biological markers of
466 oxidative stress: Applications to cardiovascular research and practice. *Redox*
467 *Biol.* 2013, 1, 483–491.
- 468 [18] Ohmura, H., Mokuno, H., Sawano, M., Hatsumi, C., et al., Lipid compositional
469 differences of small, dense low-density lipoprotein particle influence its oxidative
470 susceptibility: possible implication of increased risk of coronary artery disease in

- 471 subjects with phenotype B. *Metabolism*. 2002, 51, 1081–7.
- 472 [19] Gómez, M., Vila, J., Elosua, R., Molina, L., et al., Relationship of lipid oxidation
473 with subclinical atherosclerosis and 10-year coronary events in general
474 population. *Atherosclerosis* 2014, 232, 134–140.
- 475 [20] Wu, T., Willett, W.C., Rifai, N., Shai, I., et al., Is Plasma Oxidized Low-Density
476 Lipoprotein, Measured With the Widely Used Antibody 4E6, an Independent
477 Predictor of Coronary Heart Disease Among U.S. Men and Women? *J. Am. Coll.*
478 *Cardiol.* 2006, 48, 973–979.
- 479 [21] Gimeno, E., Fitó, M., Lamuela-Raventós, R.M., Castellote, A.I., et al., Effect of
480 ingestion of virgin olive oil on human low-density lipoprotein composition. *Eur. J.*
481 *Clin. Nutr.* 2002, 56, 114–20.
- 482 [22] Carmena, R., Duriez, P., Fruchart, J.-C., Atherogenic lipoprotein particles in
483 atherosclerosis. *Circulation* 2004, 109, III2-7.
- 484 [23] Kwiterovich, P.O., Clinical relevance of the biochemical, metabolic, and genetic
485 factors that influence low-density lipoprotein heterogeneity. *Am. J. Cardiol.* 2002,
486 90, 30i–47i.
- 487 [24] Hoogeveen, R.C., Gaubatz, J.W., Sun, W., Dodge, R.C., et al., Small dense low-
488 density lipoprotein-cholesterol concentrations predict risk for coronary heart
489 disease: the Atherosclerosis Risk In Communities (ARIC) study. *Arterioscler.*
490 *Thromb. Vasc. Biol.* 2014, 34, 1069–77.
- 491 [25] Kotani, K., Tsuzaki, K., Taniguchi, N., Sakane, N., LDL Particle Size and
492 Reactive Oxygen Metabolites in Dyslipidemic Patients. *Int. J. Prev. Med.* 2012,
493 3, 160–6.
- 494 [26] Kunitake, S.T., Young, S.G., Chen, G.C., Pullinger, C.R., et al., Conformation of
495 apolipoprotein B-100 in the low density lipoproteins of tangier disease.
496 Identification of localized conformational response to triglyceride content. *J. Biol.*
497 *Chem.* 1990, 265, 20739–46.
- 498 [27] Aviram, M., Lund-Katz, S., Phillips, M.C., Chait, A., The influence of the

499 triglyceride content of low density lipoprotein on the interaction of apolipoprotein
500 B-100 with cells. *J. Biol. Chem.* 1988, 263, 16842–8.

501 [28] Diffenderfer, M.R., Schaefer, E.J., The composition and metabolism of large and
502 small LDL. *Curr. Opin. Lipidol.* 2014, 25, 221–6.

503 [29] Krauss, R.M., Atherogenic lipoprotein phenotype and diet-gene interactions. *J.*
504 *Nutr.* 2001, 131, 340S–3S.

505 [30] Varbo, A., Benn, M., Nordestgaard, B.G., Remnant cholesterol as a cause of
506 ischemic heart disease: Evidence, definition, measurement, atherogenicity, high
507 risk patients, and present and future treatment. *Pharmacol. Ther.* 2014, 141,
508 358–367.

509

510 **FIGURE LEGENDS**

511

512 **Figure 1.** Effects of the Traditional Mediterranean Diet enriched with virgin olive oil
513 (TMD-VOO) or nuts (TMD-Nuts), relative to a low-fat diet, on LDL-C levels (**A-B**), ApoB
514 concentrations (**C-D**), remnant cholesterol levels (**E-F**), and estimated LDL particle size
515 (LDL-C/ApoB ratio) (**G-H**). **A,C,E,G.** Post- vs. pre-intervention changes (mean, 95%
516 CI). **B,D,F,H.** Inter-treatment differences in a multivariate linear regression model
517 adjusted for: age; sex; center of origin of the volunteer; baseline value of the variable;
518 and changes in the presence of dyslipidemia, diabetes, hypertension, and smoking
519 habit throughout the study (adjusted coefficient, 95% CI). *: $P < 0.05$; **: $P < 0.01$; ***:
520 $P < 0.001$.

