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Effects of Mediterranean Diet on plasma metabolites and their relationship with insulin resistance and gut microbiota composition in a crossover randomized clinical trial --Manuscript Draft--

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Abstract:	<p>Background & Aims</p> <p>The Mediterranean Diet (MedDiet) may decrease the cardiometabolic risk through modulation of metabolic pathways. Furthermore, the interplay between MedDiet, metabolites and microbial metabolism may improve our understanding on the metabolic effects of this diet. We aimed to evaluate the effect of the MedDiet compared to nuts supplementation on circulating metabolites and their relationship with cardiometabolic health. We further examined whether changes in the metabolomic profiles were associated with changes in gut microbiota composition in a multi-omics integrative approach.</p> <p>Methods</p> <p>Forty-four adults with Metabolic Syndrome (MetS), (aged 37-65) participated in a randomized, controlled, crossover 2-months dietary-intervention trial with a 1-month wash-out period, consuming a MedDiet or a non MedDiet plus nuts (50 g/day). Nutritional data were collected at the beginning and the end of each intervention period using 3-day dietary records, as well as fasting blood and fecal samples. Plasma metabolites (m=378) were profiled using targeted metabolomics. Associations of these metabolites with the interventions were assessed with elastic net regression analyses. Gut microbiota composition was assessed by 16S rRNA sequencing. A sparse least regression analysis combined with a canonical correlation analysis was conducted between the plasma selected metabolites and genera in order to identify the relevant dual-omics signatures discriminating the dietary interventions.</p> <p>Results</p> <p>Changes in 65 circulating metabolites were significantly associated with the MedDiet (mainly lipids, acylcarnitines, amino acids, steroids and TCA intermediates). Importantly, these changes were associated with decreases in glucose, insulin and HOMA-IR. The network analysis identified two main clusters of genera with an opposite behaviour towards selected metabolites, mainly PC species, ChoE(20:5), TGs and medium/long-chain acylcarnitines.</p> <p>Conclusion</p> <p>Following a MedDiet, rather than consuming nuts in the context of a non-MedDiet was associated with a specific plasma metabolomic profile, which was also related to</p>

metabolic improvements in adults with MetS. The identified correlated network between specific bacteria and metabolites suggests an interplay between diet, circulating metabolites and gut microbiota.

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Effects of Mediterranean Diet on plasma metabolites and their relationship with insulin resistance and gut microbiota composition in a crossover randomized clinical trial

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Abstract

Background & Aims The Mediterranean Diet (MedDiet) may decrease the cardiometabolic risk through modulation of metabolic pathways. Furthermore, the interplay between MedDiet, metabolites and microbial metabolism may improve our understanding on the metabolic effects of this diet. We aimed to evaluate the effect of the MedDiet compared to nuts supplementation on circulating metabolites and their relationship with cardiometabolic health. We further examined whether changes in the metabolomic profiles were associated with changes in gut microbiota composition in a multi-omics integrative approach.

Methods Forty-four adults with Metabolic Syndrome (MetS), (aged 37-65) participated in a randomized, controlled, crossover 2-months dietary-intervention trial with a 1-month wash-out period, consuming a MedDiet or a non MedDiet plus nuts (50 g/day). Nutritional data were collected at the beginning and the end of each intervention period using 3-day dietary records, as well as fasting blood and fecal samples. Plasma metabolites (m=378) were profiled using targeted metabolomics. Associations of these metabolites with the interventions were assessed with elastic net regression analyses. Gut microbiota composition was assessed by 16S rRNA sequencing. A sparse least regression analysis combined with a canonical correlation analysis was conducted between the plasma selected metabolites and genera in order to identify the relevant dual-omics signatures discriminating the dietary interventions.

Results Changes in 65 circulating metabolites were significantly associated with the MedDiet (mainly lipids, acylcarnitines, amino acids, steroids and TCA intermediates). Importantly, these changes were associated with decreases in glucose, insulin and HOMA-IR. The network analysis identified two main clusters of genera with an opposite behaviour towards selected metabolites, mainly PC species, ChoE(20:5), TGs and medium/long-chain acylcarnitines.

Conclusion Following a MedDiet, rather than consuming nuts in the context of a non-MedDiet was associated with a specific plasma metabolomic profile, which was also related to metabolic

1 improvements in adults with MetS. The identified correlated network between specific bacteria
2 and metabolites suggests an interplay between diet, circulating metabolites and gut microbiota.
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12 **Keywords:** Mediterranean diet, nuts, insulin resistance, plasma metabolome, gut microbiota
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16 **Abbreviations:** CVD; cardiovascular disease, GC-QTOF-MS; gas chromatography time-of-
17 flight mass spectrometry, HOMA-IR; homeostasis model assessment of insulin resistance, LC-
18 Q-TOF-MS; liquid chromatography quadrupole time-of-flight mass spectrometry, LC-QqQ-
19 MS; liquid chromatography with triple-quadrupole mass spectrometry, LPC;
20 lysophosphocholine, LPE; lysophosphoethanolamine, MedDiet; Mediterranean diet, MetS;
21 metabolic syndrome, PC; phosphocholine, T2D; type-2 diabetes, TG; triglyceride, TMA;
22 trimethylamine, TMAO; trimethylamine N-oxide, TCA; tricarboxylic acid
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1 **1. Introduction**

2 Diet may contribute to the risk of chronic conditions through the modulation of several
3 biological pathways [1]. Therefore, in the era of precision nutrition, a better understanding of
4 the biological consequences following a dietary intervention is required to build a functional
5 readout of the major dietary factors related to health and disease. Circulating metabolites reflect
6 individual dietary intakes, endogenous metabolic processes and other sources of variability in
7 metabolism as the interplay with gut microbiota, which objectively allows assessing the
8 complex metabolic responses to dietary patterns. Previous dietary interventions integrating
9 nutrition with metabolomics have focused on a single nutrient or specific food [2], without
10 accounting for all interactions and failing to consider important aspects such as the substitution
11 effects of other nutrients and the synergistic or antagonistic interactions between nutrients
12 within a dietary pattern.

13 Accumulating data from epidemiological and clinical trials have demonstrated that consuming
14 a Mediterranean Diet (MedDiet) is a useful dietary strategy for the prevention of cardiovascular
15 diseases (CVD), obesity and type 2 diabetes (T2D) [3]. The cardiometabolic benefits of this
16 dietary pattern could be partially explained by its effects on insulin resistance [4], oxidative
17 stress and inflammation improvements. However, the metabolic pathways through which the
18 MedDiet exerts its beneficial effects has been little investigated. Previous randomised trials
19 have identified changes in specific metabolites after the consumption of foods typically
20 included in the MedDiet such as nuts [5]. On the contrary, few intervention studies exist on the
21 effects of the MedDiet, which is also rich in other types of foods such as vegetables, fruits,
22 legumes, whole grains, red wine and extra-virgin olive oil, on circulating metabolites [6], while
23 the relationship between diet-induced changes in metabolomic profiles and cardiometabolic
24 risk factors remains largely unknown. Furthermore, the role of gut microbiota in influencing
25 host circulating metabolites in the context of dietary interventions has not been explored yet.

1 In the present study, we examined changes in plasma metabolites after following a MedDiet
2 compared to the consumption of a single healthy food such as nuts in the context of a non-
3 MedDiet among participants with Metabolic Syndrome (MetS) and whether these changes can
4 be mediated by differences in gut microbiota composition in a multi-omics integrative
5 approach.

6 **2. Material & Methods**

7 **2.1 Study design and population**

8 METADIET is a randomized, controlled, crossover, non-blinded dietary-intervention trial
9 (**Supplemental Figure 1**) in which eligible participants were community-dwelling adults aged
10 25-60 years, with overweight/obesity (body mass index (BMI) 25-35 kg/m²) and MetS
11 according to harmonized ATPIII diagnosis criteria [7], and who regularly consumed a non-
12 MedDiet (scoring <7 in the MedDiet score used in PREDIMED-Plus trial and volunteers with
13 ≥ 7 were excluded from the trial [8]). The primary outcome of METADIET was evaluating
14 changes in gut microbiota composition after following either a MedDiet or a regular diet
15 supplemented with nuts (unpublished results), while changes in plasma metabolites and
16 cardiometabolic risk factors were secondary outcomes. The exclusion criteria were: following
17 a MedDiet considered as a punctuation ≥ 7 in the PREDIMED-Plus MedDiet score tool [8],
18 presence of T2D; several chronic diseases (inflammatory, infectious, chronic obstructive
19 pulmonary, neoplasia, endocrine, or hematological diseases), regular consumption of nuts (≥ 90
20 g/week); changes in body weight (> 5 kg in the last 3 months); non-controlled hypertension;
21 LDL cholesterol > 160 mg/dL; triglycerides > 400 mg/dL; use of anti-inflammatory, corticoids,
22 hormones or antibiotics; alcohol, smoking or drug abuse; consumption of prebiotics, probiotics
23 or laxatives. Participants were recruited from primary care centers affiliated with the University
24 Hospital of Sant Joan and through advertisements in different media. Written informed consent

1 was obtained from all study participants. The Institutional Review Board approved the study
2 protocol which accomplishes the ethical standards of the Declaration of Helsinki.

