

1 **Grape seed proanthocyanidins correct**
2 **dyslipidemia associated with a high-fat diet**
3 **in rats and repress genes controlling**
4 **lipogenesis and VLDL assembling in liver**

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14

15 **Abstract**

16 **Objective:** To determine whether proanthocyanidins can protect against dyslipidemia induced by a high-
17 fat diet (HFD) and to address the mechanisms that underlie this hypolipidemic effect.

18 **Design and measurements:** Female Wistar rats were fed on a HFD for 13 weeks. They were divided into
19 two groups, one of which was treated with a grape seed proanthocyanidin extract (25 mg/kg of body
20 weight) for 10 days. Plasma and liver lipids were measured by colorimetric and gravimetric analysis.

21 Liver, muscle and adipose tissue were used to study the expression of genes involved in the synthesis and
22 oxidation of fatty acids and lipoprotein homeostasis by real-time RT-PCR.

23 **Results:** The administration of proanthocyanidins normalized plasma triglyceride and LDL-cholesterol
24 (both parameters significantly increased with the HFD) but tended to decrease hypercholesterolemia and
25 fatty liver. Gene expression analyses revealed that proanthocyanidins repressed both the expression of
26 hepatic key regulators of lipogenesis and very low density lipoprotein (VLDL) assembling such as
27 SREBP1, MTP and DGAT2, all of which were overexpressed by the HFD.

28 **Conclusion:** These findings indicate that natural proanthocyanidins improve dyslipidemia associated with
29 HFDs, mainly by repressing lipogenesis and VLDL assembly in the liver, and support the idea that they
30 are powerful agents for preventing and treating lipid altered metabolic states.

31 **Keywords:** high-fat diet; liver; proanthocyanidins; triglycerides; SREBP1; MTP

32 **Introduction**

33 Hypertriglyceridemia is a strong predictor of atherogenic cardiovascular disease (CVD).¹ In both the
34 metabolic syndrome and type 2 diabetes, hypertriglyceridemia is the result of increased plasma
35 concentration of very low density lipoprotein (VLDL).² This increase is the consequence of
36 overproduction of VLDL by the liver, and the possible delayed catabolism of these lipoproteins, caused
37 by insulin resistance.³ In turn, elevated VLDL and hypertriglyceridemia reduce the high-density
38 lipoprotein (HDL) level and generate small, dense low-density lipoprotein (LDL) due to lipid exchange.⁴
39 High serum triglyceride (TG) levels, low serum HDL-cholesterol (HDL-C) levels and a preponderance of
40 small, dense LDL particles are the ‘atherogenic lipid triada’ characteristic of the dyslipidemia that
41 commonly occurs in the metabolic syndrome.⁵ It has also been suggested that the fact that cardiovascular
42 risk indexes are lower in obese patients who lose weight may be closely connected to a reduction in
43 VLDL secretion by the liver.⁶

44

45 Flavan-3-ols and their oligomeric condensation products, proanthocyanidins (PA), are the most common
46 group of flavonoids in the American diet.⁷ PA can be found in such common foodstuffs as cereals,
47 legumes, fruits, vegetables and beverages (red wine and tea, in particular).⁸ The health benefits of PA
48 have been most studied in tea, cocoa and grape seed, each of which has a characteristic and specific
49 oligomeric composition that conditions its biological activity. Grape seed proanthocyanidin extracts
50 (GSPE) reduce foam cells,⁹ prevent aortic atherosclerosis¹⁰ from developing in a hamster atherosclerosis
51 model, decrease oxidized LDL in hypercholesterolemic humans⁹ and improve endothelial function by
52 modifying NO production.¹¹ Consequently, they protect against atherosclerosis and CVD. Besides their
53 antioxidant activity and their effects on the vascular endothelium, the antiatherogenic properties of PA are
54 also related to an improved serum lipid profile. Plasma TG and apolipoprotein B (apoB) levels are
55 reduced by GSPE in normolipidemic rats,¹² and by lyophilized grape powder in postmenopausal
56 women.¹³ The lipoprotein profile is also improved in both healthy subjects and hemodialysis patients by
57 concentrated red grape juice.¹⁴

