

1

2 **Influence of different types of LEDs lights on the formation of volatile sulfur**  
3 **compounds in white and rosé wines**

4 Authors: A.M. Mislata <sup>1,2</sup>, M. Puxeu<sup>1</sup>, M. Nadal<sup>1</sup>, S. de Lamo<sup>1</sup>, M. Mestres<sup>2</sup> and R.  
5 Ferrer-Gallego<sup>1\*</sup>

6 <sup>1</sup>VITEC- Centro Tecnológico del Vino, Ctra. Porrera Km.1, 43730 Falset (Tarragona),  
7 Spain

8 <sup>2</sup>Sensometria Instrumental (i-Sens), Department of Analytical Chemistry and Organic  
9 Chemistry, Universitat Rovira i Virgili, Tarragona, 43007, Spain

10 \* Raúl Ferrer-Gallego e-mail: raul.ferrer@vitec.wine

11 **Abstract**

12 The effect of LEDs light on the formation of volatile sulfur compounds (VSCs) and the  
13 final sensory quality of white and rosé wines was evaluated. Thus, different  
14 commercial wines were exposed for ten days to three types of lights. All wine samples  
15 were analyzed throughout the exposure period to determine the usual oenological  
16 parameters together with some other chemical characteristics (color evolution;  
17 riboflavin, cysteine and methionine photodegradation), VSC amounts and sensory  
18 characteristics. The results showed that the wines exposed to ultraviolet light suffered  
19 greater degradation of the aromatic precursors, mainly riboflavin, and had higher  
20 concentrations of VSCs. Regarding LED lights, these produced minimal degradative  
21 effects. So that we can consider this type of light as an alternative to reduce the  
22 economic impact that currently occurs due to the photodegradation of bottled wines.

23 **Keywords:** amino acids, wine, riboflavin, aroma, LED lights, sensory analysis

24

25        **1. Introduction**

26        Sustainable development is currently one of the main goals of the European Union  
27        Commission as well power consumption has become one of the most considerable  
28        environmental issues. Light-Emitting Diodes (LED), ubiquitous in modern electronic  
29        devices and lighting, offer a high series of advantages, especially from a sustainability  
30        point of view. From an oenological standpoint, the recent emergence of high-power  
31        LEDs brings new opportunities to modernize wineries as this type of lighting offers an  
32        exceptional flux range, better light distribution efficiency, versatility, longer shelf life,  
33        and best power conversion, among others (Mills, 2004). Thus, the replacement of  
34        existing lighting in wineries with LED technology can generate significant savings of up  
35        60 % when an optimal control and automation system is used (Vela, 2017).

36        On the other hand, the effect of light radiation on the quality of wine is a recurring  
37        theme. There are studies from the 70s in which already it was observed that the  
38        quality of Champagnes sold in supermarkets was lower than those sold in specialized  
39        stores and that this was due to UV-visible radiation exposure (A. Maujean, Haye, M.,  
40        Feuillat, M., 1978). Later it was found that what was happening was a photochemical  
41        transformation that promoted an unwanted change in the wine properties which was  
42        called *Goût de Lumière* (D'Auria, Emanuele, Mauriello, & Racioppi, 2003) and it has  
43        been suggested that the most damaging wavelengths in bottled wines are those below  
44        520 nm (Clark, Dias, Smith, Ghiggino, & Scollary, 2011; A. Maujean, Haye, M., Feuillat,  
45        M., 1978). This change occurs in color, odor, and flavor and depends on several  
46        factors. The most important ones are; i) the chemical composition of wines, ii) the  
47        irradiation conditions, and iii) the exposure time (Grant-Preece, Barril, Schmidtke,

48 Scollary, & Clark, 2017). Among the different compounds degradable by light, one of  
49 the most studied is riboflavin, which is generated in significant quantities during  
50 alcoholic fermentation (Mattivi, Monetti, Vrhovsek, Tonon, & Andrés-Lacueva, 2000;  
51 Santos, García-Ramírez, & Revuelta, 1995). This molecule has a chemical structure that  
52 absorbs radiation between 300 and 510 nm with maximum absorbance at 370 and 442  
53 nm. Therefore, if wine is irradiated with fluorescent lamps that provide mercury  
54 emissions with maxima around 313, 365, 405, 436, 546, and 578 nm, and phosphor  
55 emissions with maxima around 480 and 580 nm (Spikes, 1981), this molecule will be  
56 irretrievably affected. As for LED lamps, they emit light in wavelength ranges from 350  
57 nm to 750 nm and when dealing with cool white LEDs (commonly used in the wine  
58 industry) they show a maximum emission at 450 nm. However, this technology allows  
59 designing lights that minimize or even remove the emission of some wavelengths so it  
60 could minimize the risk of wine degradation.

61 Apart from riboflavin, many other molecules can be degraded due to light exposure.  
62 Some of them have been related to the Fenton reaction which consumes free SO<sub>2</sub> and  
63 promotes the oxidation of some compounds such as tartaric acid to glyoxylic acid, a  
64 known precursor of the xanthylium cation pigments. The formation of this compound  
65 has been correlated with an increased level of browning, so these chemical  
66 degradations, induced by UV-visible radiation, usually imply loss of wine quality (Clark,  
67 2008; Clark, Prenzler, & Scollary, 2007; George, Clark, Prenzler, & Scollary, 2006;  
68 Maury, Clark, & Scollary, 2010). In addition to these color changes, also UV-visible light  
69 exposure has been found to accelerate oxygen consumption which promotes obtaining  
70 an increasingly reducing atmosphere. These conditions directly affect the aromatic  
71 composition of wine since they favor the appearance of new compounds, among

72 which volatile sulfur compounds (VSCs) stand out (Grant-Preece, Barril, Schmidtke,  
73 Scollary, & Clark, 2017; Haye, 1977). In fact, although VSCs can be formed during the  
74 fermentation process, the photosensitivity of some sulfur-amino acids found in bottled  
75 wine can also lead to their formation. These photo-oxidation reactions also involve  
76 significant amounts ( $> 50 \mu\text{g/L}$ ) of the riboflavin molecule in its triplet excited state. As  
77 this is a strong oxidant, it can easily react with reductant molecules such as methionine  
78 or cysteine, giving rise to other unstable and photosensitive chemical species, which, in  
79 turn, decompose, into more stable compounds such as sulfides, disulfides and some  
80 thiols (Daniela Fracassetti, Tirelli, Limbo, Mastro, Pellegrino, & Ragg, 2020; Kinzurik,  
81 Herbst-Johnstone, Gardner, & Fedrizzi, 2016). These VSCs have a significant impact on  
82 the wine aroma, as they are the key contributors to some undesirable wine flavors  
83 described as rotten eggs, cabbage, or cooked onions.

84 Currently, the photo-oxidation of wine is in the framework for wine researchers (D.  
85 Fracassetti, Di Canito, Bodon, Messina, Vigentini, Foschino, et al., 2021; Daniela  
86 Fracassetti, Tirelli, Limbo, Mastro, Pellegrino, & Ragg, 2020; Lan, Li, Yang, Li, Yuan, &  
87 Guo, 2021). However, little information regarding the influence of the currently used  
88 LEDs on the quality of the wine has been reported. This research aims to evaluate the  
89 use of new LED lights to avoid bottled wines spoilage by determining the photo-  
90 degradation of riboflavin, cysteine, and methionine, the formation of VSCs, and  
91 changes in color throughout the time of light exposure.

## 92 **2. Material and Methods**

### 93 **2.1. Chemicals and reagents**

94 Thiophene, 2-propanethiol and methyl ethyl sulfide were used as internal standards  
95 for the analysis of aromas. Hydrogen sulfide, methanol, ethanol, dimethyl sulfide,  
96 diethyl sulfide, dimethyl disulfide, and diethyl disulfide were used as sulfur volatile  
97 standards. The other standards used were riboflavin, cysteine, and methionine. All of  
98 them were supplied by Sigma-Aldrich (Darmstadt, Germany) with a purity  $\geq$  of 90 %.  
99 HCl and HNO<sub>3</sub> solvents were of high quality and were supplied by Panreac (AppliChem,  
100 Barcelona, Spain).

## 101 2.2. Samples and experimental design

102 Six commercial white wines representative from Albariño variety (Rias Baixas  
103 Appellation of Origin) and three rosé wines from Garnacha variety (Rioja Appellation of  
104 Origin) from 2017 and 2018 chosen randomly were evaluated. All bottles of wine were  
105 subjected to direct light incidence by using three different types of lamps. Light A (LA)  
106 was achieved by using an ultraviolet light lamp like the current one used in wineries,  
107 warehouses, and supermarkets. Light B (LB) is a type of LED that minimized the 400-  
108 450 nm radiation and light C (LC) as an LED lamp that eliminated the radiation emitted  
109 into the 400-450nm region. To carry out the experiment, a black wooden box was  
110 designed into which the bottles were introduced in a horizontal position and the  
111 tested lamps were in the upper part (Figure S1A and B). A box for each type of light  
112 was built and, during the experiment, the three boxes containing the wine samples  
113 were kept in darkness in a controlled temperature room ( $20 \pm 2$  °C). All controls were  
114 stored in darkness also in a controlled temperature room ( $20 \pm 2$  °C). The exposure was  
115 carried out for 10 days. Samples were taken at different analysis times (3, 6, 24, 48,  
116 120, and 240 hours). All treatments were performed in duplicate.

### 117 2.3. Oenological parameters

118 Titratable acidity and pH were potentiometrically measured with an automatic titrator  
119 (TitroMatic Hach from Crison®) and turbidity with a HACH TL2310 turbidimeter. Free  
120 and total sulfur content, alcoholic degree and volatile acidity were determined by  
121 using an infrared spectroscopy (FTIR) system (WineScan™ by FOSS, Hilleroed,  
122 Denmark), internally calibrated according to OIV reference methods.

123 Colorimetric measurements were performed by using a Helios-α spectrophotometer  
124 (Thermo Fisher Scientific, Waltham, MA USA). CIELAB color parameters and color  
125 differences between samples in darkness and samples exposed to light ( $\Delta E^*_{ab} =$   
126  $((\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2)^{1/2}$ ) were calculated.