3 Participants who met the inclusion criteria were randomized, using a computer-generated
4 random-number table, either to follow a MedDiet or to consume 50 g/day of mixed nuts
5 (almonds, walnuts, hazelnuts provided free) in the context of their regular non-MedDiet for 2
6 months. After a 1-month washout period with no treatment, participants crossed over to the
7 other treatment condition for the last 2 months of the study. During the MedDiet period, trained
8 dietitians instructed participants in personalized face-to-face interviews to follow a
9 Mediterranean dietary pattern. Participants were encouraged to adhere to the 17-point scale
10 MedDiet [8] used in PREDIMED Plus study, emphasizing daily consumption of at least 2
11 servings of vegetables and 3 fruits, and weekly consumption of ≥ 3 servings of legumes, ≥ 5
12 servings of whole-grain cereals and pasta ≥ 3 servings of fish and seafood and the use of extra
13 virgin olive oil as the main culinary fat. Also, a decreased consumption of red meat and
14 processed foods to less than 1 serving/week, and reduced use of butter and margarine, white
15 bread and sweetened beverages was recommended. Participants were provided with biweekly
16 menus and seasonal recipes to facilitate adherence to the MedDiet intervention. During the nuts
17 intervention period, dietitians did not provide any other dietary advice rather than the
18 consumption of 50 g/day of mixed nuts that were provided by free, and also written culinary
19 advice to include nuts in regular meals with soups, creams or as side food. Adherence to the
20 interventions was assessed either by the validated 17-item MedDiet score [8] or counting the
21 empty nuts-packaging returned at the beginning and at the end of each intervention period, and
22 during two intermediate control visits scheduled after 15 and 30 days from the beginning of
23 each intervention period. Nutritional data was collected in each sampling visit using 3-day
24 dietary records and nutrient and energy intakes were calculated using Spanish food composition
25 tables [9,10]. Biological samples were collected at the beginning and at the end of each

1 intervention period (**Supplemental Figure 1**). Anthropometric data, blood pressure and
2 biochemical measurements were conducted following regular protocols in the clinical practice
3 and are detailed in the **Supplemental methods**.

4 **2.2 Plasma metabolomics profiling**

5 A total of 378 metabolites were quantified using a multiplatform approach including gas and
6 liquid chromatography coupled to high-resolution mass spectrometry (LC-qTOF-MS, LC-
7 QqQ-MS and GC-QTOF-MS) (**Supplemental Table 2**). The analytical procedures are
8 specified in the **Supplemental methods**. Information about the mean and SD of the metabolites
9 used in the present is shown in **Supplemental Table 2**.

10 **2.3 16S rRNA sequencing and data processing**

11 A detailed description of fecal samples collection, microbial DNA extraction and metagenomic
12 analysis can be found in the **Supplemental methods**. Briefly, microbial DNA from fecal
13 samples collected at the beginning and at the end of each intervention, was performed with
14 QIAmpPowerFecal DNA kit (Qiagen, Germany) with an additional lysing step (FastPrep-24
15 5G Homogenizer, MPBiomedicals). Hypervariable region V4 from 16S rRNA was amplified
16 using the Ion Metagenomics kitTM (Life Technology, Carlsbad, California).

17 **2.4 Statistical analyses**

18 Descriptive data of participants are presented as means and 95%CI for quantitative variables,
19 and as percentages for categorical variables. Changes in anthropometric, biochemical and
20 nutritional data were analyzed by using linear mixed-models analysis of variance with
21 intervention groups and periods modelled as fixed factors, baseline values as covariates and
22 subjects as a random effect. To account for multiple testing, we adjusted P for treatment and P
23 for treatment * period of the crude and multivariable-adjusted associations with the use of the
24 Benjamini-Hochberg false discovery rate (FDR) procedure [11]. An FDR-adjusted P-
25 value < 0.05 was considered to be statistically significant. The effect size was estimated via the

1 calculation of Cohen's, which shows the magnitude of the difference in changes in biochemical
2 and anthropometrical parameters between MedDiet and nuts interventions (i.e. Cohen's *d* value
3 of 0.2, 0.5 and 0.8 indicate small, medium and large effect size respectively) [12]. From a total
4 of 378 plasma metabolites profiled, 5 metabolites with missing values >20% were removed and
5 for those with less than 20% we used the random forest imputation approach [13] ("missForest"
6 function of "randomForest" R package version 4.6-14). Metabolites were approximated to a
7 normal distribution with the rank-based inverse normal transformation. Due to the high
8 dimensionality and collinear nature of the data, logistic regression with elastic net penalty was
9 implemented in the "glmnet" (R package, version 3.0-2) (alpha=0.5) to select the metabolites
10 associated with the dietary interventions. We performed 10-fold cross-validation (CV) to find
11 the optimal value of the tuning parameter that results in a mean squared error within 1-SD of
12 the minimum [14]. The performance of the model was examined based on parameters of
13 lambda.min. For reproducibility purposes, regression coefficients were reported using 9-10
14 iterations of the 10-fold CV elastic regression approach in the whole dataset. We also performed
15 a sensitivity analysis adjusting for energy intake. A multi-metabolite score was calculated as
16 the weighted sum of the selected metabolites with weights equal to regression coefficients from
17 the elastic net regression model. Linear regression models were fitted to examine the association
18 between the derived score and changes in glucose, insulin and HOMA-IR adjusting for age,
19 sex, changes in body weight, dietary interventions and value for the respective outcome traits
20 at the baseline examination. All analyses were performed using R statistical package 3.6.1
21 (www.r-project.org) (R Development Core Team, 2012). Metabolites selected from the elastic
22 net models were mapped into a metabolic network to identify key pathways and enrichment
23 analysis by using MetaboAnalyst 4.0.

24 Integration of metabolomics and 16S rRNA sequencing data was performed using DIABLO
25 analysis in mixOmics package of R (version 6.10.9, for which we applied a full design matrix

1 with a value of 1 to seek for linear combinations of variables from each omic dataset that are
2 maximally correlated between the two dietary interventions. Changes in relative abundances of
3 153 genera (normalized using `clr` function from the R package “compositions” package, version
4 1.40-4) and changes in the 65 selected metabolites were used. The `block.splsda` function with
5 full weighted design and 10-fold CV was used to identify the optimal number of components
6 for each omic dataset. Model performance was also evaluated using a 10-fold CV approach in
7 order to choose the best number of latent components to be included for each omic dataset. We
8 then evaluated the highly correlated omic variables which were able to discriminate the
9 MedDiet from the nuts supplementation diet. Each model was built on 80% of participants
10 (training) and the remaining 20% was used as the test set. A global overview of the correlation
11 structure at the component level was represented with the `plotDiablo` function. A circosplot
12 representing the correlations between variables of different types represented on the side
13 quadrants was obtained with the `circosplot` function. Also, a network analysis was implemented
14 by `network` function, in order to display the association network for regularized canonical
15 correlation analysis and sparse partial least square regression.

16 **3. Results**

17 Of the 50 participants initially included in the study, 6 dropped out for personal reasons
18 resulting in 44 participants with available metabolomic data (**Figure 1**). No significant
19 differences in participants’ baseline characteristics were observed between the two
20 interventions (**Table 1**). Medium effect sizes (Cohen’s $d = 0.5$) were observed for glucose,
21 insulin and HOMA-IR, whereas large effect size (Cohen’s $d > 1$) was observed for the MedDiet
22 score. The mean \pm SD for the 17-items MedDiet score was 11.59 \pm 2.41 [increase; 4.61 (3.74,
23 5.48)] after the MedDiet and 9.30 \pm 3.54 [increase; 0.36 (-0.82, 1.55)] after the nuts
24 supplemented diet. Significant decreases in the levels of glucose, insulin and the values of
25 HOMA-IR were observed in the MedDiet compared to the nuts supplementation diet (**Table**

1 **1).** Significant differences in total lipids, saturated fatty acids (SFA), and protein intake were
2 observed between interventions. (**Supplemental Table 3**). A higher increase of fruits, legumes,
3 fish and a decrease of alcohol consumption was found after the MedDiet compared to the
4 regular diet supplemented with nuts, whereas nuts consumption was significantly increased in
5 the nuts intervention group (**Supplemental Table 3**).