58

59 Although PA can exert part of their hypolipidemic effect by inhibiting the absorption of dietary lipids and
60 diminishing chylomicron secretion by enterocytes, the liver has an important role in reducing plasma TG
61 through GSPE. We have determined that GSPE limit VLDL secretion by repressing lipogenic genes and
62 the microsomal transfer protein (MTP; the key controller of VLDL assembling)¹² and by overexpressing
63 the carnitine palmitoiltransferase-1 (CPT1; the key controller of free fatty acid (FFA) oxidation)¹⁵ in
64 mouse liver. GSPE exert some of these effects by a pathway that involves overexpressing the nuclear
65 receptor small heterodimer partner (SHP) and repressing the transcription factor sterol regulatory
66 element-binding protein 1 (SREBP1).¹⁵

67

68 As overproduction of VLDL and hypertriglyceridemia are at the basis of atherogenic dyslipidemia, and
69 GSPE can inhibit VLDL secretion in a healthy situation, the objectives of this study were to determine
70 whether proanthocyanidins could prevent rats from developing atherogenic dyslipidemia and to establish
71 the mechanism underlying this hypolipidemic effect. To this end, we have used rats fed on a high-fat diet
72 (HFD) as a model of atherogenic dyslipidemia and we have determined the role of liver and extrahepatic
73 tissue in normalizing the lipid profile.

74 **Methods**

75 Proanthocyanidin extract

76 GSPE was kindly provided by Les Dérives Résiniques et Terpéniques (Dax, France). This
77 proanthocyanidin extract contained essentially monomeric (21.3%), dimeric (17.4%), trimeric (16.3%),
78 tetrameric (13.3%) and oligomeric (5–13 units; 31.7%) proanthocyanidins.

79 Animals

80 Female Wistar rats weighing 150g were purchased from Charles River (Barcelona, Spain). The Animal
81 Ethics Committee of our University approved all procedures. The animals were housed in animal quarters
82 at 22°C with a 12-h light/dark cycle (light from 0800 hours to 2000 hours) and were fed ad libitum with
83 standard chow diet (Panlab, Barcelona, Spain). After 5 days, 12 rats were fed ad libitum with standard
84 chow plus a cafeteria diet as an HFD model which had 13.6% fats, 21% carbohydrates and 9% protein.
85 The cafeteria diet consisted of the following foods: cookies with foie-gras and cheese triangles, sweets,
86 bacon, biscuits, chocolate, croissants, carrots and sugary milk. Six rats were kept on the standard chow
87 diet (control group).

88 After 13 weeks, rats feeding on the HFD were trained to lick condensed milk (1 ml) and were divided into
89 two groups. One group was supplemented every day, at 0900 hours, with 25 mg of GSPE per kg body
90 weight dissolved in condensed milk (HFD-GSPE group). The other group received the same volume of
91 condensed milk (HFD group).

92 On day 10 of the GSPE treatment, all the rats were killed at 1400 hours by beheading and the blood was
93 collected using heparin as the anticoagulant. Plasma was obtained by centrifugation and stored at -80°C
94 until analysis. Liver, leg muscle and mesenteric adipose tissue were excised and frozen immediately in
95 liquid nitrogen and stored at -80°C until RNA and lipid extraction.

96 Plasma and liver lipid analysis

97 Plasma total cholesterol (TC) was measured with an enzymatic colorimetric kit (QCA, Barcelona, Spain).
98 HDL-C was measured, using the same kit, after treating the plasma with phosphotungstic acid to
99 precipitate the non-HDL lipoproteins.¹⁷ The LDL-cholesterol (LDL-C) was measured after plasma
100 treatment with polyvinyl sulfate and polyethylene glycol monomethyl ether to precipitate LDL
101 lipoproteins. LDL-C was calculated as TC minus cholesterol in plasma after LDL precipitation.¹⁸ TGs
102 were assayed using an enzymatic colorimetric kit (QCA). FFA were measured using an enzymatic
103 colorimetric kit (Wako chemicals GmbH, Madrid, Spain).

104

105 Liver lipids (0.5 g) were extracted using the Folch method.¹⁹ An aliquot of extract was used to measure
106 the total lipids by gravimetry. The rest of the extract was evaporated to dryness and redissolved in 2%
107 triton X-100 to determine the TG and TC using the same kits that were used for plasma quantification.