### 127 2.4. Analysis of aromatic precursors by UPLC

#### 128 2.4.1. Riboflavin

129 For the determination and quantification of riboflavin (vitamin B12) the method  
130 described by (Andrés-Lacueva, Mattivi, & Tonon, 1998) was followed with slight  
131 modifications. Thus, all wine samples were passed through a 0.2 μm syringe filter  
132 (Millipore Corporation, Bedford, MA, USA) and directly injected into the ultra-  
133 performance liquid chromatograph (UPLC 1260 Series, Agilent, Palo Alto, USA) with a  
134 fluorescence detector. The solvents used were solvent A, 0.05 M NaH<sub>2</sub>PO<sub>4</sub> buffer at pH  
135 3.0 with H<sub>3</sub>PO<sub>4</sub>, and solvent B, acetonitrile. The calibration lines used for quantitation  
136 were carried out by preparing standard solutions under dim light, amber glass bottles,  
137 and vials. The concentration range was from 5 to 200 μg/L. The different riboflavin  
138 standard solutions were prepared by diluting a concentrated solution (10.000 μg/L)  
139 with suitable amounts of a mixture of 20 % solvent B and 80 % solvent A. Then each

140 one was filtered and injected into the UPLC. Both the standard and samples were  
141 analyzed in duplicate.

#### 142 2.4.2. Amino Acids

143 Methionine and cysteine were determined following a procedure previously optimized  
144 and validated (Roda, Martín, Mislata, Castaño, Puxeu, & Ferrer-Gallego, 2019). Wine  
145 samples were filtered using a 0.2  $\mu\text{m}$  Millex syringe filter (Millipore Corporation,  
146 Bedford, MA, USA) and subjected to automatic pre-column derivatization with *o*-  
147 phthaldialdehyde (OPA Reagent, Agilent) before the injection to the UPLC (Agilent  
148 UPLC 1260 Series (Palo Alto, USA) equipped with DAD detector and autosampler. The  
149 solvents used were, solvent A: 40 mM  $\text{Na}_2\text{HPO}_4$  at pH 7.8, and solvent B: ACN: MeOH:  
150 water (45:45:10, v/v/v). Detection was performed at  $\lambda = 338$  nm. The identification of  
151 the amino acids was carried out by comparing their retention times with those of the  
152 pure reference standards. Standard calibration curves were used to quantify amino  
153 acids, whose concentration ranges were from 0.1 to 5 mg/L for cysteine and from 1 to  
154 50 mg/L for methionine. The different standard amino acid solutions were prepared by  
155 diluting a concentrated solution (1000 mg/L) in a synthetic wine solution (130 mL of  
156 pure ethanol with 5.5 g of tartaric acid in one liter of Milli-Q water). Then each was  
157 filtered and injected into the UPLC. Both the standard and the samples were run in  
158 duplicate. All samples for analysis were injected in duplicate ( $n = 2$ ).

#### 159 2.5. Analysis of aromatic sulfur compounds by GC-FPD

160 The analysis of aromas was carried out according to the method developed by M.  
161 Mestres et al. 1999 (Mestres, Martí, Busto, & Guasch, 1999), with some minor  
162 modifications. To avoid loss of volatile compounds during sample handling, the wines

163 were refrigerated at -4 °C for 24 hours in a cold room. Once a wine bottle was opened,  
164 the sample to be analyzed was prepared by using a dry ice bath under a stream of  
165 nitrogen gas to provide an inert atmosphere. Specifically, 10 mL of wine together with,  
166 100 µL of 2-propanal to eliminate interference caused by the sulfur content of wine,  
167 and 100 µL of a mixture composed of 1000 µg/L of each internal standard were placed  
168 into a 20 mL glass vial which contained 2.7 g of NaCl and 0.15 g of  
169 ethylenediaminetetraacetic acid (EDTA).

170 The volatile sulfur compounds were extracted and concentrated using the headspace-  
171 solid phase microextraction (HS-SPME) technique with Carboxen/PDMS fibers of 85  
172 µm. The samples were equilibrated for 20 min at 35 °C with shaking and, afterwards,  
173 the fiber was exposed to the headspace for 30 minutes at the same temperature in the  
174 GC auto sampler. The fiber desorption was carried out into a TDU-thermal desorption  
175 unit coupled to a cold equipment, making a temperature ramp from -60 °C to 260 °C,  
176 which allowed the retention of extremely volatile sulfur compounds. After 120  
177 seconds, the unit was heated up to 260 °C and the sample was injected into a GC  
178 7890A gas chromatograph (Agilent Technologies) equipped with a flame photometric  
179 detector (FPD) (G3348B, Agilent Technologies). The separation was carried out using  
180 an HP-1 column (30 m x 0.320 mm x 4.00 µm, Agilent), with helium as carrier gas at a  
181 flow rate of 2.09 mL/min. The oven temperature ramp was: 35 °C to 260 °C in 8 min.  
182 The detector temperature was 200 °C and it was fed with 75 mL/min of hydrogen, and  
183 86 mL/min of synthetic air.

184 The quantification was carried out by means of calibration curves of standards. With  
185 the use of the standards, the retention times of each of the compounds were

186 identified and determined. The results of the volatile compounds were expressed as  
187 quantitative data expressed in  $\mu\text{g/L}$  as a response in consideration to the internal  
188 standard. All analyzes were done in duplicate.

## 189 2.6. Sensory analysis

190 The quantitative descriptive analysis (QDA) was performed by a tasting panel  
191 accredited by ISO 8586: 2010, and in a sensory standardization room (ISO 8589: 2007).  
192 Due to the pandemic situation arising from COVID 2019, the sensory panel was limited  
193 to four expert tasters.

194 The sensory analysis was carried out in 9 sessions of 3 blocks made up of 7 wines each  
195 session, where the panelists did not know the identification of the sample in each of  
196 the glasses, nor the type of light, nor the exposure time of the same.

197 Therefore, a total of 189 wines were tasted, grouped into 9 references by type of wine,  
198 and in turn by type of light and by the time they were exposed. The analysis was based  
199 on the olfactory phase. The aromatic descriptors considered were fresh fruit,  
200 candied/dried fruit, spicy, floral, pastry, evolution, reduction and 'light taste'. All the  
201 data obtained were processed with the FIZZ software (Biosystems, V.2.47B). And a  
202 specific tasting sheet was designed according to the aforementioned olfactory  
203 descriptors.

## 204 2.7. Statistical analysis

205 A simple analysis of variance (ANOVA) was carried out and the data were presented as  
206 the mean  $\pm$  standard deviation. The Tukey procedure was used and differences in p

207 values <0.05 were considered significant. StatGraphics Centurion XVI (Manugistics Inc.,  
208 Pockville, MD, USA) was the program used.

### 209 **3. Results**

#### 210 3.1. Oenological analysis

211 The oenological parameters of the different samples were determined in duplicate  
212 after 0, 3, 6, 24, 48, 120, and 240 hours of light exposure. Table 1 shows the mean  
213 values obtained for each sample at the beginning of the experiment (samples in  
214 darkness or 0 hours of exposure), and after 240 hours (10 days) exposed to the three  
215 types of light. As can be seen, the values found at the beginning of the experiment are  
216 comparable to those found after 10 days to light exposure, except when dealing with  
217 SO<sub>2</sub> and color. Wines presented values of total SO<sub>2</sub> lower than 100 mg/L avoiding  
218 problems related to greater wine reduction when working at high SO<sub>2</sub> amounts (Lopes,  
219 Silva, Pons, Tominaga, Lavigne, Saucier, et al., 2009; Ugliano, 2013). Some changes on  
220 the free and total SO<sub>2</sub> were found in samples exposed to lights. In general, free sulfur  
221 was more affected in samples exposed to LB after 10 days of exposure. According to  
222 other authors (Lan, Li, Yang, Li, Yuan, & Guo, 2021), the exposure to light accelerated  
223 the consumption of free SO<sub>2</sub> compared to controls (in darkness). Furthermore, UV  
224 irradiation resulted in a greater decrease in free SO<sub>2</sub> than white light after 160 days of  
225 storage. This fact raises the possibility of thinking that although after 10 days LB was  
226 the light that most affected free SO<sub>2</sub>, it could be that if we increased this exposure  
227 time it was LA (UV light) that increased the loss of free sulfur in the wines.

228 Regarding color, in general the absorbance of the wines at 420 nm remained stable in  
229 the wines exposed to lights for 10 days compared to 0h. As Diaz et al 2021 (Díaz,

230 Castro, Ubeda, Loyola, & Laurie, 2021) observed until the concentration of free SO<sub>2</sub>  
231 was not remarkably low, the absorbance at 420 nm of the wines remained without  
232 differences between the wines kept in the light and those kept in the dark. In the  
233 Cielab coordinates, it was mainly the *b\** coordinate that presented the most significant  
234 differences in the wines exposed to LA after 10 days compared to 0h. Table 1 shows  
235 how the *b\** values are lower in the wines exposed to LA (UV light), which indicates a  
236 greater loss of yellow color or a greater browning of the wines. As observed by Lan et  
237 al 2021 (Lan, Li, Yang, Li, Yuan, & Guo, 2021), the exposure to ultraviolet light affected  
238 the color parameters more compared to the control (in the dark). However, when  
239 calculating *AE\*ab*, all the results obtained were less than 1.0 CIELAB unit (table 1).  
240 However, the wines exposed to LA return values very close to 1 so that, if we consider  
241 the dispersion, they will exceed it, although not in all cases Therefore, it is probable  
242 that if the experiment was extended along time, color differences will be appeared and  
243 detected with the naked eye by the human eye in LA samples (Gonnet, 1998).