6 **Table 2** shows 65 selected metabolites ranked from the highest to the lowest elastic net positive
7 and negative regression coefficients for the MedDiet. Thirty-five metabolites were positively
8 associated with the MedDiet; hydroxyperoxide-eicosapentanoic acid (HpEPE), testosterone,
9 phosphatidylcholines (PC) (40:6, 35:1), trimethylamine (TMA), succinic acid, cholesterol
10 esters (ChoE) (17:0, 20:5), tauro lithocholic acid, amino acids (threonine, cystathione, histidine,
11 phenylalanine), lysophosphatidylcholines (LPC) (19:0-sn1, 22:6, 16:1), carnitine species (C5-
12 OH, C5:1, C18:2, C2:0, C16:0-OH), triglycerides (TG) (56:6, 56:7, 56:5) and
13 lysophosphoethanolamine (LPE) 20:5-sn1. Among the 29 metabolites negatively associated
14 with the MedDiet, the highest regression coefficients were found for LPE18:3-sn1 followed by
15 sphingomyelin (SM) (36:0, 41:1, 32:2), 9-OxoODE, carnitines (C12:0, C5-M-DC, C12:0-OH-
16 a), taurine and its bile acid, taurocholic acid, linolenic acid-iso2 and -iso1,
17 dehydroepiandrosterone sulfate, androsterone sulfate-iso4, androsterone sulfate-iso2, PCs
18 (33:2, 34:1e, 32:0, 34:2, 32:2), TG 47:0, ChoE 22:5, dihomo- γ -linolenic acid-iso2 and
19 hydroxyoctadecadienoic acid (HODE)-iso1, hydroperoxyoctadecadienoic acid (HpODE).

20 In order to take into account the significant difference in energy intake, we have conducted a
21 sensitivity analysis which shown that 29 metabolites of the 65 previously selected from the
22 unadjusted model were associated to the dietary interventions (**Supplemental Table 4**). The
23 enrichment analysis of the selected metabolites revealed that most of them were involved in
24 amino acids metabolism. The metabolic pathways with the highest impact were related to
25 taurine, glycine, serine and threonine metabolism, phenylalanine and tryptophan biosynthesis,

1 together with tricarboxylic acid (TCA). A moderate impact was also found for the metabolism
2 of alpha-linolenic acid, glycerophospholipid and unsaturated fatty acid (**Figure 2**). The linear
3 regression models revealed significant inverse associations of the multi-metabolite score with
4 changes in glucose, insulin and HOMA-IR (**Table 3**).

5 In **Figure 3** we plotted the multi-omic analysis including the selected metabolites from the
6 elastic net regression and the 153 genera depicting gut microbial composition. This analysis
7 includes a total of 38 subjects with available data after filtering procedures in gut microbiota
8 (**Supplemental Figure 1**). No significant differences in the general characteristics of
9 participants included in both analyses were observed (**Supplemental Table 1**). Considering a
10 correlation cut-off value of 0.2 we were able to find two main clusters of genera differentially
11 correlated with a set of metabolites. The first cluster consisted of an uncultured genus of
12 Lachnospiraceae, *Ruminococcaceae* UCG002, *Lachnoclostridium* and some genera from
13 Prevotellaceae family which were positively correlated with changes of C16-OH, C12:0, C12-
14 OH, PC35:1, PC40:6, and TGs 56:6, 46:7, 56:5 and also with ChoE 20:5. On the contrary,
15 negative correlations with changes in phosphoethanolamine and taurine were found for this
16 cluster. The second cluster, constituted by an uncultured genus from Christensenellaceae
17 family, *Oxalobacter*, Clostridiales family XII, *Ruminococacceae* UCG009, *Terrisporobacter*,
18 and a genus from Clostridiales family, presented the same associations of the first cluster but
19 with an opposite sign.

20 **4. Discussion**

21
22 In the present study, we identified changes in several plasma metabolites associated with
23 following either a MedDiet or a non-MedDiet supplemented with nuts in participants with
24 overweight/obesity and MetS. These metabolites mainly included lipid species and
25 acylcarnitines but also amino acids, steroids and TCA intermediates. Importantly, metabolites
26 changes induced by the MedDiet, compared to a single healthy food like nuts in the context of

1 a regular diet, were associated with improvements in participants' metabolic risk profile,
2 independent of changes in body weight and dietary interventions indicating their promising use
3 in understanding the biological mechanisms behind the effects of the MedDiet.

4 The effect of the MedDiet on circulating metabolites has been poorly investigated. In a case-
5 cohort study nested within the Prevention with Mediterranean Diet (PREDIMED) trial,
6 participants following a MedDiet had a 1-year significant reduction in several lipid species,
7 mainly PCs, PEs, ChoE and TGs, compared to participants in the low-fat diet control group,
8 although only changes in ChoE remained statistically significant after correcting for multiple
9 comparisons [15]. Similarly, in the Metabolic Syndrome Reduction in Navarra (RESMENA)
10 trial conducted among 72 subjects with obesity and at least two features of MetS, who received
11 an energy-restricted MedDiet or a low-fat diet, displayed significant changes in several lipid
12 species, mainly PCs, LPCs and SMs after 2 months [16]. A more recent cross-sectional study
13 that identified a metabolic profile of adherence to the MedDiet revealed that out of 67
14 metabolites, 45 were lipids [17]. Accordingly, in our study, following the MedDiet compared
15 with non-MedDiet supplemented with nuts was associated with decreases in PCs and SMs.
16 Previous observational studies have suggested a relationship between elevated levels of PC and
17 increased risk of coronary artery disease and mortality [18], but in the PREDIMED study
18 polyunsaturated PCs with at least 5 double bonds conferred CVD protection [19]. PCs are direct
19 substrates for the formation of SMs by SM-synthase [20], which are hydrolyzed by
20 sphingomyelinases activated by inflammatory cytokines and oxidative stress, to produce
21 ceramides [21]. Whether increases in PC40:6 induced by the MedDiet are related to metabolic
22 benefits requires further investigation. ChoE and TGs with >56 carbon atoms and >3 double
23 bonds have been inversely associated with T2D [22] and in our study were increased after the
24 MedDiet. These lipids could represent a pathway through which the MedDiet may have
25 favorably affected insulin resistance.

1 The increase in plasma α -tocopherol after the end of the MedDiet suggests a good adherence to
2 this dietary pattern [23]. Furthermore, given that the MedDiet and nuts are high in unsaturated
3 fats, it is not surprising that a large proportion of lipid species consist of PUFAs [17]. Therefore,
4 the increases in plasma LPE20:5 and LPC22:6 after the MedDiet may reflect the trend to a
5 higher intake of fish, since plasma EPA and DHA has been identified as the most suitable
6 biomarker of acute changes in 20:5(n-3) and 22:6(n-3) fatty acids intake [24]. Furthermore, an
7 EPA-derived lipoxygenase metabolite, HpEPE, was highly associated with the MedDiet. Our
8 results are in line with a previous study in which higher fish intake was associated with higher
9 levels of highly unsaturated lipid metabolites containing 22:6 and 20:5 (n-3) [17]. An
10 involvement of the metabolism of lysophospholipids in cardiometabolic health has been
11 suggested, although, findings from recent lipidomic studies of LPC have been controversial
12 [25,26]. In a previous *in vitro* study, HpEPE was found to inhibit platelet aggregation [27]. On
13 the contrary, a positive association with the nuts supplemented diet was found for α -linolenic
14 acid (ALA), which could also explain the increases in LPE18:3 and LPC18:3 after following
15 this intervention. ALA is an essential n-3 fatty acid, mainly deriving from the diet, especially
16 from nuts that has been related to cardiovascular health benefits [28]. Metabolites produced by
17 the linoleic acid metabolism such as 9-OxoODE, 13-HpODE and its catabolic product 13-
18 HODE increased after the nuts intervention, whereas 15-HETE produced by arachidonic acid
19 increased after the MedDiet. Further studies are needed to understand the role of these
20 metabolites on inflammation regulation.

21 Changes in different acylcarnitines were found to be associated with the dietary interventions
22 indicating a different role of these dietary patterns on the fatty-acids β -oxidation and energy
23 metabolism [29]. Two long-chain and three short-chain acylcarnitines were associated with the
24 MedDiet, while one short- and two medium-chain acylcarnitines were associated with the nuts
25 intervention. Previous studies have suggested that pathways related to acylcarnitine metabolism

1 can be influenced by diet. In a previous cross-sectional study, a metabolite profile composed of
2 medium- and long-chain acylcarnitines was inversely associated with the Western dietary
3 pattern [30]. On the other hand, long-chain acylcarnitines were associated with veganism in the
4 EPIC-Oxford cohort [31]. In the PREDIMED study, high short- and medium-chain
5 acylcarnitines levels were associated with a higher risk of CVD [32], while short- and long-
6 chain acylcarnitines with higher T2D risk [33].