521

522 **Figure 2.** Effects of the Traditional Mediterranean Diet enriched with virgin olive oil
523 (TMD-VOO) or nuts (TMD-Nuts), relative to a low-fat diet, on the resistance of LDL
524 particles against oxidation (LDL lag time) (**A-B**) and LDL oxidation (**C-D**). **A,C.** Post- vs.
525 pre-intervention changes (mean, 95% CI). **B,D.** Inter-treatment differences in a
526 multivariate linear regression model adjusted for: age; sex; center of origin of the
527 volunteer; baseline value of the variable; and changes in the presence of dyslipidemia,
528 diabetes, hypertension, and smoking habit throughout the study (adjusted coefficient,
529 95% CI). *: $P < 0.05$; **: $P < 0.01$.

530

531 **Figure 3.** Effects of the Traditional Mediterranean Diet enriched with virgin olive oil
532 (TMD-VOO) or nuts (TMD-Nuts), relative to a low-fat diet, on the cholesterol content in
533 LDL particles (**A-B**), and the percentage of LDL proteins other than apolipoprotein B
534 (**C-D**). **A,C.** Post- vs. pre-intervention changes (mean, 95% CI). **B,D.** Inter-treatment
535 differences in a multivariate linear regression model adjusted for: age; sex; center of
536 origin of the volunteer; baseline value of the variable; and changes in the presence of

537 dyslipidemia, diabetes, hypertension, and smoking habit throughout the study (adjusted
538 coefficient, 95% CI). *: $P < 0.05$; **: $P < 0.01$.

539

540 **Figure 4.** Effects of the Traditional Mediterranean Diet enriched with virgin olive oil
541 (TMD-VOO) or nuts (TMD-Nuts), relative to a low-fat diet, on the cytotoxicity of LDL
542 particles in macrophages (**A-B**). **A.** Post- vs. pre-intervention changes (mean, 95% CI).
543 **B.** Inter-treatment differences in a multivariate linear regression model adjusted for:
544 age; sex; center of origin of the volunteer; baseline value of the variable; and changes
545 in the presence of dyslipidemia, diabetes, hypertension, and smoking habit throughout
546 the study (adjusted coefficient, 95% CI). *: $P < 0.05$.

547 **TABLES**

548

549 **Table 1.** Baseline characteristics of the volunteers in the three intervention groups.

550

VARIABLES	TMD-VOO N=71	TMD-Nuts N=68	Low-fat diet N=71	P-value
Age (years)	66.5 ± 6.34	65.1 ± 6.85	64.7 ± 6.58	0.270
Sex (% male)	45.1%	61.8%	47.9%	0.111
Body Mass Index (kg/m ²)	30.2 ± 3.96	29.2 ± 3.92	29.7 ± 3.98	0.386
Waist Circumference (cm)	99.8 ± 10.7	102 ± 10.2	101 ± 11.5	0.489
Leisure-time physical activity (MET·min/day)	156 (67.5-247)	169 (59.1-323)	150 (15.5-332)	0.782
Smoking status (% of smokers)	16.9%	11.8%	12.7%	0.642
Type 2 diabetes (% of diabetic patients)	76.1%	76.5%	84.5%	0.380
Hypertension (% of hypertensive patients)	47.9%	55.9%	38.0%	0.107
Dyslipidemia (% of dyslipidemic patients)	83.1%	77.9%	85.9%	0.458
Fasting glucose (mg/dL)	105 (92.5-127)	118 (96.0-140)	105 (94.0-128)	0.470
Triglycerides (mg/dL)	108 (90.7-157)	105 (73.0-147)	115 (97.0-140)	0.610
Total cholesterol (mg/dL)	206 ± 39.1	198 ± 35.9	210 ± 38.4	0.231
HDL cholesterol (mg/dL)	49.8 ± 11.8	49.2 ± 10.8	49.2 ± 10.6	0.932
LDL cholesterol (mg/dL)	129 ± 30.0	125 ± 30.1	135 ± 33.0	0.190
Apolipoprotein B (mg/dL)	104 ± 22.0	97.6 ± 17.1	105 ± 22.7	0.121
Apolipoprotein B/A-I ratio (unitless)	0.78 ± 0.16	0.75 ± 0.16	0.82 ± 0.22	0.123

551 Variables are expressed as percentages (categorical variables), means \pm SD (normally
552 distributed variables) or median (1st-3rd quartile) (non-normally distributed variables).
553 *MET*: metabolic equivalent of task. *TMD-Nuts*: Traditional Mediterranean Diet enriched
554 with mixed nuts. *TMD-VOO*: Traditional Mediterranean Diet enriched with virgin olive
555 oil.