## 244 3.2. Analysis of aromatic precursors by UPLC

### 245 3.2.1. Riboflavin

246 The mean riboflavin values of the samples that remained in the dark (time 0h)  
247 presented concentrations between 138-182 µg/L in white wines, and between 152-178  
248 µg/L in rosé wines (table S1). These values indicate that the studied wines presented  
249 high concentrations of riboflavin, since in all cases they exceeded 100 µg/L. Although  
250 the concentration values agree with others found in literature, these are above 100  
251 µg/L so, according to previous studies, the sample wines are at risk of triggering the  
252 appearance of volatile sulfur compounds in wine (D. Fracassetti, Limbo, Pellegrino, &

253 Tirelli, 2019; Mattivi, Monetti, Vrhovsek, Tonon, & Andrés-Lacueva, 2000). In general,  
254 rosé wines presented more abrupt degradation of riboflavin than white wines  
255 throughout the time of exposure to the lights. Furthermore, this greater degradation  
256 of riboflavin was observed in all the wines exposed mainly to light LA followed by LB  
257 (figure 1). However, it is worth noting the behavior of the wines with exposure to LC,  
258 because the riboflavin concentrations practically did not vary throughout the exposure  
259 time. This behavior is probably due to wavelengths in the 370 to 450 nm range.  
260 Wavelengths at which LA light emits and LC light completely removes, can probably  
261 excite riboflavin to the short-lived singlet state and then to the triplet state, which  
262 then participates in photo-oxidation reactions through various mechanisms (Grant-  
263 Preece, Barril, Schmidtke, Scollary, & Clark, 2017).

264 In general, as figure 1 shows, white wines (W1-W6) presented different degrees of  
265 affectation. After the last exposure time (240 hours) with LA, riboflavin losses of up to  
266 44 % (W1 and W6) were observed. This decrease in riboflavin concentration occurred  
267 progressively over time of exposure. Even after only 24 hours of exposure, the wines  
268 showed losses of 17 and 13 % (W1 and W6, respectively). Regarding the values  
269 obtained in the samples exposed with LB light, in general, the wines showed a trend  
270 like that observed with LA light but with less degradation of riboflavin in all cases. With  
271 this light (LB), after 48 hours of exposure, the wines presented losses between 11 and  
272 16 % and these degradation values increased up to 35-38 % after the last exposure  
273 time (240h) (W1 and W6, respectively). In general, both lights, LA and LB, behaved  
274 similarly in the degradation of riboflavin in white wines with the exception of W4 and  
275 W5 wines in which there was hardly any degradation with LB (and LC) but where the  
276 degradation of riboflavin reached values up to 20 % after 240 h of exposure to LA. The

277 green color of the bottle of these two samples, which unlike the rest of the wines had  
278 bottles of clear color, could be a crucial factor in the lowest photodegradation showed.  
279 Different wavelengths of light will be transmitted depending on the glass color, and  
280 exposing wine to light at wavelengths close to 370 or 442 nm is particularly effective in  
281 inducing riboflavin degradation and the formation of volatile sulfur compounds,  
282 especially when using transparent glass bottles (Dias, Smith, Ghiggino, & Scollary,  
283 2012; D. Fracassetti, Gabrielli, Encinas, Manara, Pellegrino, & Tirelli, 2017).

284 In the case of rosé wines, as shown in figure 1, the pronounced degradations produced  
285 with exposure to light LA over the time in all wines (W7-W9) stand out, reaching losses  
286 of up to 85 % (W8) after 10 days of exposure. Even after 24 hours of exposure, this  
287 decrease was already around 30 % of degradation. And only after 6 hours a loss of 10  
288 % of riboflavin was produced. As for rosé wines exposed to LB light, they showed a  
289 similar trend to that observed with LA light, but with less degradation of riboflavin over  
290 time. After the last time of exposure, they showed degradation of around 45 %, half of  
291 the degradation produced with LA.

292 Finally, it should be emphasized again that in general none of the white and rosé wines  
293 exposed to LC light showed riboflavin degradation over time.

### 294 3.2.2. Methionine and Cysteine

295 Amino acids such as cysteine and methionine represent the most important form of  
296 total nitrogen in musts and wines. In addition to being the most important precursors  
297 of certain volatile sulfur compounds (Bekker, Wilkes, & Smith, 2018), due to their  
298 polyfunctional character, amino acids have great chemical reactivity compared to  
299 carbonyl compounds, in particular with sugars, according to the Maillard reaction

300 (Marchand, De Revel, & Bertrand, 2000). This Strecker degradation of methionine and  
301 cysteine to aldehydes by  $\alpha$ -dicarbonyl compounds formed during fermentation or  
302 oxidation contributes to the evolution of aroma in bottled wine (Ugliano, 2013).

303 The initial content of cysteine ranged between 0.35 and 1.79 mg/L in white wines and  
304 between 0.56 and 0.86 mg/L in rosé wines (table S2), values that are within the usual  
305 ones (Bekker, Wilkes, & Smith, 2018). As can be seen in figure 2, on the whole, the  
306 wines suffered decreases in cysteine after exposure to light, mainly with LA light,  
307 reaching decreases of around 30-45 % in white wines, and even up to 55 % in rosé  
308 wines (W9). In the case of exposure to LB light, these decreases in cysteine were  
309 slightly less than those produced with LA, reaching values of around 20-35% in white  
310 wines and up to 50 % in rosé wines (W9). However, the cysteine decreases of the  
311 samples with exposure to LC light were between 10-35 %, up to three times less than  
312 the losses obtained with LA.

313 This degradation of the amino acid cysteine observed in all the wines studied may have  
314 as a consequence the formation of VSCs through Strecker degradation of sulfur amino  
315 acids, as already observed by Pripis et al 2000 (Pripis-Nicolau, De Revel, Bertrand, &  
316 Maujean, 2000). Cysteine can react with the  $\alpha$ -dicarbonyl compound in wine to form  
317 hydrogen sulfide, methanethiol, and other volatile compounds (Marchand, De Revel, &  
318 Bertrand, 2000)

319 The initial concentrations of methionine in the wines ranged between 3.6 and 6.0 mg/L  
320 in white wines and between 4.8 and 8.1 mg/L in rosé wines (table S2), values similar to  
321 those found in literature (D. Fracassetti, Limbo, Pellegrino, & Tirelli, 2019; Grant-  
322 Preece, Barril, Schmidtke, Scollary, & Clark, 2017). Furthermore, the range of

323 concentrations in wine was around between 1 and 37 mg/L (Bekker, Wilkes, & Smith,  
324 2018). Figure 2 shows how in general, after 10 days of exposure, the wines exposed to  
325 LA light presented further degradation of methionine, reaching losses of around 10-30  
326 % in white wines and up to 40 % in rosé wines. (W9). In the case of samples exposed to  
327 LB light, generally this decrease in methionine was slightly less than with LA, but  
328 decreases in methionine were achieved in the same range (around 10-30 %). However,  
329 it should be noted that W1, W4, W5, W6, and W7 wines exposed to LC light, did not  
330 exceed 10 % loss of methionine in the last exposure time.

331 These decreases in methionine may be due to the presence of riboflavin, which acts as  
332 a photosensitizer in its degradation. Fracassetti et al 2019 (D. Fracassetti, Limbo,  
333 Pellegrino, & Tirelli, 2019) observed that methionine proved to be stable in a  
334 hydroalcoholic acid solution without riboflavin since no degradation occurred in the  
335 model wine enriched with methionine (3 mg/L) and exposed to light for two hours. In  
336 contrast, the presence of riboflavin caused methionine degradation of up to 27 %,  
337 depending on the initial concentrations of the two compounds. Furthermore, when  
338 riboflavin and methionine concentrations were high, then the amounts of degraded  
339 methionine increased as well, which demonstrated the influence of the methionine  
340 concentration on its own degradation.

### 341 3.3. Analysis of volatile sulfur composition

342 The analysis of volatile sulfur compounds (VSCs) was carried out as these can be  
343 generated by the degradation of riboflavin and the amino acids, methionine, and  
344 cysteine, later in post-bottling (Bekker, Day, Holt, Wilkes, & Smith, 2016; D. Fracassetti,  
345 Limbo, Pellegrino, & Tirelli, 2019). In total six compounds were detected and

346 quantified, which were grouped in three aromatic families based on their chemical  
347 structures: thiols, sulfides and disulfides.

348 The thiol family consisted of hydrogen sulfide ( $H_2S$ ), methanethiol (MeSH), and  
349 ethanethiol (EtSH). Although, as in other studies (Bekker, Day, Holt, Wilkes, & Smith,  
350 2016; Siebert, Solomon, Pollnitz, & Jeffery, 2010), EtSH was not detected in any of the  
351 samples analyzed. Regarding  $H_2S$  and MeSH, these are the main compounds  
352 responsible for the formation of "reducing" aromas after bottling (Ugliano, 2013;  
353 Ugliano, Dieval, Siebert, Kwiatkowski, Aagaard, Vidal, et al., 2012; Ugliano, Kolouchova,  
354 & Henschke, 2011). As shown in figure 3, in general both compounds, increased their  
355 concentrations throughout the light exposure time but this increase was much higher  
356 when dealing with the exposure to light A, followed by light B, and to a lesser extent,  
357 light C.

358 Hydrogen sulfide is a characteristic compound for providing wines with unpleasant  
359 aromas of rotten eggs, decomposing algae, or wastewater when its concentration is  
360 higher than its odor threshold (OT) of  $1.6 \mu\text{g/L}$  in vinica matrix (Siebert, Solomon,  
361 Pollnitz, & Jeffery, 2010). As can be seen in figure 3a, all the wines except for W4  
362 exceeded the OT concentration of  $H_2S$  just after 6 hours of exposure with both LA and  
363 LB light. In the case of exposure to LC light, this sensory limit was not or slightly  
364 exceeded. In general, it should be noted that rosé wines (W7-W9) presented the  
365 highest concentrations of this compound after exposure to all lights. In general, the  
366 highest concentrations of  $H_2S$  were obtained in the wines exposed to LA. In some  
367 cases, these wines exposed to LA came to present twice the concentration of  $H_2S$  than

368 that obtained with LB light, and even three times higher than that obtained with LC  
369 light.

370 After the last exposure time (240 hours) with light LA, in general, the samples  
371 continued to show an increase in the concentration of H<sub>2</sub>S compared to the 6 hours of  
372 exposure. However, after 10 days (240 hours) of exposure to LB light, only rosé wines  
373 were affected. The same happened with LC light, which mainly affected rosé wines but  
374 to a lesser extent than with previous lights, being half the concentration obtained with  
375 LB.