7 Besides the impact of the study dietary interventions on lipid metabolism and fatty acid
8 oxidation, changes in amino acid metabolism were also observed in our study. Following the
9 MedDiet was associated with increases in plasma amino acids such as phenylalanine, histidine,
10 cystathionine and threonine. Previous studies have suggested that a Western dietary pattern or
11 an omnivorous diet are associated with increased levels of amino acids like phenylalanine
12 [30,34]. This could be due to the higher content of phenylalanine in animal proteins. In our
13 study, participants assigned to the MedDiet intervention may have increased circulating amino
14 acids concentrations possibly due to the trend for a higher fish consumption as compared to the
15 nuts group. On the other hand, the sulfur-containing amino acid, cystine, derivative of cysteine
16 mainly found in almonds [35] was associated with the nuts intervention group.

17 Increased plasma concentrations of TCA-related metabolites (i.e., succinate) after the MedDiet
18 compared to nuts group is a novel finding. Recent evidence suggests that succinate is not only
19 a TCA intermediary metabolite but also a by-product of gut microbial carbohydrate and amino
20 acid fermentation [36] with pleiotropic functions, such as resolution of inflammation associated
21 with obesity [37]. Regarding other gut microbiota-derived metabolites, increases in TMA
22 concentrations were found after the MedDiet possibly due to the higher fish consumption
23 observed following this dietary pattern than the non-MedDiet supplemented with nuts. TMA
24 can be generated in the colon from choline or L-carnitine or metabolic retro-conversion of
25 trimethylamine N-oxide (TMAO) to TMA by the gut microbiota [38]. In a study with 40 healthy

1 young men, consumption of meals producing TMAO provided directly from fish led to a
2 significant increase in postprandial plasma TMA levels. However, eggs or beef consumption,
3 providing choline and L-carnitine, respectively, resulted only in negligible TMA increases [39].

4 At the same time, the MedDiet was associated with increases in tauro lithocholic acid, while the
5 regular diet plus nuts with increases in taurine and its conjugated bile acid (taurocholic acid).
6 Recent animal studies showed an increase in taurocholic acid in mice fed with a high-saturated
7 fat diet [40], which may result in microbiota dysbiosis that can perturb immune homeostasis.
8 Similarly, in our study, the higher intake of saturated fatty acids in the nuts group may explain
9 these findings. In addition, previous *in vitro* studies have demonstrated that treatment with
10 tauro lithocholic acid significantly increased nitric oxide production [41], while taurocholic acid
11 exerted an arrhythmogenic effect [42].

12 Another novel finding of our study was increased in testosterone after MedDiet and increases
13 in three androsterone sulfate species following the nuts supplemented diet. Whether both diets
14 provide sufficient nutrients for precursors of steroidogenesis [43] is a hypothesis that needs to
15 be examined. Interestingly, obese men with low testosterone concentrations are more prone to
16 metabolic disturbances such as insulin resistance [44]. A recent *in vivo* study also found a
17 relationship between lower plasma concentrations of sulfated steroids, such as
18 dehydroepiandrosterone sulfate and impaired glucose tolerance [45].

19 The dual-omic analysis demonstrated a correlation between a cluster enriched by medium-and
20 long-chain acylcarnitines, PCs, ChoE and TGs with >56 carbon atoms and >3 double bonds
21 and a cluster of bacteria belonging to Firmicutes (Lachnospiraceae, *Lachnoclostridium* and
22 Ruminococcaceae) and genera from Prevotellaceae family. Genera from the Lachnospiraceae
23 family were associated to a higher MedDiet adherence [46] whereas higher abundances of
24 *Lachnoclostridium* were reported after a 3 months MedDiet intervention [47]. Similarly, plant-
25 based foods diets were associated with a Prevotella enterotype [48]. On the other hand, the

1 second cluster, constituted of *Oxalobacter* and genera from Christensenellaceae family together
2 with *Ruminococcaceae* UCG009 and genera from Clostridiales family may reflect the effect of
3 the nuts supplemented diet on these microbes. Nuts are rich in oxalate, the main substrate for
4 *Oxalobacter formigenes*, and their consumption has been related with a higher concentration of
5 gastricoxalate [49]. Christensenellaceae is reported higher in relative abundance in humans with
6 an omnivorous diet, relative to vegetarians [46]. Potential mechanisms through which the
7 identified microbiota may affect circulating acylcarnitines, PCs, TGs and ChoE may involve
8 short-chain fatty acids (SCFAs). Some of the genera (uncultured Lachnospiraceae,
9 *Lachnoclostridium*, *Ruminococcaceae* UCG002, uncultured Prevotellaceae and
10 Christensenellaceae) identified in our study are involved in production of SCFAs, which can
11 be absorbed into the bloodstream and affect lipid metabolism [50].

12 Our study has several strengths such the crossover, randomized, controlled design, thus
13 balancing the potential intra-individual variability in metabolites and gut microbiota with a
14 longer washout period to avoid a potential carryover effect, even if we cannot completely
15 discard due to the crossover design of the study. With regards to limitations, the relatively small
16 sample size and the high intra-variability of gut microbiota composition could limit to reach
17 statistically significance for some genera. Also, the lack of blinding, despite its unfeasibility,
18 represents a potential bias that should be taken into account. Furthermore, the 16S rRNA
19 analysis does not allow to identify bacterial species, leaving the taxonomy at genus level.
20 Finally, diablo analysis is one of the computational methods for inferring correlation networks
21 which is computationally challenging since sensitivity, specificity and precision have not been
22 evaluated in reference to either real or theoretical data sets and these limitations should be
23 accounted when interpreting our findings.

24 **5. Conclusion**

1 In conclusion, our study demonstrated that following a MedDiet, rather than the consumption
2 of a non-MedDiet supplemented with nuts, was associated with changes in the plasma
3 metabolome that were associated with insulin resistance improvements among participants with
4 MetS. Medium- and long-chain acylcarnitines, two PCs, and TGs and ChoE with >56 carbon
5 atoms and >3 double bonds appeared to be related with a specific gut microbiota composition
6 supporting an interplay between diet, circulating metabolites and gut microbiota.

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9 Centre for Omics Sciences (COS) Joint Unit of Universitat Rovira i Virgili-Eurecat. Data
10 generated and analysed in the framework of the METADIET are not publicly available due to
11 national data regulations and for ethical reasons, because study participants only gave their
12 consent for the use of their data by the METADIET investigators. However, collaboration for
13 data analyses can be requested by sending a letter to the corresponding author (Mònica Bulló).

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25 **Author Contribution**

1 MB designed the research and was the coordinator of subject's recruitment; MB, SG, JG-G,
2 LC-B, PA, AP-G, AR recruited participants and conducted the research, SG, JG-G and LC-B
3 obtained data and conducted the bioinformatics analysis; SG, JG-G and CP conducted the
4 statistical analysis; SG, CP, MB, drafted the paper; all the authors revised the manuscript for
5 important intellectual content, and read and approved the final version.

6 **Conflict of interest statement**

7 The authors declare that they have no competing interests.

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References

1. Schulze MB, Martínez-González MA, Fung TT, Lichtenstein AH, Forouhi NG. Food based dietary patterns and chronic disease prevention. *BMJ*. 2018;361:1–6.
2. Tuccinardi D, Farr OM, Upadhyay J, Oussaada SM, Klapa MI, Candela M, et al. Mechanisms underlying the cardiometabolic protective effect of walnut consumption in obese people: A cross-over, randomized, double-blind, controlled inpatient physiology study. *Diabetes, Obes Metab*. 2019;21(9):2086–95.
3. Papadaki A, Nolen-Doerr E, Mantzoros CS. The effect of the mediterranean diet on metabolic health: A systematic review and meta-analysis of controlled trials in adults. *Nutrients*. 2020;12(11):1–21.
4. Mirabelli M, Chiefari E, Arcidiacono B, Corigliano DM, Brunetti FS, Maggisano V, et al. Mediterranean diet nutrients to turn the tide against insulin resistance and related diseases. *Nutrients*. 2020.
5. Malik VS, Guasch-Ferre M, Hu FB, Townsend MK, Zeleznik OA, Eliassen AH, et al. Identification of plasma lipid metabolites associated with nut consumption in US men and women. *J Nutr*. 2019;149(7):1215–21.
6. Meslier V, Laiola M, Roager HM, De Filippis F, Roume H, Quinquis B, et al. Mediterranean diet intervention in overweight and obese subjects lowers plasma cholesterol and causes changes in the gut microbiome and metabolome independently of energy intake. *Gut*. 2020;69(7):1258–68.
7. Alberti KGMM, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, et al. Harmonizing the metabolic syndrome: A joint interim statement of the international diabetes federation task force on epidemiology and prevention; National heart, lung, and blood institute; American heart association; World heart federation; International . Vol. 120, *Circulation*. 2009. p. 1640–5.