108 Gene expression analyses

109 Total RNA from the liver was obtained using a NucleoSpin RNA2 kit (Macherie-Naegel, Germany) and
110 total RNA from muscle and adipose tissue was obtained using Trizol reagent (Invitrogen, Barcelona,
111 Spain) following the manufacturer's protocol. Additional purification and DNase treatment was
112 performed using a NucleoSpin RNA2 kit (Macherie-Naegel, Germany). cDNA was synthesized from 2
113 µg of total RNA using the Taqman Reverse transcription reagent kit (Applied Biosystems). A total of 20
114 ng of cDNA was subjected to quantitative RT-PCR amplification using Taqman Master Mix (Applied
115 Biosystems). Specific Taqman probes (Applied Biosystems) were used for different genes. Ppia was used
116 as an endogenous control. Real-time quantitative PCR reactions were performed using the ABI Prism

117 7300 SDS Real-Time PCR system (Applied Biosystems).

118 Statistical analysis

119 Results are reported as mean \pm s.e.m. of six animals. Group means were compared with an independent-
120 samples Student's t-test ($P \leq 0.05$) using SPSS software.

121 **Results**

122 GSPE treatment prevents dyslipidemia induced by high-fat diet

123 The body weight of rats fed with an HFD was significantly higher (approximately 40%) than those in the
124 control group (Table 1). After 13 weeks of HFD administration, initial body mass of the rats increased by
125 126%, whereas that of rats fed with the standard chow diet only increased by 73%. The body weight of
126 HFD-fed rats reduced slightly when treated with GSPE for 10 days (Table 1).

127 HFD also significantly increased liver mass, hepatic lipids, TG and TC levels (Table 1). GSPE treatment
128 slightly lowered TG and TC content in liver, although the total lipids remained as elevated as in
129 nontreated HFD rats.

130 HFD-fed rats were normogluceic (results not shown) but presented hypertriglyceridemia and
131 hypercholesterolemia and increased HDL-C and LDL-C (Table 2). Moreover, the HDL-C/LDL-C ratio,
132 calculated to evaluate the atherosclerosis risk, was reduced in HFD-fed rats, which indicated a greater risk
133 of atherosclerosis.

134 Treatment with GSPE reversed the dyslipidemia induced by the HFD. The plasma levels of TG and LDL-
135 C decreased to the same values observed in the control group of normolipidemic rats (Table 2).

136 Consequently, the atherogenic risk index HDL-C/ LDL-C improved in GSPE-treated animals.

137 Additionally, FFA levels, which were not affected by the HFD, were significantly reduced by GSPE
138 treatment (Table 2). Plasma TC and HDL-C, however, were only slightly reduced (Table 2).

139 GSPE treatment counteracts hepatic overexpression of SREBP1, MTP and DGAT2 induced by a high-fat
140 diet

141 The liver governs the homeostasis of circulating lipids and lipoproteins. Thus, we used reverse
142 transcription-PCR to analyze the differential expression of key genes controlling TG and cholesterol
143 metabolism in the liver (Table 3) and gain further insight into how GSPE improves the plasma lipid
144 profile. We have chosen genes that encode key proteins in cholesterol pathways (LDL-receptor, CYP7A1
145 and HMG-CoA reductase), in fatty acid oxidation (CPT1-a), in TG synthesis (DGAT2) and in VLDL
146 assembling (apoB and MTP). We have also selected two nuclear receptors, namely FXR and SHP, and
147 the transcription factor SREBP1 because they govern the expression of key lipid metabolism genes in the
148 liver and are involved in the molecular mechanism used by GSPE in the liver.

149 The liver of HFD rats showed a significant repression of SHP and CPT1-a in concert with a significant
150 overexpression of DGAT2 when compared to the control group. SREBP1 and MTP also showed a slight
151 overexpression. This expression profile in the liver of HFD rats suggests active TG synthesis and VLDL
152 assembling as well as impaired fatty acid oxidation, which is consistent with the fatty liver,
153 hypercholesterolemia and hypertriglyceridemia present in these animals.