376 Another of the thiols studied was methanethiol (MeSH). This compound is  
377 characteristic for providing wines with aromas related to descriptors such as  
378 putrefaction and cooked cabbage. In addition, it should be noted that this compound  
379 presents very low OT values (0.3 µg/L) which makes it play an important role in the  
380 perception of 'reducing' aromas in wine (D. Fracassetti & Vigentini, 2018; Nguyen  
381 Dang-Dung, 2012). As can be seen in figure 3.b, only after 6 hours of exposure to LA  
382 light, all wines (except W4 and W6) exceeded the OT concentration of MeSH.  
383 However, only two white wines (W3 and W5) and two rosés (W8 and W9) exceeded  
384 the OT with exposure to LB light. And in the case of exposing the samples in LC, only  
385 two wines (W3 and W9) slightly exceeded the sensory limit. After the last analysis time  
386 (240 hours), the samples exposed to LA light increased their MeSH concentrations  
387 concerning the previous time, mainly highlighting the large increase produced in W9  
388 (5.5 µg / L), since it was practically the double that obtained after 6 hours. In the case  
389 of exposure to LB light, generally, the wines that were already affected after 6 hours,  
390 followed an increase in concentration. And finally, in general, the wines exposed to LC

391 light did not show great changes in the concentrations of MeSH throughout the  
392 exposure time.

393 As seen in other studies, cysteine and methionine are the main sources of H<sub>2</sub>S and  
394 MeSH formation (Bekker, Wilkes, & Smith, 2018; Perpète, Duthoit, De Maeyer, Imray,  
395 Lawton, Stavropoulos, et al., 2006). But in addition, riboflavin plays a key role in the  
396 induction of methionine degradation (D. Fracassetti, Limbo, Pellegrino, & Tirelli, 2019).  
397 This fact could explain the relationship observed between the higher degradations of  
398 these amino acids (figure 2) and the higher concentrations of these aromatic  
399 compounds (figure 3) obtained in the studied wines and exposed to LA light, mainly in  
400 the case observed in W9.

401 Therefore, exposure to LA light favored the appearance of thiols in the samples, mainly  
402 in rosé wines. The LB light affected to a lesser extent than the LA, but emphasizing the  
403 increase of H<sub>2</sub>S mainly in rosé wines. And especially, it should be noted that the LC  
404 light hardly affected the samples in the formation of thiols, except in rosé wines where  
405 the concentration of H<sub>2</sub>S increased, although to a lesser extent compared to the other  
406 lights.

407 The next family studied was the sulfides. This family consisted of dimethyl sulfide  
408 (Me<sub>2</sub>S) and diethyl sulfide (Et<sub>2</sub>S) (figure 4). As shown in figure 4, and as was the case in  
409 the thiol family, in general, both compounds, Me<sub>2</sub>S and Et<sub>2</sub>S, increased their  
410 concentrations throughout the exposure time, in all wines. In general, this increase  
411 was much greater with the exposure of light LA, followed by light LB and to a lesser  
412 extent with light LC.

413 Dimethyl sulfide (Me<sub>2</sub>S) is a compound that is associated with cabbage, asparagus,  
414 corn, or vegetable flavors when present in high concentrations, since it has an odor  
415 threshold of 25 µg/L (Mestres, Busto, & Guasch, 2000; Siebert, Solomon, Pollnitz, &  
416 Jeffery, 2010; Ullrich, Neef, & Schmarr, 2018). Factors that influence Me<sub>2</sub>S formation in  
417 wines after bottling include grape variety, viticulture and winemaking processes, and  
418 wine pH value. These factors likely affect the Me<sub>2</sub>S precursor compounds (Bekker, Day,  
419 Holt, Wilkes, & Smith, 2016; Escudero, Campo, Fariña, Cacho, & Ferreira, 2007;  
420 Ugliano, 2013). As shown in figure 4a, in general rosé wines and some white wines (W4  
421 and W6) presented the highest concentrations of the compound Me<sub>2</sub>S under the  
422 exposure of all the lights, presenting very high values after 6 hours of exposure to the  
423 light LA. Also, it should be noted that W9 was the only wine that exceeded the OT of  
424 this compound. However, in the particular case of the wines exposed to both LB and LC  
425 light, none of them exceeded the OT, at this time. After 10 days of exposure with LA  
426 light, all rosé wines exceeded the OT, in addition to white wine W6. In the case of  
427 exposure to LB light, on the whole, the wines behaved similarly to those exposed to LA  
428 light. However, it should be noted that none of the samples exposed to LC light  
429 presented values higher than OT for Me<sub>2</sub>S.

430 As is already known, riboflavin and methionine can be precursors of Me<sub>2</sub>S (D'Auria,  
431 Emanuele, Mauriello, & Racioppi, 2003; A. Maujean, Haye, Feuillat, Thomas, & Petit,  
432 1978). Therefore, when observing figures 1 and 2, the highest degradations of  
433 riboflavin and methionine are obtained in samples from W6 to W9, which also coincide  
434 with the highest concentrations of Me<sub>2</sub>S obtained (Figure 4a).

435 The second compound analyzed within the sulfide family was **diethyl sulfide**. This  
436 compound is characteristic because it has garlic as the main aromatic descriptor. In  
437 addition, it has a great reducing influence on wines since it has a very low sensory  
438 limit, OT 0.93 µg/L (Mestres, Busto, & Guasch, 2000; Siebert, Solomon, Pollnitz, &  
439 Jeffery, 2010; Ullrich, Neef, & Schmarr, 2018). In figure 4.b it can be seen how the  
440 white wine W2, and all the rosés (W7-W9), in general, presented the highest  
441 concentrations of Et<sub>2</sub>S after being exposed to all the lights. After the 6-hour exposure  
442 time to LA light, three of the study wines (W2, W7 and W9) presented Et<sub>2</sub>S values  
443 higher than their OT. In the case of exposure with the LB and LC lights, only two wines  
444 (W2 and W9) exceeded the OT value. After 10 days of exposure to LA light, in general,  
445 all the wines exceeded the OT of Et<sub>2</sub>S. In the case of the exposure with the LB and LC  
446 lights, it was the rosé wines (W7-W9) and the W2 white wine, which exceeded the OT  
447 of Et<sub>2</sub>S (with the exception of the W8 with LC). However, it should be noted that  
448 exposure of the wines to LC light over time did not dramatically increase the  
449 concentrations of the Et<sub>2</sub>S compound.

450 Therefore, exposure to LA light favored the appearance of sulfides in the samples,  
451 mainly in rosé wines. The LB light affected less than the LA light. And the LC light, in  
452 general, did not favor the formation of sulfides in the wines over time.

453 And finally, there is the family of disulfides, in which the compounds dimethyl disulfide  
454 (Me<sub>2</sub>S<sub>2</sub>) and diethyl disulfide (Et<sub>2</sub>S<sub>2</sub>) were evaluated. As in the previous families (thiols  
455 and sulfides), both compounds were increasing their concentrations throughout the  
456 exposure time to the lights. In addition, generally this increase was greater with the  
457 exposure of light LA, followed by light LB and to a lesser extent with light LC.

458 Dimethyl disulfide is a characteristic compound for providing aromas related to cooked  
459 cabbage, sulfurous or onion, and for having a high odor threshold (29 µg/L) (Siebert et  
460 al 2010; Ullrich et al 2017). As shown in figure 5a, in general all wines (except W5)  
461 were affected by LA light, followed by LB, over time. After 6 hours of exposure to LA  
462 light, all wines exceeded the OT of Me<sub>2</sub>S<sub>2</sub> (except W4 and W5). This could be because  
463 the W4 and W5 wines had the green bottle color, which can act as a protector in the  
464 formation of the Me<sub>2</sub>S<sub>2</sub> compound. However, with LB light exposure, only rosé wines  
465 (W7 and W8) exceeded OT. It should be noted that exposure to LC light, in general, did  
466 not affect the increase in the concentration of Me<sub>2</sub>S<sub>2</sub>, since none of the wines  
467 exceeded the OT. After the last analysis time (240 hours), all samples exceeded the  
468 sensory limit of Me<sub>2</sub>S<sub>2</sub> with exposure to LA light. In the case of exposure to LB, the  
469 highest increases in Me<sub>2</sub>S<sub>2</sub> occurred in the same wines as with exposure to LA, but with  
470 lower concentrations. And in the case of exposure with LC light, it should be noted that  
471 in general none of the wines exceeded the OT of Me<sub>2</sub>S<sub>2</sub> (except W7, which slightly  
472 exceeded it).

473 Maujean and Seguin, 1983a, b (A. Maujean, Haye, Feuillat, Thomas, & Petit, 1978),  
474 observed that two MeSH molecules could produce dimethyl disulfide. This mechanism  
475 has been investigated in white wines that are responsible for an unpleasant aroma,  
476 generally called 'lumière'. In this study, when comparing the values obtained from the  
477 degradation of riboflavin (figure 1) and those obtained from the compound of Me<sub>2</sub>S<sub>2</sub>  
478 (figure 5a), it can be observed that the wines with the highest degradation of riboflavin  
479 were those with the highest concentration of Me<sub>2</sub>S<sub>2</sub>, in whites wines W1, W3, W6 and  
480 in all rosé wines (W7-W9).

481 The second disulfide analyzed was diethyl disulfide ( $\text{Et}_2\text{S}_2$ ). This compound gives wines  
482 aromas associated with bad smells or onion notes when concentrations are higher  
483 than  $4.3 \mu\text{g/L}$  (OT) (Mestres, Busto, & Guasch, 2000; Siebert, Solomon, Pollnitz, &  
484 Jeffery, 2010; Ullrich, Neef, & Schmarr, 2018). As can be seen in figure 5.b, in general,  
485 the samples that presented higher concentrations were rosé wines and W2 white  
486 wine, with exposure to LA and followed by exposure to LB. However, it should be  
487 noted that exposure of the samples to LC light did not affect the increase in  $\text{Et}_2\text{S}_2$ . After  
488 6 hours of exposure with LA, almost all wines (except W1, W4, and W5) exceeded the  
489 OT. In the case of exposure to LB, it was the same wines as those with LA that  
490 exceeded the OT of  $\text{Et}_2\text{S}_2$ , presenting similar concentrations. However, with exposure  
491 to LC none of the samples presented concentrations higher than OT. After 10 days of  
492 exposure to LA, all the wines presented values higher than OT (except W1). The same  
493 happened with the exposure to LB, where most of the wines exceeded the OT (except  
494 W1 and W3). However, it should be noted that in exposure to LC, none of the wines  
495 presented higher OT values (except W7).