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51
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56
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58
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64
65
8. Martínez-González MA, Buil-Cosiales P, Corella D, Bulló M, Fitó M, Vioque J, et al. Cohort profile: Design and methods of the PREDIMED-Plus randomized trial. *Int J Epidemiol.* 2019;48(2):387-388o.
9. Moreiras O, Cabrera L. Tablas de composición de alimentos [Food composition tables]. 5th ed. Granada: Universidad de Granada; 2005.
10. Moreiras, O.; Cabrera, L.; Cuadrado C. Tablas de Composición de Alimentos. (Spanish Food Composition Tables). 17th ed. madrid: Pirámide; 2015. 16 p.
11. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *J R Stat Soc Ser B.* 1995;
12. Cohen J. Statistical power: analysis for the behavioural sciences. New York: Academic Press. *Statistical Power Analysis for the Behavioural Science.* 1977.
13. Gromski P, Xu Y, Kotze H, Correa E, Ellis D, Armitage E, et al. Influence of Missing Values Substitutes on Multivariate Analysis of Metabolomics Data. *Metabolites.* 2014;4(2):433–52.
14. Friedman J, Hastie T, Tibshirani R. Regularization paths for generalized linear models via coordinate descent. *J Stat Softw.* 2010;33(1):1–22.
15. Toledo E, Wang DD, Ruiz-Canela M, Clish CB, Razquin C, Zheng Y, et al. Plasma lipidomic profiles and cardiovascular events in a randomized intervention trial with the Mediterranean diet. *Am J Clin Nutr.* 2017;106(4):973–83.
16. Bondia-Pons I, Martinez JA, de la Iglesia R, Lopez-Legarrea P, Poutanen K, Hanhineva K, et al. Effects of short- and long-term Mediterranean-based dietary treatment on plasma LC-QTOF/MS metabolic profiling of subjects with metabolic syndrome features: The Metabolic Syndrome Reduction in Navarra (RESMENA) randomized controlled trial. *Mol Nutr Food Res.* 2015;59(4):711–28.
17. Li J, Guasch-Ferré M, Chung W, Ruiz-Canela M, Toledo E, Corella D, et al. The

- Mediterranean diet, plasma metabolome, and cardiovascular disease risk. *Eur Heart J*. 2020;41(28):2645–56.
18. Schlitt A, Blankenberg S, Yan D, Von Gizycki H, Buerke M, Werdan K, et al. Further evaluation of plasma sphingomyelin levels as a risk factor for coronary artery disease. *Nutr Metab*. 2006;3.
 19. Razquin C, Liang L, Toledo E, Clish CB, Ruiz-Canela M, Zheng Y, et al. Plasma lipidome patterns associated with cardiovascular risk in the PREDIMED trial: A case-cohort study. *Int J Cardiol*. 2018;253:126–32.
 20. Li Z, Vance DE. Phosphatidylcholine and choline homeostasis. Vol. 49, *Journal of Lipid Research*. 2008. p. 1187–94.
 21. Cutler RG, Kelly J, Storie K, Pedersen WA, Tammara A, Hatanpaa K, et al. Involvement of oxidative stress-induced abnormalities in ceramide and cholesterol metabolism in brain aging and Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2004;
 22. Papandreou C, Bulló M, Zheng Y, Ruiz-Canela M, Yu E, Guasch-Ferré M, et al. Plasma trimethylamine-N-oxide and related metabolites are associated with type 2 diabetes risk in the Prevención con Dieta Mediterránea (PREDIMED) trial. *Am J Clin Nutr*. 2018;108(1):163–73.
 23. Bach A, Serra-Majem L, Carrasco JL, Roman B, Ngo J, Bertomeu I, et al. The use of indexes evaluating the adherence to the Mediterranean diet in epidemiological studies: a review. *Public Health Nutr*. 2006;
 24. Browning LM, Walker CG, Mander AP, West AL, Madden J, Gambell JM, et al. Incorporation of eicosapentaenoic and docosahexaenoic acids into lipid pools when given as supplements providing doses equivalent to typical intakes of oily fish¹⁻⁴. *Am J Clin Nutr*. 2012;96(4):748–58.
 25. Law SH, Chan ML, Marathe GK, Parveen F, Chen CH, Ke LY. An updated review of

lysophosphatidylcholine metabolism in human diseases. *International Journal of Molecular Sciences*. 2019.

26. Papandreou C, Bulló M, Ruiz-Canela M, Dennis C, Deik A, Wang D, et al. Plasma metabolites predict both insulin resistance and incident type 2 diabetes: A metabolomics approach within the Prevención con Dieta Mediterránea (PREDIMED) study. *Am J Clin Nutr*. 2019;109(3):635–47.
27. Takenaga M, Hirai A, Terano T, Tamura Y, Kitagawa H, Yoshida S. Comparison of the in vitro effect of eicosapentaenoic acid (EPA)-derived lipoxygenase metabolites on human platelet function with those of arachidonic acid. *Thromb Res*. 1986;41(3):373–84.
28. Lázaro I, Rueda F, Cediel G, Ortega E, García-García C, Sala-Vila A, et al. Circulating Omega-3 Fatty Acids and Incident Adverse Events in Patients With Acute Myocardial Infarction. *J Am Coll Cardiol*. 2020;76(18):2089–97.
29. Longo N, Frigeni M, Pasquali M. Carnitine transport and fatty acid oxidation. *Biochim Biophys Acta - Mol Cell Res*. 2016;1863(10):2422–35.
30. Bouchard-Mercier A, Rudkowska I, Lemieux S, Couture P, Vohl MC. The metabolic signature associated with the Western dietary pattern: A cross-sectional study. *Nutr J*. 2013;12(1).
31. Schmidt JA, Rinaldi S, Ferrari P, Carayol M, Achaintre D, Scalbert A, et al. Metabolic profiles of male meat eaters, fish eaters, vegetarians, and vegans from the EPIC-Oxford cohort. *Am J Clin Nutr*. 2015;102(6):1518–26.
32. Guasch-Ferré M, Zheng Y, Ruiz-Canela M, Hruby A, Martínez-González MA, Clish CB, et al. Plasma acylcarnitines and risk of cardiovascular disease: Effect of Mediterranean diet interventions. *Am J Clin Nutr*. 2016;103(6):1408–16.
33. Guasch-Ferré M, Ruiz-Canela M, Li J, Zheng Y, Bulló M, Wang DD, et al. Plasma

- acylcarnitines and risk of type 2 diabetes in a mediterranean population at high cardiovascular risk. *J Clin Endocrinol Metab.* 2019;104(5):1508–19.
- 1
2
3
4
5 34. Xu J, Yang S, Cai S, Dong J, Li X, Chen Z. Identification of biochemical changes in
6
7 lactovegetarian urine using ¹H NMR spectroscopy and pattern recognition. *Anal*
8
9 *Bioanal Chem.* 2010;396(4):1451–63.
10
11
12 35. Venkatachalan M, Sathe SK. Chemical composition of selected edible nut seeds. *J*
13
14 *Agric Food Chem.* 2006;
15
16
17 36. Fernández-Veledo S, Vendrell J. Gut microbiota-derived succinate: Friend or foe in
18
19 human metabolic diseases? *Rev Endocr Metab Disord.* 2019;20(4):439–47.
20
21
22 37. Keiran N, Ceperuelo-Mallafre V, Calvo E, Hernández-Alvarez MI, Ejarque M, Núñez-
23
24 Roa C, et al. SUCNR1 controls an anti-inflammatory program in macrophages to
25
26 regulate the metabolic response to obesity. *Nat Immunol.* 2019;20(5):581–92.
27
28
29 38. Papandreou C, Moré M, Bellamine A. Trimethylamine n-oxide in relation to
30
31 cardiometabolic health—cause or effect? Vol. 12, *Nutrients.* 2020.
32
33
34 39. Cho CE, Taesuwan S, Malysheva O V., Bender E, Tulchinsky NF, Yan J, et al.
35
36 Trimethylamine-N-oxide (TMAO) response to animal source foods varies among
37
38 healthy young men and is influenced by their gut microbiota composition: A
39
40 randomized controlled trial. *Mol Nutr Food Res.* 2017;61(1).
41
42
43 40. Rinninella E, Cintoni M, Raoul P, Lopetuso LR, Scaldaferri F, Pulcini G, et al. Food
44
45 components and dietary habits: Keys for a healthy gut microbiota composition. Vol.
46
47 11, *Nutrients.* 2019.
48
49
50
51 41. Devkota S, Wang Y, Musch MW, Leone V, Fehlner-Peach H, Nadimpalli A, et al.
52
53 Dietary-fat-induced taurocholic acid promotes pathobiont expansion and colitis in Il10-
54
55 */-* mice. Vol. 487, *Nature.* 2012. p. 104–8.
56
57
58 42. Sheikh Abdul Kadir SH, Miragoli M, Abu-Hayyeh S, Moshkov A V., Xie Q, Keitel V,
59
60
61
62
63
64
65

et al. Bile acid-induced arrhythmia is mediated by muscarinic M2 receptors in neonatal rat cardiomyocytes. *PLoS One*. 2010;5(3).