154 In contrast, the liver of HFD-fed rats treated with GSPE for 10 days showed a significant repression of
155 SREBP1, MTP and DGAT2 versus the HFD group. GSPE treatment also increased SHP, although the
156 expression was lower than that of the control group. GSPE treatment did not affect the expression of
157 CPT1-a, which remained repressed as in the HFD group. The changes induced by GSPE treatment in
158 HFD-fed rats strongly suggest that proanthocyanidins repress TG synthesis and VLDL assembling,
159 processes that are exacerbated by an HFD diet. We observed, however, that neither GSPE nor HFD
160 affected the expression of the genes involved in cholesterol metabolism, which suggests that the effects of
161 both the HFD diet and GSPE on plasma cholesterol are not linked to changes in hepatic gene expression.

162 Given that plasma TG levels heavily depend on extrahepatic uptake, we quantified lipoprotein lipase
163 (LPL) and CPT1 expression in adipose tissue and muscle (results not shown). Neither an HFD diet nor
164 GSPE treatment induced any significant change in the mRNA levels of the enzymes controlling the
165 uptake of TG and fatty acid oxidation.

166 **Discussion**

167 Metabolic syndrome and obesity are associated with an increased risk of CVD, in part, due to their
168 association with atherogenic dyslipidemia.²⁰ Overproduction of VLDL and hypertriglyceridemia are the
169 basis of atherogenic dyslipidemia,⁴ so managing VLDL secretion and hypertriglyceridemia could be a
170 good strategy for reducing the risk of CVD associated with these pathologies. PA, a group of flavonoids
171 that can be found in common foodstuffs,⁸ actively reduce plasma TG and apoB in normolipidemic rats,¹²
172 in hamsters fed a hypercholesterolemic diet¹⁰ and in humans.¹¹ In a previous study we showed that an
173 acute dose of GSPE decreases VLDL secretion by repressing MTP and lipogenic genes in
174 normolipidemic rats.¹² This study, then, intended to determine, first, the ability of PA to correct
175 atherogenic dyslipidemia associated with obesity and, second, to gain insight into the mechanisms that
176 underlie the improvement of plasma TG levels, the effects of PA on the lipid metabolism in liver and in
177 extrahepatic tissues.

178

179 With this purpose, we chose a cafeteria diet as an HFD model to induce obesity in rats. The cafeteria diet
180 is a feeding regime in which animals are offered a choice of several palatable food items of varied
181 composition, appearance and texture in addition to their normal chow diet.²¹ This diet induces obesity
182 due to hyperphagia,²¹ and mimics human behavior when the control system for food intake is
183 overwhelmed by psychological or social influences. Rats fed with this diet for 15 weeks showed obesity,
184 hypertriglyceridemia, hypercholesterolemia, elevated plasma LDL-C and fatty liver. Ten days of oral
185 intake of GSPE normalized plasma TG and LDL-C. Therefore, the reduction in TG levels and LDL-C, in
186 association with the increased HDL-C/LDL-C ratio, determined an improvement in the atherosclerotic
187 risk after GSPE intake. However, GSPE did not correct obesity, and only slightly reduced
188 hypercholesterolemia and fatty liver.

189

190 In a cafeteria diet approach, it is very difficult to exactly quantify the ingestion of each food, so we cannot
191 be sure that GSPE treatment changed the food pattern. However, although GSPE treatment can induce
192 changes in food preferences, which can indirectly lead to a plasma lipid reduction, a direct effect of GSPE
193 could not be discounted because in previous studies we found a considerable reduction in plasma TGs and
194 LDL-C in other experimental models that consumed no food after GSPE treatment. In this study with
195 female hyperlipidemic rats, the effects of chronic GSPE treatment on plasma lipids are similar to those
196 observed in a previous study on the effects of acute GSPE treatment with male normolipidemic rats.¹² In
197 this study, GSPE treatment reduced plasma TG, TC and LDL-C levels by about 40, 13 and 40% with
198 respect to animals fed the HFD, respectively. In the previous study, GSPE treatment reduced plasma TG,
199 TC and LDL-C levels by about 50, 12 and 43% with respect to the untreated animals, respectively. GSPE,
200 then, is also a powerful agent for reducing plasma TG and LDL-C in dyslipidemia associated to HFD.
201 GSPE treatment was also effective in both male and female rats, so it seems that sex does not affect GSPE
202 efficiency on plasma lipids.