496 Therefore, both exposures to LA light and LB light favored the appearance of disulfides  
497 in the samples, mainly after 10 exposure times. However, it should be noted that the  
498 LC light hardly affected the samples in the formation of both disulfides.

499

#### 500 3.4. Sensory analysis

501 Finally, the chemical analysis of the volatile composition of the wines was completed  
502 by sensory analysis. The results of the sensory analysis seem to confirm the patterns  
503 observed in the chemical analysis of aromatic compounds (figure S2). In general, the

504 wines showed a greater appearance of reduction and 'lumiere' aromas after the  
505 longest exposure time, mainly with exposure to LA light and LB light, although less  
506 intense with the latter (LB). However, the wines exposed to LC light showed hardly any  
507 reduction or 'lumiere' notes. In addition, in the analysis, it was observed how  
508 throughout the exposure time the fresh fruit and the floral notes, are decreasing,  
509 presenting a greater intensity of the same all the controls in general.

510

#### 511 **4. Conclusions**

512 Our findings highlighted that both the degradation of methionine and to a greater  
513 extent the degradation of riboflavin resulted in the formation of high levels of VSCs. As  
514 expected, the photodegradation of riboflavin was greatly affected by the light emitted  
515 in the region of 400-450 nm (LA), reaching losses of up to 80 % after 10 days of  
516 exposure. However, it should be noted that wines exposed to new LEDs, eliminated by  
517 this wavelength region (LC), showed conservation of riboflavin concentration  
518 throughout the exposure time. Consequently, the wines exposed to LA presented  
519 higher concentrations of thiols, sulfides, and disulfides, exceeding the sensory limits in  
520 most cases. Wines with LB behaved similarly but presenting less intense riboflavin  
521 degradation than with LA, and the formation of VSCs but in lower concentrations than  
522 with LA. However, LC was the light with which the wines were preserved over time  
523 without presenting high concentrations of VSCs. Demonstrating that simply by  
524 eliminating this wavelength range (400-450), new LEDs sources can completely avoid  
525 the appearance of unpleasant aromas in wines related to VSCs. Further research

526 should be done to evaluate the effect of different factors such as glass color, bottle  
527 thickness or type of wines.

528

529 **Figure captions**

530 Figure 1. Riboflavin degradation in % of all wines (W1-W9) throughout the time of  
531 exposure (0-240 hours) to the different study lights (LA, LB and LC).

532 Figure 2. Degradation of methionine and cysteine in percentage of all wines (W1-W9)  
533 throughout the exposure time (0-6-240 hours) at different LEDs lights (LA, LB and LC).

534 Figure 3. Aromatic family of thiols, hydrogen sulfide (a) and methanethiol (b)  
535 concentrations ( $\mu\text{g/L}$ ) obtained in all wines (W1-W9) throughout the exposure time (0-  
536 6-240 hours) with each of the study lights (LA, LB and LC). Samples ( $n = 2$ ).

537 Figure 4. Aromatic family of sulfides, dimethyl sulfide (a) and diethyl sulfide (b)  
538 concentrations ( $\mu\text{g/L}$ ) obtained in all wines (W1-W9) throughout the exposure time (0-  
539 6-240 hours) with each of the study lights (LA, LB and LC). Samples ( $n = 2$ ).

540 Figure 5. Aromatic family of disulfides, dimethyl disulfide (a) and diethyl disulfide (b)  
541 concentrations ( $\mu\text{g/L}$ ) obtained in all wines (W1-W9) throughout the exposure time (0-  
542 6-240 hours) with each of the study lights (LA, LB and LC). Samples ( $n = 2$ ).

543 Figure S1. a) Experimental design of the black wooden box where the wines were  
544 exposed to the three study lights. b) Spotlights which were located in the upper part of  
545 the black box. c) Wine bottles exposed to light, and protected with aluminum foil for  
546 protection until analysis.

547 Figure S2. Aromatic profiles of W1-W9 wines obtained with exposure to the three  
548 types of lights (LA, LB and LC) throughout the exposure time (0 - 240 hours). Results  
549 obtained by the mean of the scores given by the tasters.

550

## 551 **Acknowledgements**

552 The authors would like to acknowledge Ministerio de Ciencia, Innovación y  
553 Universidades for their financial support in RETASTELED Project (RTC-2017-6646-2) and  
554 also thank Grupo Prilux and the Bodegas Martin Codax and Ramon Bilbao for its  
555 commitment and advice.

## 556 **References**

- 557 Andrés-Lacueva, C., Mattivi, F., & Tonon, D. (1998). Determination of riboflavin, flavin  
558 mononucleotide and flavin-adenine dinucleotide in wine and other beverages by high-  
559 performance liquid chromatography with fluorescence detection. *Journal of*  
560 *Chromatography A*, 823(1), 355-363.
- 561 Bekker, M. Z., Day, M. P., Holt, H., Wilkes, E., & Smith, P. A. (2016). Effect of oxygen exposure  
562 during fermentation on volatile sulfur compounds in Shiraz wine and a comparison of  
563 strategies for remediation of reductive character. *Australian Journal of Grape and*  
564 *Wine Research*, 22(1), 24-35.
- 565 Bekker, M. Z., Wilkes, E. N., & Smith, P. A. (2018). Evaluation of putative precursors of key  
566 'reductive' compounds in wines post-bottling. *Food Chem*, 245, 676-686.
- 567 Clark, A. C. (2008). The production of yellow pigments from (+)-catechin and dihydroxyfumaric  
568 acid in a model wine system. *European food research and technology = Zeitschrift fur*  
569 *Lebensmittel-Untersuchung und -Forschung. A*, 226(4), 925-931.
- 570 Clark, A. C., Prenzler, P. D., & Scollary, G. R. (2007). Impact of the condition of storage of  
571 tartaric acid solutions on the production and stability of glyoxylic acid. *Food Chem*,  
572 102(3), 905-916.
- 573 D'Auria, M., Emanuele, L., Mauriello, G., & Racioppi, R. (2003). On the origin of "Goût de  
574 Lumiere" in champagne. *Journal of Photochemistry and Photobiology A: Chemistry*,  
575 158(1), 21-26.
- 576 Dias, D. A., Smith, T. A., Ghiggino, K. P., & Scollary, G. R. (2012). The role of light, temperature  
577 and wine bottle colour on pigment enhancement in white wine. *Food Chem*, 135(4),  
578 2934-2941.
- 579 Díaz, I., Castro, R. I., Ubeda, C., Loyola, R., & Laurie, V. F. (2021). Combined effects of sulfur  
580 dioxide, glutathione and light exposure on the conservation of bottled Sauvignon  
581 blanc. *Food Chem*, 356.

582 Escudero, A., Campo, E., Fariña, L., Cacho, J., & Ferreira, V. (2007). Analytical characterization  
583 of the aroma of five premium red wines. Insights into the role of odor families and the  
584 concept of fruitiness of wines. *Journal of Agricultural and Food Chemistry*, 55(11),  
585 4501-4510.

586 Fracassetti, D., Di Canito, A., Bodon, R., Messina, N., Vigentini, I., Foschino, R., & Tirelli, A.  
587 (2021). Light-struck taste in white wine: Reaction mechanisms, preventive strategies  
588 and future perspectives to preserve wine quality. *Trends in Food Science and*  
589 *Technology*, 112, 547-558.

590 Fracassetti, D., Gabrielli, M., Encinas, J., Manara, M., Pellegrino, I., & Tirelli, A. (2017).  
591 Approaches to prevent the light-struck taste in white wine. *Australian Journal of Grape*  
592 *and Wine Research*, 23(3), 329-333.

593 Fracassetti, D., Limbo, S., Pellegrino, L., & Tirelli, A. (2019). Light-induced reactions of  
594 methionine and riboflavin in model wine: Effects of hydrolysable tannins and sulfur  
595 dioxide. *Food Chem*, 298.

596 Fracassetti, D., Tirelli, A., Limbo, S., Mastro, M., Pellegrino, L., & Ragg, E. M. (2020).  
597 Investigating the Role of Antioxidant Compounds in Riboflavin-Mediated Photo-  
598 Oxidation of Methionine: A 1H-NMR Approach. *ACS Omega*, 5(40), 26220-26229.

599 Fracassetti, D., & Vigentini, I. (2018). Occurrence and Analysis of Sulfur Compounds in Wine. In  
600 *Grapes and Wines - Advances in Production, Processing, Analysis and Valorization*, (pp.  
601 225-251): INTECH, Open Access book.

602 George, N., Clark, A. C., Prenzler, P. D., & Scollary, G. R. (2006). Factors influencing the  
603 production and stability of xanthylum cation pigments in a model white wine system.  
604 *Australian Journal of Grape and Wine Research*, 12(1), 57-68.

605 Gonnet, J. F. (1998). Colour effects of co-pigmentation of anthocyanins revisited - 1. A  
606 colorimetric definition using the CIELAB scale. *Food Chem*, 63(3), 409-415.

607 Grant-Preece, P., Barril, C., Schmidtke, L. M., Scollary, G. R., & Clark, A. C. (2017). Light-induced  
608 changes in bottled white wine and underlying photochemical mechanisms. *Critical*  
609 *Reviews in Food Science and Nutrition*, 57(4), 743-754.

610 Haye, B., Maujean, A., Jacquemin, C., Feuillat, M. (1977). Contribution a l'étude des "Gouts de  
611 lumière" dans le vin de Champagne. I.- Asepects analytiques - dosage des mercaptans  
612 et des thiols dans les vins.-. *Connaissance Vigne et Vin*, 11(3), 243-254.

613 Kinzurik, M. I., Herbst-Johnstone, M., Gardner, R. C., & Fedrizzi, B. (2016). Hydrogen sulfide  
614 production during yeast fermentation causes the accumulation of ethanethiol, S-ethyl  
615 thioacetate and diethyl disulfide. *Food Chem*, 209, 341-347.