43. Salas-Huetos A, Bulló M, Salas-Salvadó J. Dietary patterns, foods and nutrients in male fertility parameters and fecundability: A systematic review of observational studies. *Hum Reprod Update*. 2017;23(4):371–89.
44. Niskanen L, Laaksonen DE, Punnonen K, Mustajoki P, Kaukua J, Rissanen A. Changes in sex hormone-binding globulin and testosterone during weight loss and weight maintenance in abdominally obese men with the metabolic syndrome. *Diabetes, Obes Metab*. 2004;6(3):208–15.
45. Ma J, Yue J, Huang R, Liao Y, Li S, Liu W. Reversion of aging-related DHEAS decline in mouse plasma alleviates aging-related glucose tolerance impairment by potentiation of glucose-stimulated insulin secretion of acute phase. *Biochem Biophys Res Commun*. 2018;500(3):671–5.
46. De Filippis F, Pellegrini N, Vannini L, Jeffery IB, La Stora A, Laghi L, et al. High-level adherence to a Mediterranean diet beneficially impacts the gut microbiota and associated metabolome. *Gut*. 2016;65(11).
47. Pagliai G, Russo E, Niccolai E, Dinu M, Di Pilato V, Magrini A, et al. Influence of a 3-month low-calorie Mediterranean diet compared to the vegetarian diet on human gut microbiota and SCFA: the CARDIVEG Study. *Eur J Nutr*. 2020;59(5):2011–24.
48. Wu GD, Chen J, Hoffmann C, Bittinger K, Chen YY, Keilbaugh SA, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science* (80-). 2011;334(6052):105–8.
49. Ritter MMC, Savage GP. Soluble and insoluble oxalate content of nuts. *J Food Compos Anal*. 2007;20(3–4):169–74.
50. Chambers ES, Preston T, Frost G, Morrison DJ. Role of Gut Microbiota-Generated

Short-Chain Fatty Acids in Metabolic and Cardiovascular Health. Vol. 7, Current
Nutrition Reports. 2018. p. 198–206.

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Table 1. Baseline and changes in biochemical and anthropometrical parameters (n=44)

Characteristics	Mediterranean Diet		Nuts		MedDiet vs Nuts (changes)		
	Baseline	Change	Baseline	Change	P _{treat₁}	P _{treat*period¹}	Effect size Cohen's <i>d</i>
Age	50.73 (48.65, 52.80)	0 (0, 0)	50.73 (48.65, 52.80)	0 (0, 0)	N/A	N/A	
Weight (Kg)	84.81 (83.01; 86.61)	-0.7 (-1.21, -0.19)	84.43 (82.64; 86.22)	-0.02 (-0.47, 0.42)	0.156	0.580	
Waist Circumference (cm)	102.79 (101.49; 104.10)	-1.1 (-2.22, 0.02)	101.77 (100.35; 103.19)	0.74 (-1.93, 3.41)	0.617	0.726	
SBP (mmHg)	134.85 (133.14; 136.56)	-1.4 (-5.14, 2.35)	132.48 (130.74; 134.21)	-1.73 (-6.61, 3.16)	0.894	0.580	
DBP (mmHg)	85.26 (84.13; 86.39)	-0.99 (-2.99, 1.01)	84.35 (82.96; 85.75)	-2.17 (-5.53, 1.19)	0.617	0.639	
Total Cholesterol (mg/dL)	215.09 (210.09; 220.09)	-7.82 (-14.99, -0.64)	212.41 (207.10; 217.72)	-0.16 (-9.04, 8.73)	0.189	0.639	
LDLc (mg/dl)	136.93 (132.86; 141.00)	-6.66 (-13.33, 0.02)	133.27 (128.95; 137.60)	-2.65 (-8.31, 3.01)	0.474	0.381	
HDLc (mg/dL)	50.48 (48.71; 52.24)	-1.23 (-3.13, 0.68)	50.61 (48.84; 52.39)	0.02 (-1.56, 1.61)	0.565	0.793	
VLDLc (mg/dL)	27.68 (25.77; 29.59)	0.07 (-2.47, 2.61)	28.52 (26.91; 30.14)	-1.44 (-4.55, 1.67)	0.617	0.681	
Triglycerides (mg/dL)	138.66 (129.12; 148.20)	0.07 (-12.44, 12.58)	142.84 (134.75; 150.94)	13.25 (-29.51, 56.01)	0.617	0.681	
Glucose (mg/dL)	100.55 (98.61; 102.48)	-3.18 (-6.53, 0.17)	97.45 (95.77; 99.14)	2.25 (-0.47, 4.97)	0.046	0.580	0.53 (0.10, 0.96)
Insulin (mcUI/mL)	14.46 (13.33; 15.58)	-1.44 (-3.1, 0.21)	12.44 (11.33; 13.54)	2.35 (0.03, 4.66)	0.046	0.210	0.56 (0.12, 0.99)
HOMA-IR	3.65 (3.32; 3.98)	-0.42 (-0.96, 0.12)	1.01 (2.73; 3.29)	0.8 (0.03, 1.56)	0.046	0.210	0.55 (0.11, 0.97)
Zonulin (ng/mL)	40.08 (38.93; 41.23)	0.52 (-1.67, 2.7)	40.45 (39.51; 41.40)	1.04 (-1.04, 3.11)	0.770	0.830	
Items MedDiet score	6.98 (6.61; 7.35)	4.61 (3.74, 5.48)	8.93 (8.46; 9.40)	0.36 (-0.82, 1.55)	0.001	0.381	-1.22 (-1.53, -0.89)

All values are given as means (95% CI). Changes in anthropometric and biochemical parameters between dietary interventions were analyzed analyzed by using linear mixed-effects analysis of variance with intervention groups and periods modelled as fixed factors, baseline values as covariates and subjects as random effect. Cohen's *d* indicates the effect size of MedDiet vs. Nuts group. Abbreviations: SBP; systolic blood Pressure, DBP; diastolic blood pressure, LDLc; low density lipoprotein cholesterol, HDLc; high density lipoprotein cholesterol, VLDLc; very low-density lipoprotein cholesterol, HOMA-IR; homeostatic model assessment of insulin resistance, MedDiet; Mediterranean Diet, N/A; not available.

¹Adjusted with the Benjamini-Hochberg False Discovery Rate method.

Table 2. Plasma metabolites ranked from the highest to the lowest elastic net positive or negative regression coefficients for the MedDiet intervention.

Metabolites	Coefficient	Metabolites	Coefficient
HpEPE	0.958	LPE18:3-sn1	-0.649
Testosterone	0.579	SM36:0	-0.563
PC40:6	0.537	9-OxoODE	-0.517
TMA	0.518	C12:0	-0.334
Succinic acid	0.407	C5-M-DC	-0.320
ChoE(17:0)	0.388	Taurine	-0.284
Taurolithocholic acid	0.380	Linolenic acid-iso2	-0.283
Threonine	0.356	SM41:1	-0.282
LPC19:0-sn1	0.306	Dehydroepiandrosterone sulfate	-0.281
C5-OH	0.304	Hydroxyproline trans	-0.236
3-Phosphoglyceric acid	0.291	PC33:2	-0.235
LPC 22:6	0.278	Linolenic acid-iso1	-0.229
Cystathione	0.273	androsterone sulfate-iso4	-0.222
Histidine	0.231	PC34:1 e	-0.218
C5:1	0.219	Taurocholic acid	-0.215
C18:2	0.215	PC32:0	-0.213
Glycoursodeoxycholic acid	0.205	TG47:0	-0.198
Phenylalanine	0.199	ChoE(22:5)	-0.191
C2:0	0.197	PC34:2	-0.188
Glycerol-1-phosphate	0.172	dihomo- γ -linolenic acid-iso2	-0.188
TG56:6	0.166	androsterone sulfate-iso2	-0.178
PC38:2	0.161	C12_0-OH-a	-0.172
alpha-tocopherol	0.145	HODE-iso1	-0.140
TG56:7	0.139	HpODE	-0.134
15-HETE	0.131	Cystine	-0.112
ChoE(20:5)	0.128	9.12.13-TriHOME	-0.109
LPE20:5-sn1	0.095	SM32:2	-0.106
Fumaric acid	0.089	Phosphoethanolamine	-0.096
3-Hydroxybutyric acid	0.088	LPC18:3-sn1	-0.078
PC35:1	0.088	PC32:2	-0.059
Nervonic acid	0.075		
C16:0-OH	0.058		
cis-10 heptadecenoic acid	0.041		
LPC16:1	0.026		
TG56:5	0.025		

Abbreviations: ChoE: cholesterol ester; HETE: hydroxyeicosatetraenoic acid; HpEPE: hydroxyperoxy-eicosapentaenoic acid; HpODE: hydroperoxyoctadecadienoic acid; LPC: lysophosphatidylcholine; LPE: lysophosphoethanolamine; OxoODE: oxo-octadecadienoic acid; PC: phosphatidylcholine; SM: sphingomyelin; TG: triglycerides; TMA: trimethylamine-N-oxide; TriHOME: trihydroxyoctadecenoic acid

Table 3. Linear regression analysis examining the associations of 1-SD increase of multi-metabolite score with changes of main cardio-metabolic parameters.