203

204 The liver gene expression of rats fed an HFD indicated active lipogenesis and impaired fatty acid
205 oxidation together with increased assembling of VLDL, which points to the liver as an important
206 contributor to HFD-induced atherogenic dyslipidemia. Oral GSPE treatment repressed SREBP1, DGAT2
207 and MTP. These three proteins are key regulators of VLDL synthesis and secretion. SREBP1 activates the
208 expression of several genes involved in FFA and TG synthesis, as well as other components of the
209 regulatory machinery of lipid metabolism. Although it can be regulated at different levels, its
210 transcriptional repression has been linked to lower FFA and TG synthesis and release from the liver,
211 leading to hypolipidemic states.²² Therefore, the repressive effects of GSPE on SREBP1 may account for
212 the lower plasma levels of TG. Together with SREBP1, DGAT2 is a key enzyme in the FFA

213 reesterification process that delivers TG to the nascent VLDL.²³ As the availability of lipids is a key
214 factor that drives the synthesis and secretion of VLDL,²³ the repression of DGAT-2 and SREBP1
215 suggests that lipoprotein release may be blocked at the primary level, that is, fewer lipids are available for
216 lipoprotein assembly. The repression of MTP also suggests a decreased assembly of VLDL, because MTP
217 is responsible for the association of apoB with lipids and the intracellular trafficking of the newly
218 synthesized VLDL. Altogether, the repression of MTP, SREBP1 and DGAT2 by GSPE suggests a VLDL
219 synthesis blockage at two different levels: first, lipid availability is decreased and then lipoprotein
220 assembly. Moreover, GSPE did not modify the expression of the LDL-receptor, indicating that the
221 decrease in LDL-C by GSPE was not due to an increased uptake of LDL by the liver. Altogether, these
222 results strongly suggest that GSPE improved dyslipidemia in HFD rats by reducing VLDL secretion,
223 which, in turn, led to lower levels of TG and lower LDL production.

224

225 In contrast, GSPE did not increase CPT1-a expression, suggesting that PA did not normalize FFA
226 oxidation in liver. Nevertheless, increased FFA oxidation by GSPE cannot be ruled out because CPT1-a
227 activity is tightly regulated by its physiological inhibitor malonyl-CoA, which physiologically regulates
228 β -oxidation depending on the availability of fatty acids and glucose.²⁴ SREBP1 activates the expression
229 of acetyl-CoA synthetase, the synthesizing enzyme of malonyl-CoA,²⁵ so repression of SREBP1 by
230 GSPE could result in a lower concentration of malonyl-CoA in hepatocytes and, therefore, a lower
231 repression of CPT-1a. For this reason, the effect of GSPE on FFA oxidation still requires further study.

232

233 SHP is a nuclear receptor that acts as an inducible repressor of other nuclear receptors and transcription
234 factors. With this mechanism, it controls lipogenesis and cholesterol metabolism in liver.²⁶ Thus, the low
235 levels of SHP expression in HFD-fed rats could be behind the exacerbated expression of SREBP1, which,
236 in turn, induces overexpression of lipogenic genes, fatty liver and hypertriglyceridemia.²⁵ In a previous

237 study we showed that GSPE uses an SHP-dependent molecular mechanism to reduce TG secretion in
238 HepG2,15 so the slight increase in SHP expression induced by GSPE could be sufficient to reduce TG
239 secretion, but not enough to counteract fatty liver.

240

241 To assess whether lipoprotein uptake and metabolization can account for the hypolipidemic effects of
242 GSPE, LPL expression was analyzed in both muscle and adipose tissue. HFD and GSPE were found to
243 have no effect, suggesting that uptake of TG by these tissues does not affect TG levels in plasma as much
244 as the production of TG by the liver.

245

246 Although GSPE affects the target genes of lipid metabolism in the liver of HFD rats in almost the same
247 way as it does in normolipidemic rats,12 there are some differences, mainly in MTP and CYP7A1
248 expression. MTP expression is not modified by GSPE in normolipidemic rats but it is strongly repressed
249 in HFD rats. On the other hand, CYP7A1 expression is increased by GSPE in normolipidemic rats but it
250 is not affected in HFD rats. These differences can be explained in a variety of ways. In normolipidemic
251 rats GSPE was administered acutely whereas in the present experiment it was administered chronically.
252 The effects of acute administration may be a little different from the effects of chronic administration,
253 where the organism is better adapted to receiving GSPE. Moreover, the metabolic stress induced by the
254 HFD, which exacerbates VLDL assembly, hyperlipidemia and fatty liver, may modify the response to
255 GSPE.