616 Lan, H., Li, S., Yang, J., Li, J., Yuan, C., & Guo, A. (2021). Effects of light exposure on chemical  
617 and sensory properties of storing Meili Rosé wine in colored bottles. *Food Chem*, 345,  
618 128855.

619 Lopes, P., Silva, M. A., Pons, A., Tominaga, T., Lavigne, V., Saucier, C., Darriet, P., Teissedre, P.  
620 L., & Dubourdieu, D. (2009). Impact of oxygen dissolved at bottling and transmitted  
621 through closures on the composition and sensory properties of a Sauvignon Blanc wine  
622 during bottle storage. *J Agric Food Chem*, 57(21), 10261-10270.

623 Marchand, S., De Revel, G., & Bertrand, A. (2000). Approaches to wine aroma: Release of  
624 aroma compounds from reactions between cysteine and carbonyl compounds in wine.  
625 *Journal of Agricultural and Food Chemistry*, 48(10), 4890-4895.

626 Mattivi, F., Monetti, A., Vrhovsek, U., Tonon, D., & Andrés-Lacueva, C. (2000). High-  
627 performance liquid chromatographic determination of the riboflavin concentration in  
628 white wines for predicting their resistance to light. *J Chromatogr A*, 888(1-2), 121-127.

629 Maujean, A., Haye, M., Feuillat, M., Thomas, J. C., & Petit, D. (1978). Contribution à l'étude des  
630 « goûts de lumière » dans le vin de champagne. II. Influence de la lumière sur le  
631 potentiel d'oxydoreduction. Corrélation avec la teneur en thiols du vin. *OENO One*,  
632 12(4), 277-290.

633 Maujean, A., Haye, M., Feuillat, M. (1978). Contribution a l'étude des "Gouts de lumière" dans  
634 le vin de Champagne. II.- Influence de la lumière sur le potentiel d'oxydoreduction.  
635 Correlation avec la teneur en thiols du vin. *Connaissance Vigne et Vin*, 12(4), 277-290.

636 Maury, C., Clark, A. C., & Scollary, G. R. (2010). Determination of the impact of bottle colour  
637 and phenolic concentration on pigment development in white wine stored under  
638 external conditions. *Anal Chim Acta*, 660(1-2), 81-86.

639 Mestres, M., Busto, O., & Guasch, J. (2000). Analysis of organic sulfur compounds in wine  
640 aroma. *Journal of Chromatography A*, 881(1-2), 569-581.

641 Mestres, M., Martí, M. P., Busto, O., & Guasch, J. (1999). Simultaneous analysis of thiols,  
642 sulphides and disulphides in wine aroma by headspace solid-phase microextraction-  
643 gas chromatography. *Journal of Chromatography A*, 849(1), 293-297.

644 Mills, A. (2004). Lighting: The progress & promise of LEDs. *III-Vs Review*, 17(4), 39-41.

645 Nguyen Dang-Dung, N. L. a. A. K. P. (2012). Application of an Automated Headspace Solid  
646 Phase Micro-Extraction for the GC-MS Detection and Quantification of Reductive  
647 Sulfur Compounds in Wines. In D. B. Salih (Ed.), *Gas Chromatography in Plant Science,*  
648 *Wine Technology, Toxicology and Some Specific Applications*: InTech.

649 Perpète, P., Duthoit, O., De Maeyer, S., Imray, L., Lawton, A. I., Stavropoulos, K. E., Gitonga, V.  
650 W., Hewlins, M. J., & Dickinson, J. R. (2006). Methionine catabolism in *Saccharomyces*  
651 *cerevisiae*. *FEMS Yeast Res*, 6(1), 48-56.

652 Pripri-Nicolau, L., De Revel, G., Bertrand, A., & Maujean, A. (2000). Formation of flavor  
653 components by the reaction of amino acid and carbonyl compounds in mild conditions.  
654 *Journal of Agricultural and Food Chemistry*, 48(9), 3761-3766.

655 Roda, R., Martín, L., Mislata, A. M., Castaño, F. J., Puxeu, M., & Ferrer-Gallego, R. (2019).  
656 Effects of fertigation by elicitors enriched in amino acids from vegetal and animal  
657 origins on Syrah plant gas exchange and grape quality. *Food Research International*,  
658 125.

659 Santos, M. A., García-Ramírez, J. J., & Revuelta, J. L. (1995). Riboflavin biosynthesis in  
660 *Saccharomyces cerevisiae*. Cloning, characterization, and expression of the RIB5 gene  
661 encoding riboflavin synthase. *J Biol Chem*, 270(1), 437-444.

662 Siebert, T. E., Solomon, M. R., Pollnitz, A. P., & Jeffery, D. W. (2010). Selective determination of  
663 volatile sulfur compounds in wine by gas chromatography with sulfur  
664 chemiluminescence detection. *Journal of Agricultural and Food Chemistry*, 58(17),  
665 9454-9462.

666 Spikes, J. D. (1981). Photodegradation of Foods and Beverages. In K. C. Smith (Ed.),  
667 *Photochemical and Photobiological Reviews: Volume 6*, (pp. 39-85). Boston, MA:  
668 Springer US.

669 Ugliano, M. (2013). Oxygen contribution to wine aroma evolution during bottle aging. *Journal*  
670 *of Agricultural and Food Chemistry*, 61(26), 6125-6136.

671 Ugliano, M., Dieval, J. B., Siebert, T. E., Kwiatkowski, M., Aagaard, O., Vidal, S., & Waters, E. J.  
672 (2012). Oxygen consumption and development of volatile sulfur compounds during  
673 bottle aging of two Shiraz wines. influence of pre- and postbottling controlled oxygen  
674 exposure. *Journal of Agricultural and Food Chemistry*, 60(35), 8561-8570.

675 Ugliano, M., Kolouchova, R., & Henschke, P. A. (2011). Occurrence of hydrogen sulfide in wine  
676 and in fermentation: Influence of yeast strain and supplementation of yeast available  
677 nitrogen. *Journal of Industrial Microbiology and Biotechnology*, 38(3), 423-429.

678 Ullrich, S., Neef, S. K., & Schmarr, H. G. (2018). Headspace solid-phase microextraction and gas  
679 chromatographic analysis of low-molecular-weight sulfur volatiles with pulsed flame  
680 photometric detection and quantification by a stable isotope dilution assay. *Journal of*  
681 *Separation Science*, 41(4), 899-909.

682 Vela, R., Mazarrón, F.R., Fuentes-Pila, J., Baptista, F., Silva, L.L., García, J.L. (2017). Improved  
683 energy efficiency in wineries using data from audits. *Ciência Téc. Vitiv.*, 32(1), 62-71.



685 Table 1. Basic analysis of all wines (W1-W9), without exposure to light (time 0 hours) and after the last exposure time (240 hours) with each of the study  
686 lights (LA, LB and LC). The asterisk (\*) indicates significant differences in the samples with exposure to each of the lights (240 hours) compared to the  
687 samples in darkness (0 hours). Free sulfur (SO<sub>2</sub> F, mg/L), total sulfur (SO<sub>2</sub> T, mg/L), total acidity (TA, g/L expressed as tartaric acidity), alcoholic strength  
688 (AG, %vol), and volatile acidity (VA, g/L). Luminosity (*L*), *a* and *b* coordinates CIELAB. Samples (n = 2).