Changes	(Mean±SE)	p value
Glucose (mg/dL)	-0.72± 0.34	0.020
Insulin (mcUI/mL)	-0.57± 0.24	0.010
HOMA-IR	-0.56 ± 0.23	0.010

Values are given as means ± standard error. Model was adjusted for sex, age, weight changes, dietary interventions and value for the respective outcome traits at the baseline examination.

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Figure legend

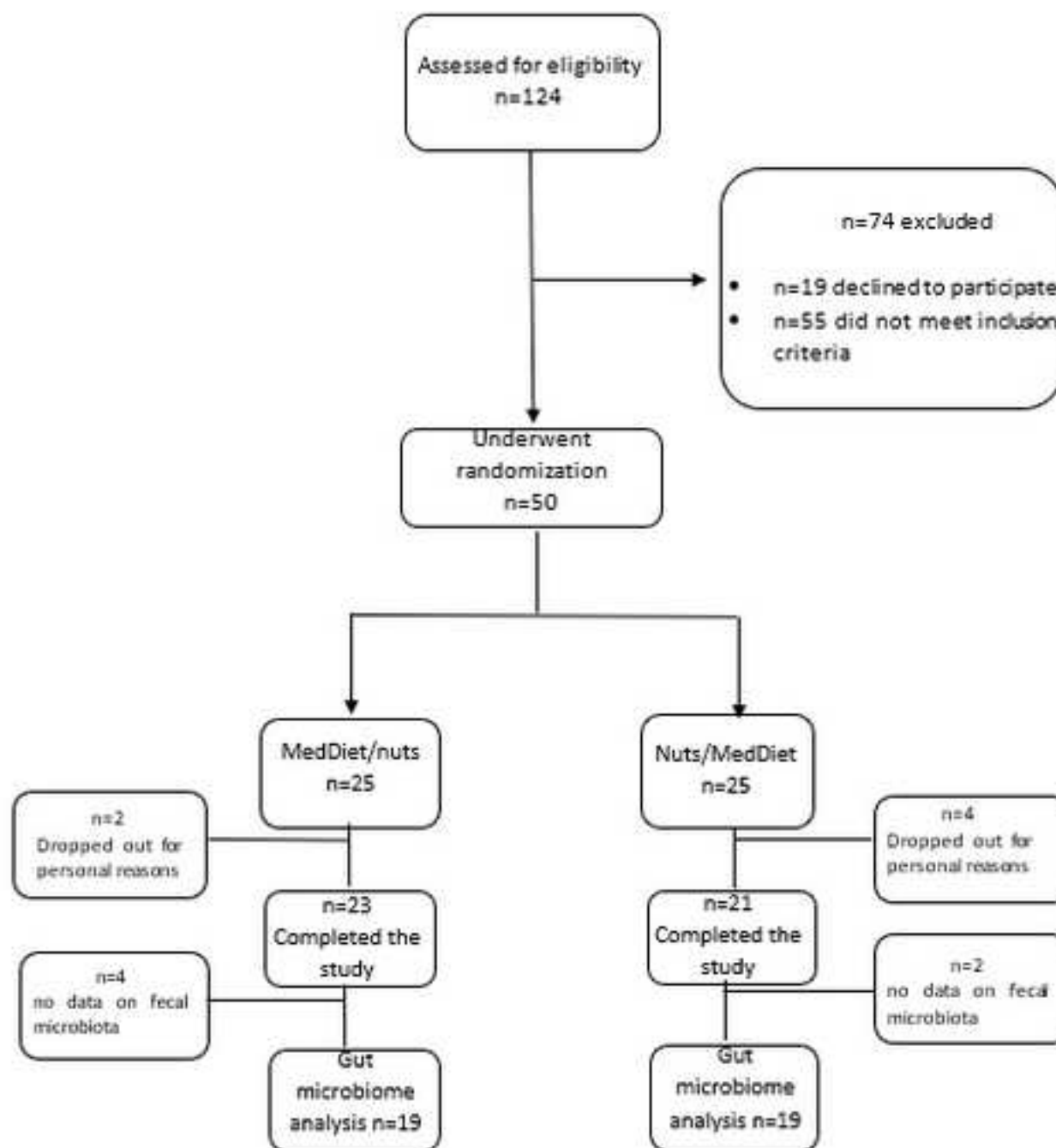
Figure 1: Flow Diagram of the study design METADIET

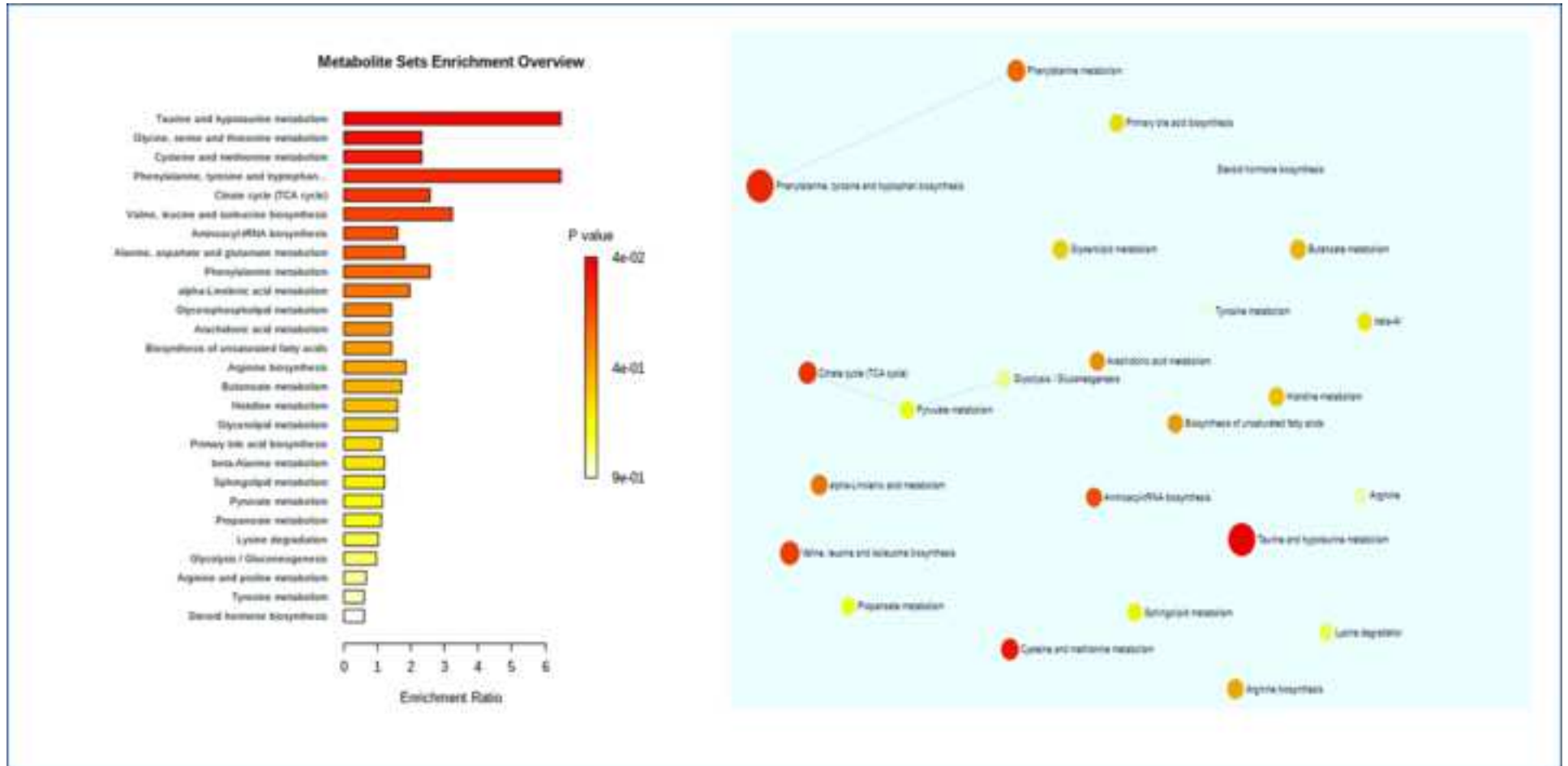
Figure 2. Metabolite set enrichment analysis