256

257 In conclusion, oral intake of GSPE improves dyslipidemia induced by HFD in rats, mainly by repressing
258 lipogenesis and VLDL assembly in the liver, which are overexpressed as a result of the HFD. Therefore,
259 increasing the intake of PA-rich foods can be a strategy for counteracting atherogenic dyslipidemia

260 associated with obesity and metabolic syndrome.

261

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- 326

327 **Tables**

328 **Table 1:** Body weight and liver lipids of rats fed with a standard diet or high-fat diet, either with or
 329 without proanthocyanidin treatment.

Parameter	Control group	HFD group	HFD-GSPE group
Body weight (g)	282 ± 9.0	392 ± 27*	380 ± 21*
Liver weight (g)	8.99 ± 0.23	11.73 ± 0.88*	10.85 ± 0.64
Liver lipids (mg/g liver)	48.7 ± 5.5	60.9 ± 5.87*	60.3 ± 0.6
Liver cholesterol (mg/g liver)	2.24 ± 0.15	3.66 ± 0.51*	2.7 ± 0.16
Liver triglyceride (mg/g liver)	4.53 ± 0.48	7.25 ± 1.01*	6.55 ± 0.55

330 *Experimental details and symbols: Values are mean ± s.e.m. of six rats. * Indicates a significant*
 331 *difference (P ≤ 0.05) versus control group.*

332 **Table 2:** Plasma lipid levels of rats fed with a standard diet or high-fat diet, either with or without
 333 proanthocyanidin treatment.

Parameter	Control group	HFD group	HFD-GSPE group
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Triglycerides (mg per 100 ml)	107.3 ± 10.6	204.0 ± 2.3*	129.4 ± 12.3†
Total cholesterol (mg per 100 ml)	57.9 ± 2.8	95.9 ± 5.7*	83.5 ± 4.5*
HDL cholesterol (mg per 100 ml)	35.6 ± 7.9	60.6 ± 4.1*	51.0 ± 4.9
LDL cholesterol (mg per 100 ml)	3.5 ± 0.1	15.2 ± 2.0*	6.6 ± 1.0†
HDL-C/LDL-C ratio	8.7 ± 2.2	4.0 ± 0.6	7.0 ± 0.2
Total C/HDL-C ratio	1.4 ± 0.03	1.6 ± 0.15	1.7 ± 0.11
Free fatty acids (mg per 100 ml)	20.5 ± 2.1	22.9 ± 2.0	14.3 ± 1.1

334 *Abbreviations: GSPE, grape seed proanthocyanidin extracts; HFD, high-fat diet. * Indicates a significant*
335 *difference (P ≤ 0.05) versus control group; † indicates that the t-test found a significant difference*
336 *between the HFD and HFD-GSPE groups.*

337 **Table 3:** mRNA levels of lipid-related genes in the liver of rats fed with a standard diet or a high-fat diet,
338 either with or without proanthocyanidin treatment.

Gene	Control group	HFD group	HFD-GSPE group
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FXR	1.05 ± 0.16	0.89 ± 0.13	1.38 ± 0.24
SHP	1.13 ± 0.20	0.29 ± 0.10*	0.46 ± 0.13*
SREBP1	0.98 ± 0.07	1.54 ± 0.42	0.58 ± 0.11
ApoB	1.43 ± 0.32	1.62 ± 0.53	1.48 ± 0.36
MTP	1.08 ± 0.17	1.49 ± 0.06	0.99 ± 0.05†
CPT1-a	1.03 ± 0.02	0.50 ± 0.09*	0.50 ± 0.09*
DGAT2	1.05 ± 0.05	1.80 ± 0.11*	1.19 ± 0.10*
CYP7A1	1.19 ± 0.29	2.63 ± 0.63	1.55 ± 0.40
HMG-CoA reductase	1.13 ± 0.21	0.72 ± 0.04	1.08 ± 0.17
LDL receptor	0.91 ± 0.16	1.05 ± 0.38	0.75 ± 0.17

339 *Abbreviations: GSPE, grape seed proanthocyanidin extracts; HFD, high-fat diet. The values are*
340 *expressed as fold change, using PPIA expression as the endogenous control.*