	SO <sub>2</sub> F	SO <sub>2</sub> T	pH	TA	AG	VA	420nm	<i>L</i>	<i>a</i>	<i>b</i>	<i>AE ab</i>
<b>W1 0h</b>	18 ± 0.0	95 ± 0.7	3.31 ± 0.0	6.3 ± 0.1	12.6 ± 0.0	0.4 ± 0.0	0.085 ± 0.0	99.1 ± 0.1	(-1.1) ± 0.0	6.6 ± 0.0	-
<b>W2 0h</b>	16 ± 0.7	96 ± 0.7	3.25 ± 0.0	6.3 ± 0.0	12.5 ± 0.0	0.4 ± 0.0	0.101 ± 0.0	98.2 ± 0.1	(-0.5) ± 0.1	6.7 ± 0.1	-
<b>W3 0h</b>	11 ± 0.0	82 ± 0.7	3.28 ± 0.0	6.1 ± 0.1	12.8 ± 0.0	0.5 ± 0.0	0.086 ± 0.0	98.7 ± 0.1	(-0.9) ± 0.0	5.7 ± 0.3	-
<b>W4 0h</b>	15 ± 0.7	89 ± 0.7	3.25 ± 0.0	6.5 ± 0.0	12.4 ± 0.0	0.4 ± 0.0	0.106 ± 0.0	98.2 ± 0.1	(-1.2) ± 0.0	7.5 ± 0.1	-
<b>W5 0h</b>	17 ± 0.0	91 ± 0.7	3.30 ± 0.0	5.9 ± 0.1	12.6 ± 0.0	0.4 ± 0.0	0.085 ± 0.0	98.7 ± 0.1	(-0.8) ± 0.0	6.1 ± 0.1	-
<b>W6 0h</b>	18 ± 0.0	79 ± 0.7	3.08 ± 0.0	6.1 ± 0.0	12.6 ± 0.0	0.4 ± 0.0	0.112 ± 0.0	97.0 ± 0.1	0.9 ± 0.0	6.9 ± 0.0	-
<b>W7 0h</b>	11 ± 0.7	60 ± 0.0	3.11 ± 0.0	5.5 ± 0.1	12.4 ± 0.0	0.3 ± 0.0	0.117 ± 0.0	96.7 ± 0.1	1.0 ± 0.0	7.9 ± 0.1	-
<b>W8 0h</b>	19 ± 1.4	81 ± 0.7	3.10 ± 0.0	5.9 ± 0.1	12.6 ± 0.0	0.3 ± 0.0	0.111 ± 0.0	96.9 ± 0.1	1.8 ± 0.0	7.5 ± 0.1	-
<b>W9 0h</b>	20 ± 1.4	87 ± 0.0	3.10 ± 0.0	6.0 ± 0.1	12.8 ± 0.0	0.3 ± 0.0	0.111 ± 0.0	97.0 ± 0.1	1.6 ± 0.0	7.4 ± 0.1	-
<b>W1 LA 240h</b>	18 ± 1.4	96 ± 4.2	3.31 ± 0.0	6.4 ± 0.0	12.6 ± 0.0	0.4 ± 0.0	0.082 ± 0.0	99.0 ± 0.2	(-1.1) ± 0.1	6.0 ± 0.3	0.6 ± 0.3
<b>W2 LA 240h</b>	20 ± 0.0	100 ± 0.7	3.25 ± 0.0	6.4 ± 0.1	12.5 ± 0.0	0.4 ± 0.0	0.090 ± 0.0	98.6 ± 0.1	(-1.1) ± 0.1 *	6.0 ± 0.1 *	1.0 ± 0.3
<b>W3 LA 240h</b>	12 ± 2.1	86 ± 2.1	3.29 ± 0.0	6.1 ± 0.0	12.8 ± 0.0	0.4 ± 0.0	0.079 ± 0.0 *	98.6 ± 0.2	(-1.0) ± 0.3	5.6 ± 0.1	0.2 ± 0.2
<b>W4 LA 240h</b>	14 ± 0.0 *	86 ± 1.4	3.25 ± 0.0	6.5 ± 0.1	12.4 ± 0.0	0.4 ± 0.0	0.103 ± 0.0	98.4 ± 0.1	(-1.1) ± 0.0	7.2 ± 0.0	0.4 ± 0.2
<b>W5 LA 240h</b>	18 ± 0.7	93 ± 0.7	3.29 ± 0.0	5.8 ± 0.0	12.6 ± 0.0	0.4 ± 0.0	0.084 ± 0.0	98.5 ± 0.1	(-1.0) ± 0.1	5.8 ± 0.0 *	0.4 ± 0.3
<b>W6 LA 240h</b>	18 ± 0.0	77 ± 1.4	3.08 ± 0.0	6.1 ± 0.0	12.6 ± 0.0	0.4 ± 0.0	0.103 ± 0.0	97.6 ± 0.2	0.9 ± 0.1	6.6 ± 0.4	0.7 ± 0.2
<b>W7 LA 240h</b>	11 ± 0.0	60 ± 0.7	3.09 ± 0.0	5.5 ± 0.0	12.4 ± 0.0	0.3 ± 0.0	0.106 ± 0.0	97.0 ± 0.0	0.9 ± 0.0	7.2 ± 0.0	0.8 ± 0.4
<b>W8 LA 240h</b>	21 ± 0.7	82 ± 1.4	3.10 ± 0.0	5.9 ± 0.0	12.6 ± 0.0	0.3 ± 0.0	0.106 ± 0.0	96.9 ± 0.1	2.0 ± 0.1	6.6 ± 0.1 *	0.9 ± 0.4
<b>W9 LA 240h</b>	20 ± 0.7	82 ± 1.4	3.09 ± 0.0	6.0 ± 0.0	12.8 ± 0.0	0.3 ± 0.0	0.103 ± 0.0 *	97.0 ± 0.1	1.9 ± 0.0	6.5 ± 0.2 *	0.9 ± 0.3
<b>W1 LB 240h</b>	18 ± 0.0	94 ± 0.7	3.30 ± 0.0	6.4 ± 0.1	12.6 ± 0.0	0.4 ± 0.0	0.083 ± 0.0	99.2 ± 0.0	(-1.0) ± 0.0	6.2 ± 0.0	0.2 ± 0.0
<b>W2 LB 240h</b>	17 ± 3.5	95 ± 7.1	3.28 ± 0.0	6.3 ± 0.1	12.6 ± 0.0	0.4 ± 0.0	0.092 ± 0.0	98.6 ± 0.1	(-0.9) ± 0.1	6.2 ± 0.1	0.3 ± 0.2
<b>W3 LB 240h</b>	12 ± 2.1	87 ± 2.1	3.29 ± 0.0	6.1 ± 0.0	12.7 ± 0.0	0.4 ± 0.0	0.082 ± 0.0	98.8 ± 0.0	(-0.9) ± 0.2	5.8 ± 0.1	0.3 ± 0.1
<b>W4 LB 240h</b>	14 ± 0.0 *	88 ± 1.4	3.25 ± 0.0	6.5 ± 0.0	12.4 ± 0.0	0.4 ± 0.0	0.106 ± 0.0	98.3 ± 0.1	(-0.9) ± 0.3	7.4 ± 0.1	0.2 ± 0.2

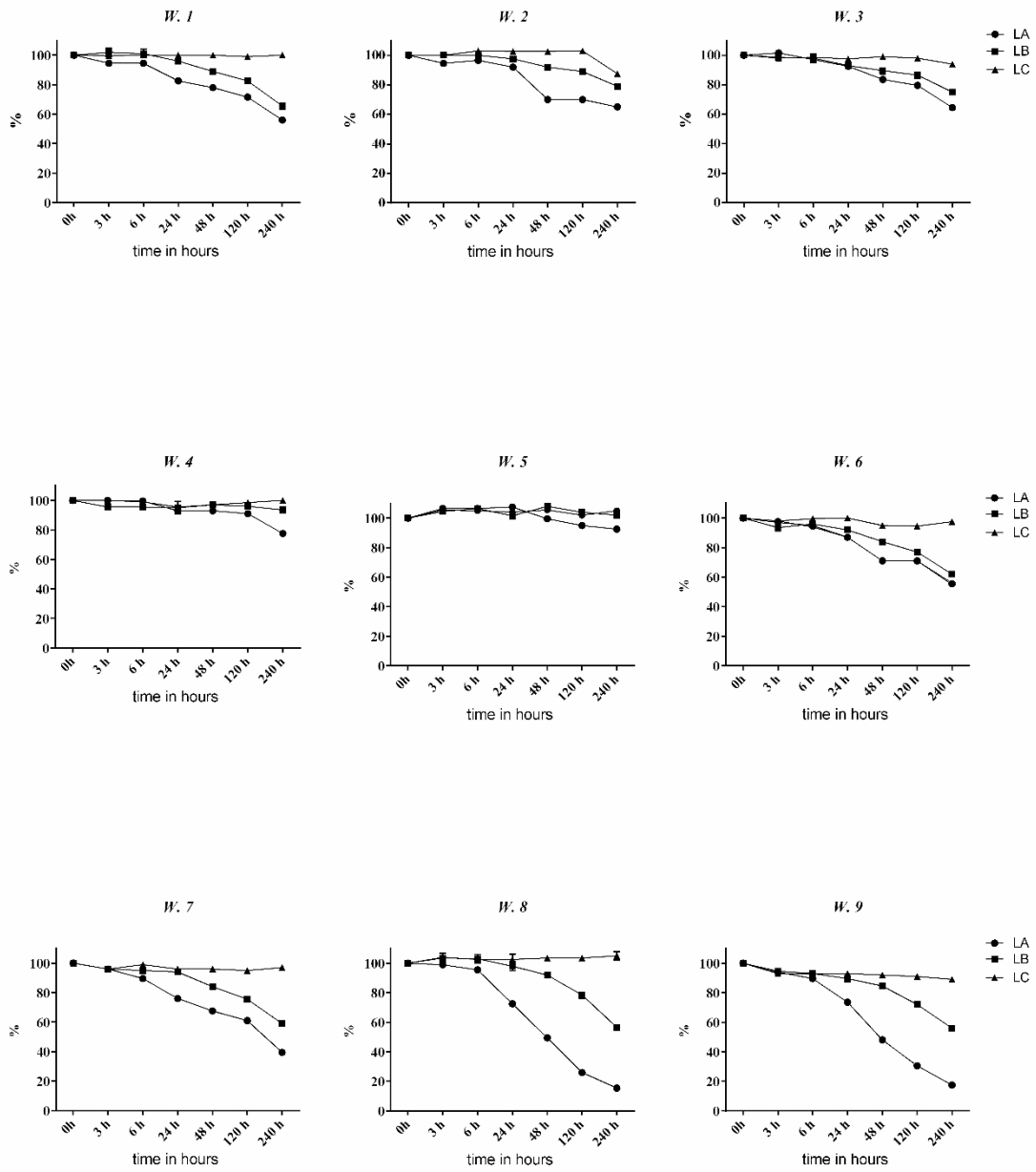
<b>W5 LB 240h</b>	15 ± 0.0 *	97 ± 1.4	3.30 ± 0.0	5.8 ± 0.0	12.3 ± 0.0	0.4 ± 0.0	0.088 ± 0.0	98.1 ± 0.1	(-1.2) ± 0.2	5.6 ± 0.0 *	0.4 ± 0.1
<b>W6 LB 240h</b>	19 ± 0.0	79 ± 0.7	3.08 ± 0.0	6.1 ± 0.0	12.6 ± 0.0	0.4 ± 0.0	0.097 ± 0.0	97.7 ± 0.1 *	0.7 ± 0.1	6.4 ± 0.1	0.4 ± 0.1
<b>W7 LB 240h</b>	11 ± 0.7	60 ± 2.1	3.10 ± 0.0	5.5 ± 0.0	12.4 ± 0.0	0.3 ± 0.0	0.117 ± 0.0	96.4 ± 0.4	1.0 ± 0.0	7.6 ± 0.2	0.7 ± 0.1
<b>W8 LB 240h</b>	21 ± 1.4	80 ± 0.7	3.09 ± 0.0	5.9 ± 0.0	12.6 ± 0.0	0.3 ± 0.0	0.113 ± 0.0	96.7 ± 0.1	1.8 ± 0.1	7.2 ± 0.1	0.7 ± 0.1
<b>W9 LB 240h</b>	14 ± 0.7 *	79 ± 0.0*	3.10 ± 0.0	6.0 ± 0.0	12.7 ± 0.2	0.3 ± 0.0	0.112 ± 0.0	96.7 ± 0.1	2.1 ± 0.2 *	7.2 ± 0.1	0.8 ± 0.2
<b>W1 LC 240h</b>	19 ± 0.0	98 ± 0.7	3.30 ± 0.0	6.4 ± 0.0	12.6 ± 0.0	0.4 ± 0.0	0.086 ± 0.0	99.0 ± 0.1	(-1.1) ± 0.1	6.4 ± 0.0	0.4 ± 0.1
<b>W2 LC 240h</b>	16 ± 0.7	98 ± 2.1	3.25 ± 0.0	6.3 ± 0.0	12.5 ± 0.0	0.4 ± 0.0	0.096 ± 0.0	98.4 ± 0.1	(-1.0) ± 0.2 *	6.4 ± 0.2	0.3 ± 0.2
<b>W3 LC 240h</b>	13 ± 1.4	85 ± 3.5	3.29 ± 0.0	6.1 ± 0.0	12.7 ± 0.0	0.4 ± 0.0	0.083 ± 0.0	98.7 ± 0.1	(-0.8) ± 0.0	5.8 ± 0.1	0.2 ± 0.2
<b>W4 LC 240h</b>	14 ± 0.0*	91 ± 1.4	3.24 ± 0.0	6.5 ± 0.0	12.4 ± 0.0	0.4 ± 0.0	0.103 ± 0.0	98.4 ± 0.1	(-1.0) ± 0.2	7.3 ± 0.1	0.2 ± 0.2
<b>W5 LC 240h</b>	16 ± 0.0	90 ± 2.8	3.30 ± 0.0	5.9 ± 0.1	12.6 ± 0.0	0.4 ± 0.0	0.086 ± 0.0	98.4 ± 0.1	(-1.0) ± 0.1	5.8 ± 0.0 *	0.4 ± 0.5
<b>W6 LC 240h</b>	18 ± 0.7	82 ± 0.7	3.08 ± 0.0	6.1 ± 0.1	12.6 ± 0.0	0.3 ± 0.0	0.102 ± 0.0	97.5 ± 0.1	0.6 ± 0.2	6.6 ± 0.1	0.3 ± 0.0
<b>W7 LC 240h</b>	12 ± 0.7	61 ± 2.1	3.09 ± 0.0	5.5 ± 0.0	12.4 ± 0.0	0.3 ± 0.0	0.115 ± 0.0	96.7 ± 0.1	0.9 ± 0.2	8.1 ± 0.3	0.6 ± 0.2
<b>W8 LC 240h</b>	18 ± 0.0	82 ± 0.0	3.10 ± 0.0	5.9 ± 0.0	12.6 ± 0.0	0.3 ± 0.0	0.113 ± 0.0	96.9 ± 0.1	1.6 ± 0.2	7.6 ± 0.1	0.4 ± 0.3
<b>W9 LC 240h</b>	20 ± 2.1	86 ± 2.1	3.09 ± 0.0	6.0 ± 0.0	12.8 ± 0.0	0.3 ± 0.0	0.113 ± 0.0	96.8 ± 0.1	1.7 ± 0.1	7.4 ± 0.1	0.5 ± 0.1