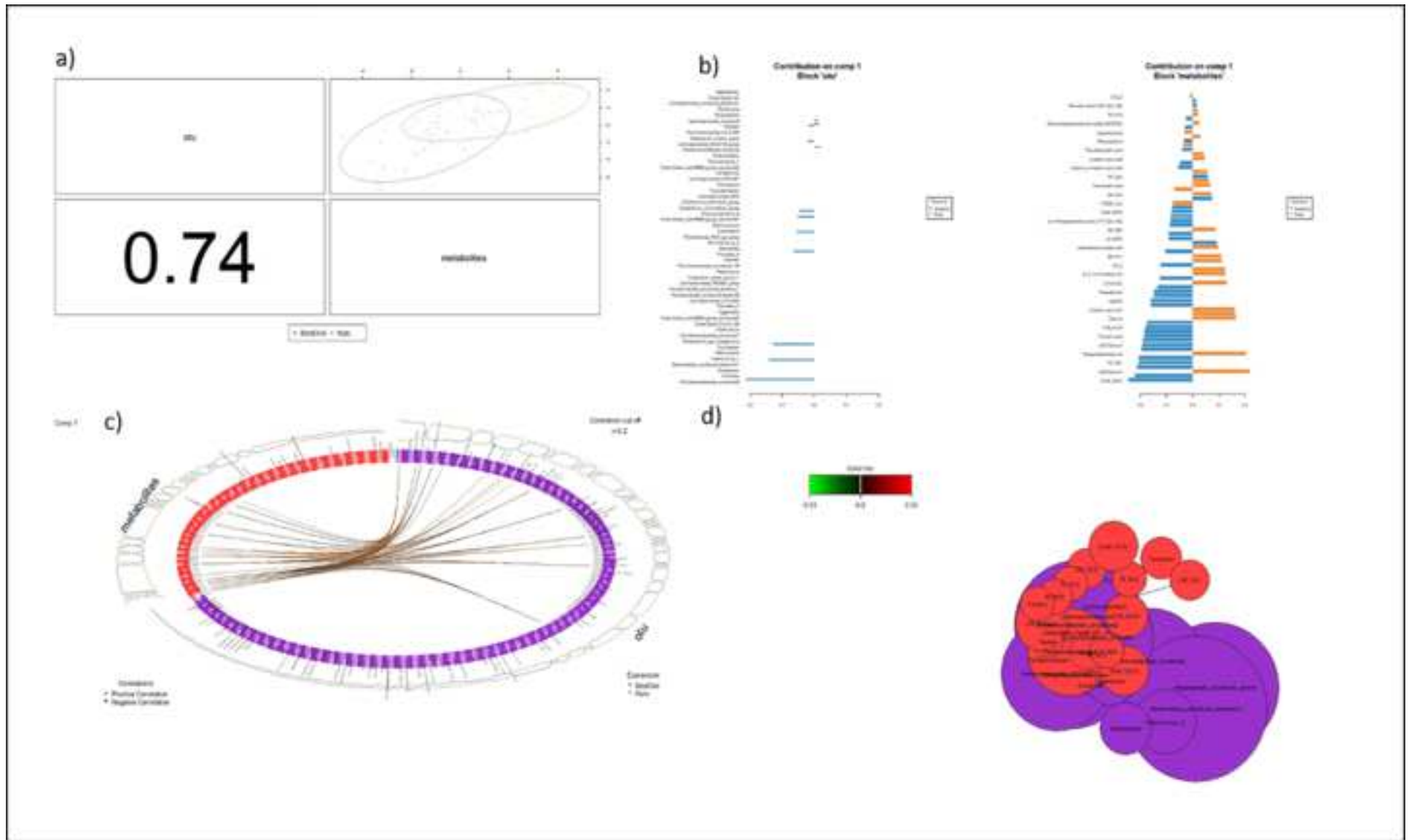
Over representation analysis (ORA) was implemented in the enrichment analysis of MetaboAnalyst 4.0 using the hypergeometric test to evaluate whether a particular metabolite set is represented more than expected by chance within the metabolites previously selected. On the left there is a bar-plot representation of this test and on the right the same information is plotted in a network with a clustering of the identified metabolic pathways

Figure 3. DIABLO graphical and numerical outputs on dual-omics analysis

a) Sample scatterplot from plotDiablo displaying the first component in each data set (upper diagonal plot) and Pearson correlation between each component (lower diagonal plot); b) Loading plot of each feature selected on the first component of both data sets, with color indicating the class with a maximal mean expression value for each feature; c) Circos plot of the final dual-omics signature; d) Relevance network visualization of the selected features at a correlation cut off value of 0.2.









CONSORT 2010 checklist of information to include when reporting a randomised trial*

Section/Topic	Item No	Checklist item	Reported on page No
Title and abstract			
	1a	Identification as a randomised trial in the title	<u>3</u>
	1b	Structured summary of trial design, methods, results, and conclusions (for specific guidance see CONSORT for abstracts)	<u>3-4</u>
Introduction			
Background and objectives	2a	Scientific background and explanation of rationale	<u>5</u>
	2b	Specific objectives or hypotheses	<u>6</u>
Methods			
Trial design	3a	Description of trial design (such as parallel, factorial) including allocation ratio	<u>6</u>
	3b	Important changes to methods after trial commencement (such as eligibility criteria), with reasons	<u>6</u>
Participants	4a	Eligibility criteria for participants	<u>6</u>
	4b	Settings and locations where the data were collected	<u>6</u>
Interventions	5	The interventions for each group with sufficient details to allow replication, including how and when they were actually administered	<u>6</u>
Outcomes	6a	Completely defined pre-specified primary and secondary outcome measures, including how and when they were assessed	<u>7</u>
	6b	Any changes to trial outcomes after the trial commenced, with reasons	<u>9</u>
Sample size	7a	How sample size was determined	<u>Not available</u>
	7b	When applicable, explanation of any interim analyses and stopping guidelines	<u>Not available</u>
Randomisation:			
Sequence generation	8a	Method used to generate the random allocation sequence	<u>6</u>
	8b	Type of randomisation; details of any restriction (such as blocking and block size)	<u>Not available</u>
Allocation concealment mechanism	9	Mechanism used to implement the random allocation sequence (such as sequentially numbered containers), describing any steps taken to conceal the sequence until interventions were assigned	<u>Not available</u>
Implementation	10	Who generated the random allocation sequence, who enrolled participants, and who assigned participants to interventions	<u>Not available</u>
Blinding	11a	If done, who was blinded after assignment to interventions (for example, participants, care providers, those	<u>Not available</u>

		assessing outcomes) and how	
Statistical methods	11b	If relevant, description of the similarity of interventions	Not available
	12a	Statistical methods used to compare groups for primary and secondary outcomes	7
	12b	Methods for additional analyses, such as subgroup analyses and adjusted analyses	7
Results			
Participant flow (a diagram is strongly recommended)	13a	For each group, the numbers of participants who were randomly assigned, received intended treatment, and were analysed for the primary outcome	9
	13b	For each group, losses and exclusions after randomisation, together with reasons	9
Recruitment	14a	Dates defining the periods of recruitment and follow-up	Not available
	14b	Why the trial ended or was stopped	Not available
Baseline data	15	A table showing baseline demographic and clinical characteristics for each group	9
Numbers analysed	16	For each group, number of participants (denominator) included in each analysis and whether the analysis was by original assigned groups	9
Outcomes and estimation	17a	For each primary and secondary outcome, results for each group, and the estimated effect size and its precision (such as 95% confidence interval)	10
	17b	For binary outcomes, presentation of both absolute and relative effect sizes is recommended	10
Ancillary analyses	18	Results of any other analyses performed, including subgroup analyses and adjusted analyses, distinguishing pre-specified from exploratory	Not available
Harms	19	All important harms or unintended effects in each group (for specific guidance see CONSORT for harms)	Not available
Discussion			
Limitations	20	Trial limitations, addressing sources of potential bias, imprecision, and, if relevant, multiplicity of analyses	16
Generalisability	21	Generalisability (external validity, applicability) of the trial findings	11-12
Interpretation	22	Interpretation consistent with results, balancing benefits and harms, and considering other relevant evidence	13-14-15
Other information			
Registration	23	Registration number and name of trial registry	2
Protocol	24	Where the full trial protocol can be accessed, if available	2
Funding	25	Sources of funding and other support (such as supply of drugs), role of funders	1-2

*We strongly recommend reading this statement in conjunction with the CONSORT 2010 Explanation and Elaboration for important clarifications on all the items. If relevant, we also recommend reading CONSORT extensions for cluster randomised trials, non-inferiority and equivalence trials, non-pharmacological treatments, herbal interventions, and pragmatic trials. Additional extensions are forthcoming: for those and for up to date references relevant to this checklist, see www.consort-statement.org.