689

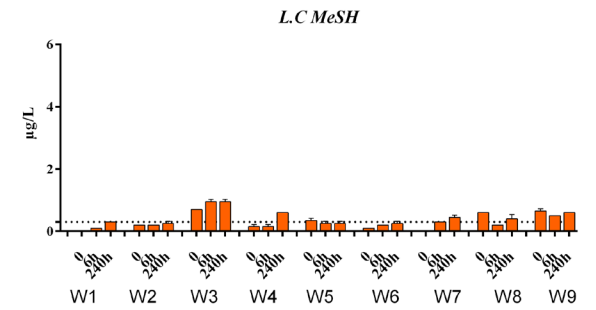
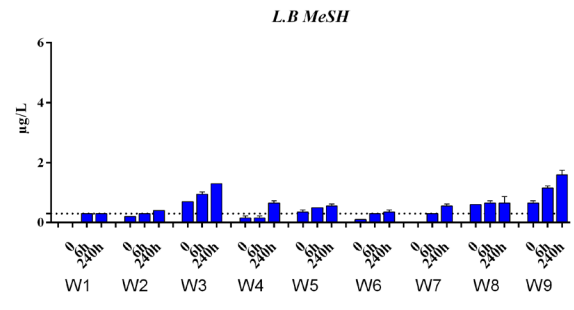
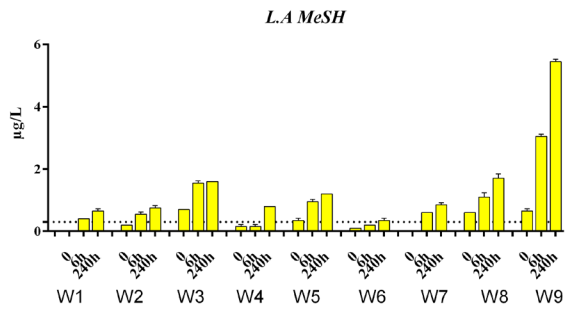
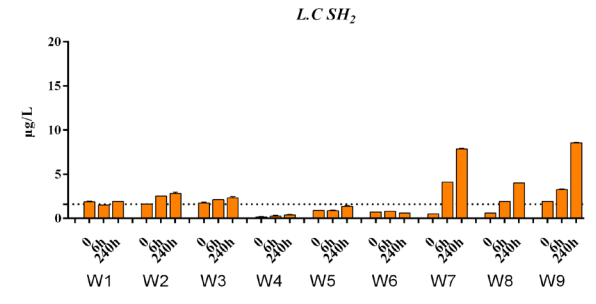
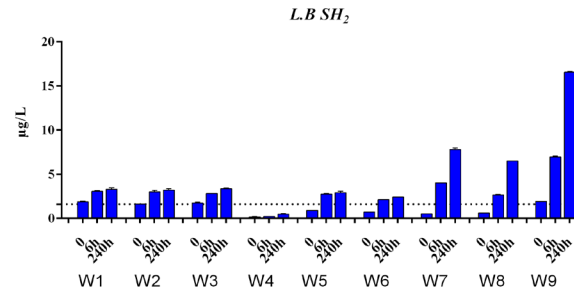
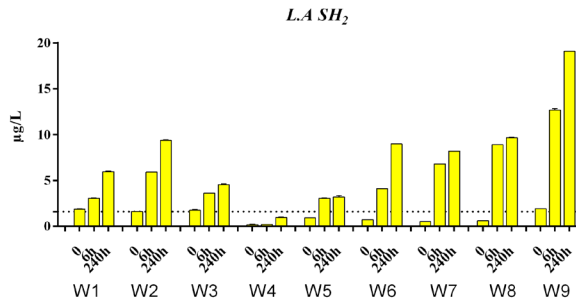
690

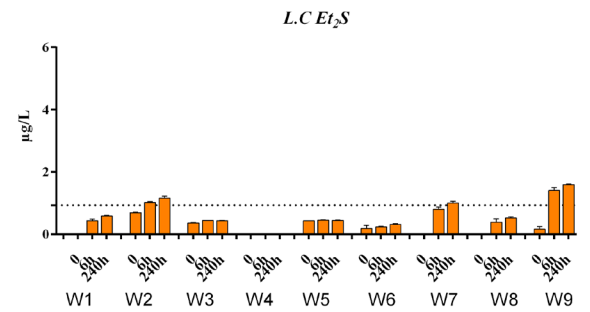
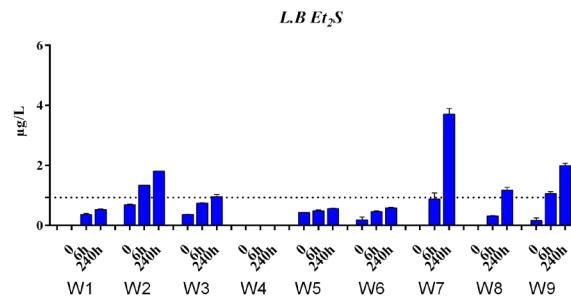
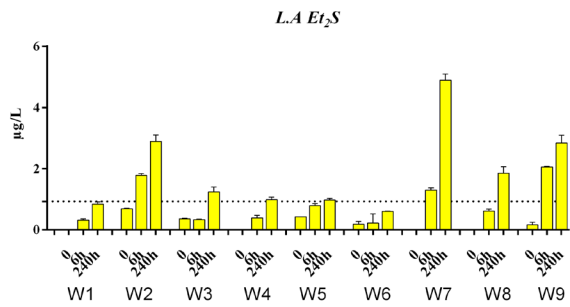
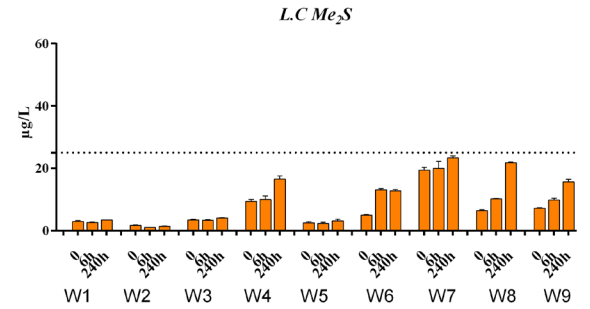
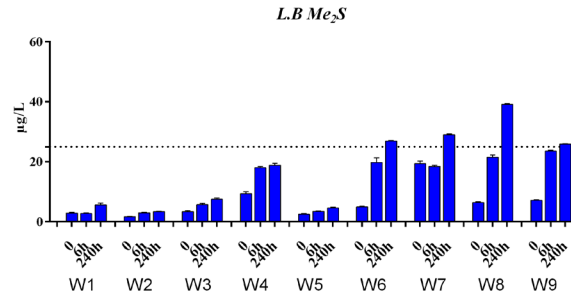
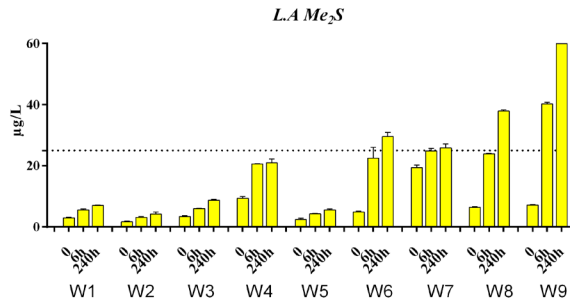
691

692









703 Figura 5.

